



PHARMACOLOGICAL ACTIVITIES OF CATHARANTHUS ROSEUS: A PERSPECTIVE REVIEW

GAJALAKSHMI S, VIJAYALAKSHMI S AND DEVI RAJESWARI V*

School of Biosciences and Technology, VIT University, Vellore-14, India

ABSTRACT

India possess a rich biodiversity of the medicinal plants that were still not explored completely. The need for the novel pharmaceutical products out from the plant has attained a great interest in the present research world due to the cost and the higher side effects that are associated with the chemically manufactured drugs. *Catharanthus roseus*, which is a potent medicinal plant many of the pharmacological actions such as antimicrobial, antioxidant, anthelmintic, antifeedant, antisterility, antidiarrheal, antidiabetic effect etc. That is used to treat many of the fatal diseases. Alkaloids were the major phytochemical constituent of the above medicinal plant and have different types possessing various medicinal uses. The pharmacology of the plant was found to be associated mostly especially with the alkaloids that occupies almost most of the parts of the plant. In this review, an attempt has been made to summarize the pharmacological effect of the above plant against various disease starting from the antimicrobial to that of the antineoplastic diseases in a precise way to help the scientist and learners to understand the basis medicinal value of the plant.

KEYWORDS: *Catharanthus roseus*, Apocynaceae, Monoterpenoid, Catharanthine, Antineoplastic.



DEVI RAJESWARI V

School of Biosciences and Technology, VIT University, Vellore-14, India

*Corresponding author

INTRODUCTION

Catharanthus roseus (L.) which is an important medicinal plant of the family Apocynaceae is used to treat many of the fatal diseases. *C. roseus* also possess good antioxidant potential. There are about two common cultivars of *C. roseus* which is named on the basis of their flower color that is the pink flowered 'Rosea' and the white flowered 'Alba'¹. *C. roseus* is extensively cultivated in northern India in order to meet their commercial and the ever increasing demand in the indigenous systems of the medicine also their need to the pharmaceutical industry. However, certain factors like the soil salinity hampers the cultivation of this plant very severely^{1, 2}. *Catharanthus roseus* which is proudly known as the Madagascar periwinkle is found to be a species of *Catharanthus* native and also endemic to Madagascar. The synonyms of the plant name include *Vinca rosea*, *Ammocallis rosea* and *Lochnera rosea*. Other English names occasionally used for the plant include Cape Periwinkle, Rose Periwinkle, Rosy Periwinkle and "Old maid"³. A traditionally used medicinal plant *Catharanthus roseus* is an erected procumbent herb or under the shrub containing latex. It grows widely to about 1m tall at the subtropical area. The leaves are found to be of oval to oblong, 2.5 – 9.0cm long and 1.0- 3.5 cm broad, glossy green, hairless, with a pale midrib and a short petiole of about 1.0- 1.8 cm long and they are arranged in the opposite pairs. The flowers looks white to dark pink with a darker red centre and a basal tube of 2.5 – 3.0 cm long with a corolla of about 2.0 – 5.0 cm diameter with five petals like lobes. The fruits are found to be a pair of follicles of about 2.0- 4.0 cm long and 3 mm broad⁴. This plant is found to be rich in their pharmacological actions that includes antibacterial, antifungal, antioxidant, anticancer and antiviral activates⁵.

PLANT DISTRIBUTION

In the wild, it is found to be an endangered plant and the main cause of their decline is the habitat destruction by the slash and burn agriculture. However, it is also widely cultivated

and naturalized in the subtropical and tropical areas of the world³. It is found to be an evergreen sub-shrub or the herbaceous plant that grows to about 1 m tall with white to dark pink flowers⁴.

PHYTOCHEMICAL COMPONENTS AND THEIR MEDICINAL USES

Alkaloids that are isolated from *C. roseus* are found to be hypotensive, sedative and possess tranquilising and anti cancerous properties. Traditionally, the plant has been used for relieving muscle pain, depression of the central nervous system and wasps stings. It is used in the cases of nose bleed, bleeding gums, mouth ulcers and sore throats. It has also been used internally for the treatment of the loss of memory, hypertension, cystitis, gastritis, enteritis, diarrhoea and the raised blood sugar levels⁶. Its application ranges widely from the prevention of cancer, cancer treatment, anti-diabetic, stomachic etc⁷. *Catharanthus roseus* was the highly exploited and studied medicinal plants as it was found to produce more than 100 monoterpenoid indole alkaloids (MIAs) that includes the two major commercially important cytotoxic dimeric alkaloids that are used in the cancer chemotherapy⁸. *C. roseus* was also found to be a good source of the non-enzymatic and enzymatic antioxidants^{9,10}. From the Traditional period itself, the plant has been used to cure diabetes and high blood pressure as it was believed to promote the insulin production or to increase the body's usage of the sugars from the food in case of diabetes¹¹. During the period of 1950s, *C. roseus* was found to possess a large number of chemicals in the alkaloid class. Alkaloids are the bitter-tasting plant compounds that contains mostly of nitrogen many of them was found to possess pain relieving or the anticancer properties. Especially two major alkaloids in *C. roseus* such as vinblastine and vincristine was developed into the prescriptions for the anticancer drugs¹². These injectable drugs and its derivatives such as vinflunine acts in several pathways and was found to interfere with the division of the cancer

cells. Recently, it was also found that certain chemicals in *C. roseus* involves in the prevention of the growth of the new blood vessels that is supporting the tumour growth¹³. Vincristine was found to be produced by *Fusarium oxysporum* which is an endophyte of this host¹⁴, while another group has isolated the vinblastine from an endophytic fungus¹⁵. *Catharanthus roseus* L. is found to be an important source of the indole alkaloids that are present in all plant parts. The plant has been used for the treatment of diabetes, fever, malaria, throat infections, chest complaints, regulation of menstrual cycles and as a euphoriant¹⁶. The physiologically important antineoplastic alkaloids such as vincristine and vinblastine are present in the leaves and the antihypertensive alkaloids are found in the roots such as ajmalicine, serpentine, and reserpine¹⁷. Vincristine and vinblastine alkaloids are found to be useful in the treatment of various types of lymphoma and leukemia^{18,19}. These *Catharanthus* alkaloids are found to be used for the treatment of both malignant and non-malignant diseases and in the platelet and platelet associated disorders¹⁶.

A US government screening program has discovered incidentally that the *Catharanthus* extracts were antineoplastic in vitro that leads ultimately to the licensing of the alkaloids such as vinblastine and vincristine, as well as some of the synthetic analogs today, as the highly toxic chemotherapy drugs. The absolute levels of vinblastine and vincristine are considered to be far too low in order to explain the activity of crude extracts of *Catharanthus*^{20, 21}. Other pharmacological uses of *C. roseus* include wound healing, analgesic, vasodilatory and hypoglycaemic²². Hence, *Catharanthus roseus* (L.) G. Don is found to be the most extensively investigated medicinal plant that is known mainly for its pharmacologically important alkaloids²³. present, the *Catharanthus* alkaloids was found to comprise a group of about 130 terpenoid indole alkaloids (TIAs)²⁴. No other single plant species was reported to produce such a wide array of the complex alkaloids²⁵. Wide differences has been noted in the compositions of the alkaloids that are

isolated from the underground and aerial tissues of the plant^{26,27}. The roots of the plant was found to accumulate ajmalicine and serpentine, which are the important components of medicines that are used for controlling the high blood pressure and other types of the cardio-vascular maladies. However, the plant has been particularly known for its economically significant leaf-specific bisindole alkaloids, vinblastine and vincristine, which are found to be the potent antineoplastic agents^{28,29} and the indispensable constituents of the most cancer chemotherapies. Vindoline that is found specifically in the green parts of the plant and not in the roots or cell suspension cultures was found to be biosynthesized from the branch-point intermediate tabersonine³⁰. Catharanthine was reported to be present in the root tissue and in the aerial part of the plant³¹. Alkaloids that are present in the plants was found to be effective in leukaemia treatment, diabetes, hypertension and menorrhagia etc³². But, only little work has been done on the anti-insect properties of the root extract of *C. roseus* when compared with that of the aerial portion³³. Vindoline and catharanthine are found to be the major monomer alkaloids. Vinblastine and vincristine are the two well known anticancer drugs that are used in the treatment of acute leukemia and Hodgkin's disease³⁴. Low "dimeric" alkaloid contents in the plants has encouraged the intense research for the alternative production methods by involving the usage of cell cultures^{35,36}, metabolic engineering³⁷, semi-synthesis^{38,39}, or even the total chemical synthesis⁴⁰. Total synthesis was found to be difficult due to the structural complexity of the molecules and also the complicated reaction steps that involves the stereochemical constraints. Various semi-synthetic procedures was developed for these alkaloids on the basis of their chemical^{38,39} or enzymatic⁴¹ coupling of the commercially available catharanthine and vindoline. A photochemical one pot synthesis has been proposed as a means of the simpler and economically feasible semi-synthesis of the vinblastine and vincristine⁴².

PHARMACOLOGICAL ACTIVITIES

ANTIOXIDANT ENZYME ACTIVITIES

An experiment was carried out in order to determine the changes in the antioxidant enzyme activities with respect to different concentrations of sodium chloride (NaCl) in *alba* and *rosea* varieties of *Catharanthus roseus* (L.) G. Don. in pot culture at various stages of growth. Especially, Superoxide dismutase (SOD), peroxidase (POX), catalase (CAT) enzymes antioxidant potentials were analysed. The result revealed the fact that the SOD activity was found to be increased at the level of 50 mM NaCl, but was reduced at further higher treatment levels. There were no significant changes obtained on the POX activity at the range of 25 mM NaCl level but showed significant increases of this activity at the next, higher levels of NaCl. From the above experiment, it was proved that *C. roseus* is an ideal plant for the cultivation in salt affected areas and we can obtain plants with the higher antioxidant and medicinal values⁴³.

ANTHELMINTHIC ACTIVITY

Helminthes infections are the chronic illnesses affecting human beings and cattle. *Catharanthus roseus* was found to be used from the traditional period as an anthelmintic agent. The anthelmintic property of *Catharanthus roseus* has been evaluated by using *Pherithema posthuma* as an experimental model and with Piperazine citrate as the standard reference. The ethanolic extract of the concentration of 250 mg/ml was found to show the significant anthelmintic activity with death time of 46.33 min whereas the standard drug at 50 mg/ml was found to show the death time of 40.67 min. This investigation supported the ethnomedical claims of *Catharanthus roseus* as an anthelmintic plant⁴⁴.

ANTI HYPERGLYCEMIC EFFECT

The effect of the daily oral administration of *Catharanthus roseus* (CR) leaf dichloromethane: methanol (1:1) extracts (500 mg/ body weight) for 20 days was tested on the blood glucose and hepatic enzymes in the normal and Alloxan induced diabetic rats. The

extract showed a significant increase in the body weight and decrease in the blood glucose, urea, cholesterol levels of the test animals. The activity of the hepatic enzymes such as hexokinase was increased whereas glucose 6-phosphatase and fructose 1, 6-bisphosphatase were found to be decreased significantly⁴⁵.

ANTINEOPLASTIC AND ANTIDIABETIC EFFECT

Different percentage of the methanolic crude extracts of *Catharanthus* was found to show the significant anticancer activity against numerous cell types in the *in vitro* condition⁴⁶ and especially greatest activity was found against the multidrug resistant tumor types⁴⁷. Several animal studies have proved that the ethanolic extracts of the leaves and flowers of *Catharanthus* has lowered the blood glucose levels^{48,49}. The aqueous extract was found to lower the blood glucose of about 20% in diabetic rats when compared to that of the dichloromethane and methanol extracts which lowered the blood glucose level to 49-58%⁵⁰. The hypoglycemic effects has appeared due to the result of the increased glucose utilization in the liver⁵¹.

IN VIVO ANTIDIARRHEAL ACTIVITY

The *in vivo* antidiarrheal activity of *C. roseus* ethanolic leaf extract was tested in the Wistar rats with castor oil as a experimental diarrhea inducing agent in addition to the pretreatment of the extract. Loperamide and atropine sulphate was used as the standard drugs. The antidiarrheal effect of ethanolic extract of *C. roseus* showed the dose dependant inhibition of the castor oil induced diarrhea at the doses of 200 and 500 mg/kg⁵².

ANTIMICROBIAL ACTIVITY OF CATHARANTHUS ROSEUS

Catharanthus roseus is found to be an important medicinal plant for the production of the novel pharmaceuticals as most of the bacterial pathogens were developing resistance against many of the available anti microbial drugs. Plants have been proved to be a

significant natural resources for the effective chemotherapeutic agents and offers a broad spectrum of activity with the greater emphasis on the preventive action. This study aims to investigate some of the anti microbial properties of this plant. The antimicrobial activity of the leaf extracts of the plant was checked against the microorganisms like *Pseudomonas aeruginosa* NCIM 2036, *Salmonella typhimurium* NCIM 2501, *Staphylococcus aureus* NCIM 5021 and was found that the extract could be used as the prophylactic agent in the treatment of many of the diseases⁵³.

TRANSCRIPTOME ANALYSIS

In *Catharanthus roseus* (L.) G. Don each plant tissue was known to produce a distinct spectrum of terpenoid indole alkaloids. But, the invaluable antineoplastic bisindole alkaloids was found to be restricted to the aerial parts of the plant and not in its underground tissues. Hence, the identification of the structural and regulatory factors that are operating distinctly in the shoot/leaf of the plant would be of a common necessity for the modulation of bisindole alkaloid biosynthesis. The elucidation of the differential gene expression in the two main tissues (leaf and root) of the plant which is well known for their distinct terpenoid indole alkaloid profiles was done indirectly through the construction and characterization of the separate cDNA libraries and directly through a strategically designed suppression subtractive hybridization by using the leaf and root cDNA populations as the tester and driver, respectively. Out of the total of 155 ESTs subjected to the homology-based classification, direct approach has yielded an EST for *sgd* (strictosidine *b*-D-glucosidase) and 16 novel ESTs⁵⁴.

EFFECT OF PERIWINKLE ON LEPIDOPTEROUS INSECT

Five fractions that were obtained from the root of the common periwinkle was evaluated for their antifeedant activity, growth regulatory potential and its effect on the fecundity and fertility against various stages of a lepidopterous insect, *Spilarctia obliqua*. The chloroform and

ethyl acetate fractions was found to be highly effective as the antifeedants, growth inhibitors and was found to cause sterility in the treated male adults. Initial instars of the test insect was found to be quite susceptible to the chloroform fraction at the concentration of 5%, whereas the late instar arvae was found to be unaffected⁵⁵.

STUDY ON THE POLYAMINES AND THE CELL CYCLE OF CATHARANTHUS ROSEUS CELLS IN CULTURE

The effect of partial depletion of polyamines (PAs) which is induced by the treatment with inhibitors of the biosynthesis of PAs, on the cell distribution at each phase of the cell cycle in *Catharanthus roseus* (L.) G. Don. cells in the suspension cultures was studied by means of using flow cytometry. Inhibitors of arginine decarboxylase (ADC) and ornithine decarboxylase (ODC) was found to accumulate more cells in the G₁ phase whereas the inhibitor of spermidine (SPD) synthase was found to show no effect on the distribution of cells. Two peaks of the endogenous level of PAs, in particular, of PUT and SPD was observed during the cell cycle. Levels of PAs was found to increase markedly prior to the synthesis of DNA in the S phase and prior to the cytokinesis. Activities of ADC was found to be much higher than that of ODC throughout the cell cycle, but both ODC and ADC changed in concert with the changes in levels of PAs⁵⁶.

ANTIOXIDANT PROPERTIES

Free radicals are found to be the fundamental of any biochemical process and hence represents an essential part of the aerobic life and metabolism and could show a dual role in our body as both the deleterious and beneficial species. The antioxidant potential of the ethanolic extracts of the roots of the two varieties of *Catharanthus roseus* L. namely 'rosea' (pink flowers) and 'alba' (white flowers) was obtained by using different systems of assay such as Hydroxyl radical-scavenging activity, superoxide radical-scavenging activity, DPPH radical-scavenging activity and nitric oxide radical inhibition method. The results obtained proved that the ethanolic extracts of

the roots of Periwinkle varieties extracts has exhibited the satisfactory scavenging effect in all the radical scavenging assays in a concentration dependent manner but *Catharanthus rosea* was found to possess more antioxidant activity than that of *Catharanthus alba*⁵⁷.

EFFECT OF AN ANTIDIABETIC EXTRACT OF CATHARANTHUS ROSEUS ON ENZYMIC ACTIVITIES

Hypoglycemic activity was detected by using the dichloromethane:methanol extract (1:1) of the leaves and twigs of *Catharanthus roseus* medicinal plant in streptozotocin (STZ) induced diabetic rat model at the dose of 500 mg/kg that was given orally for 7 and 15 days. The extract showed 48.6 and 57.6% hypoglycemic activity and further treatment for 30 days has provided complete protection against STZ challenge (75 mg/kg/i.p.×1). Enzymic activities of glycogen synthase, glucose 6-phosphate-dehydrogenase, succinate dehydrogenase and malate dehydrogenase was found to be decreased in the liver of diabetic animals which would be significantly improved after treatment with extract at dose 500 mg/kg p.o. for 7 days. Results indicated the increased metabolism of glucose in treated rats with the increased levels of lipid peroxidation⁵⁸.

REFERENCES

1. Jaleel CA, Panneerselvam R, Variations in the antioxidative and indole alkaloid status in different parts of two varieties of *Catharanthus roseus*: An important folk herb. Chinese Journal of Pharmacology and Toxicology. 2007; 1(6): 487- 494.
2. Hasegawa PM, Bressan RA, Zhu JK, Bohnert HJ, Plant cellular and molecular responses to high salinity. Annual Review on Plant Physiology, 2000; 51: 463-499.
3. Lewis WH, Elvin Lewis MPH, Medicinal Botany Plants Affecting Man's Health. John Wiley & Sons, New York; 1977.
4. Frode TS, Medeiros YSA Animal models to test drugs with potential antidiabetic

CONCLUSION

Medicinal plants were the potent source of various novel pharmaceutical products that shows ect causing potent pharmacological effect on the human beings. Instead of using the side effects causing chemical drugs, the ancient medicine could be explored to identify the novel drug formulations that are more effective with lesser side effects and also cheaper cost. Though, many of the traditional drugs were used without understanding the basic mechanism, their effect could be proved further with the help of the present technology and tools. The active compound that is responsible for the pharmacological effect could be found very easily and also commercialized as a drug product itself with proper approval from the respective organizations. The above plant was investigated from the ancient time for their phytochemical components and their pharmacological effect. The plant contains enormous phytochemical constituents of various medicinal application. The plant also possesses various activities such as antimicrobial, antioxidant, anthelmintic, antifeedant, antisterility, antidiarrheal, antidiabetic activities, etc. Hence, more work could be done on the above plant to reveal the unknown mysteries which would help the need of the present pharmaceutical world.

5. Marcone A, Ragozzino E, Seemuller, Dodder transmission of alder yellows phytoplasma to the experimental host *Catharanthus roseus* (periwinkle) Forest Pathology. 1997; 27(6): 347-350.
6. Dessisa D preliminary economic evaluation of medicinal plants in Ethiopia: trade, volume and price In: Medhin Z, Abebe D, editors. Proceedings of the National Workshop on biodiversity Conservation and Sustainable use of Medicinal Plants in Ethiopia. Addis Ababa, Ethiopia: 28 th April-May 1 st, 2001; 176-188.

7. Friis I, Gilbert MG, Chenopodiaceae. In: Edwards Mesfin T, Sebsebe D, Hedberg I, editors. Flora of Ethiopia and Eritrea; Magnoliaceae to Flacourtiaceae. Vol. 2, Published by National Herbarium of Addis Ababa University and Uppsala University, Uppsala. Sweden: 2000; 277.
8. Magnotta M, Murata J, Chen J, De Luca V, Identification of a low vindoline accumulating cultivar of *Catharanthus roseus* (L.) G. Don. by alkaloid and enzymatic profiling. *Phytochemistry*. 2006; 67: 1758–1764
9. Jaleel CA, Gopi R, Alagulakshmanan GM, Panneerselvam R. Triadimefon induced changes in the antioxidant metabolism and ajmalicine production in *Catharanthus roseus* (L.) G. Don. *Plant Science*. 2006; 171: 271–276.
10. Jaleel CA, Panneerselvam R, Variations in the antioxidative and indole alkaloid status in different parts of two varieties of *Catharanthus roseus*, an important folk herb. *Chinese Journal of Pharmacology and Toxicology*. 2007; 21: 487–494.
11. Singh SN, Vats P, Suri S, Shyam R, Kumria MM, Ranganathan S, Sridharan K, Effect of an antidiabetic extract of *Catharanthus roseus* on enzymic activities in streptozotocin induced diabetic rats. *Journal of Ethnopharmacology*, 2001; 76: 269–277.
12. Duflos A, Kruczynski A, Barret JM, Novel aspects of natural and modified vinca alkaloids. *Current Medicinal Chemistry – Anti-Cancer Agents*, 2002; 2: 55–70.
13. Zhang LB, Gou LH, Zeng SV, Preliminary study on the isolation of endophytic fungus of *Catharanthus roseus* and its fermentation to produce product of therapeutic value. *Chinese Traditional Herbal Drugs*, 2000; 11: 805–807.
14. Tung CY, Yang DB, Gou M, A preliminary study on the condition of the culture and isolate of endophytic fungus producing vincristine. *Journal of Chuxiong Normal University*, 2002; 6: 39–41.
15. Guo B, Kunming LH, A middle vinblastine fungi isolated. *Journal of Yunnan University*, 1998; 20: 214–215.
16. The Wealth of India. Raw Materials (Revised Edition), Vol. 3, C.S. Ambasta (Editor in Chief), Publication and Information Directorate, CSIR, New Delhi; 1992.
17. Mishra P, Uniyal GC, Sharma S. Pattern of diversity for morphological and alkaloid yield related traits among the periwinkle *Catharanthus roseus* accessions collected from in and around Indian Subcontinent. *Genetic Research in Crop Evolution*. 2001; 48: 273–286.
18. Farnsworth NR, Svoboda GH, Blomster RN. Antiviral activity of selected *Catharanthus* alkaloids. *Journal of Pharmaceutical Science*, 1968; 57: 2174–2175.
19. Svoboda GH, Blake DA. The phytochemistry and pharmacology of *Catharanthus roseus* (L.) G. Don. Inc. In: Taylor, W.J., Farnsworth, N.R. (eds.): *The Catharanthus alkaloids*. Marcel Decker, New York; 1975: pp. 45–84.
20. El-Sayed A, Cordell GA, *Catharanthus* alkaloids. XXXIV. Catharanthamine, a new antitumor bisindole alkaloid from *Catharanthus roseus*. *Journal of Natural Products*, 1981; 44(3): 289–93
21. El-Sayed A, Handy GA, Cordell GA, *Catharanthus* alkaloids, XXXVIII. Confirming structural evidence and antineoplastic activity of the bisindole alkaloids leurosine-N'-oxide (pleurosine), roseadine and vindolicine from *Catharanthus roseus*, *Journal of Natural Products*, 1986; 46(4): 517–27
22. Nayak BS, Lexley MPP. *Catharanthus roseus* flower has wound healing activity in Sprague Dawley rats. *BMC Complementary Alternative Medicine*. 2006; 6:41.
23. Verpoorte R, Van der Heijden R, Moreno PRH. Biosynthesis of terpenoid indole alkaloids in *Catharanthus roseus* cells. In: Cordell GA, ed. *The alkaloids*, Vol. 49. San Diego, CA: Academic Press, 1997; 221–299.

24. Van der Heijden R, Jacobs DI, Snoeijer W, Hallard D, Verpoorte R. The Catharanthus alkaloids: pharmacognosy and biotechnology. *Current Medicinal Chemistry*, 2004; 11: 607–628.
25. Blasko G, Cordell GA. Isolation, structure elucidation, and biosynthesis of the bisindole alkaloids of *Catharanthus*. In: Brossi A, Suffness M, eds. *The alkaloids*, Vol. 37. San Diego, CA: Academic Press, 1997; 1–76.
26. Mishra P, Kumar S. Emergence of periwinkle *Catharanthus roseus* as a model system for molecular biology of alkaloids: phytochemistry, pharmacology, plant biology and in vivo and in vitro cultivation. *Journal of Medicinal and Aromatic Plant Sciences*, 2000; 22: 306–337.
27. Shukla AK, Molecular studies on biosynthesis of shoot alkaloids in *Catharanthus roseus* (L.) G. Don. PhD thesis, Department of Biochemistry, University of Lucknow, India. 2005.
28. Svoboda GH, Blake DA, The phytochemistry and pharmacology of *Catharanthus roseus* (L.) G. Don. In: Taylor WI, Farnsworth NR, eds. *The catharanthus alkaloids*. New York, NY: Marcel Dekker, 1975; 45–83.
29. Neuss N, The spectrum of biological activities of indole alkaloids. In: Phillipson JD, Zenk MH, eds. *Indole and biogenetically related alkaloids*. London: Academic Press, 1980; 293–313
30. De Luca V, Balsevich J, Tyler RT, Eilert U, Panchuk BD, Kurz WGW, Biosynthesis of indole alkaloids: developmental regulation of the biosynthetic pathway from tabersonine to vindoline in *Catharanthus roseus*. *Journal of Plant Physiology*, 1986; 125: 147–156.
31. Deus-Neumann B, Stockigt J, Zenk MH, Radioimmunoassay for the quantitative determination of catharanthine. *Planta Medica*, 1987; 53: 184–188.
32. Atal CK, Kapur, BM, Cultivation and Utilization of Medicinal and Aromatic Plants, Regional Research Laboratory, Jammu Tawai. 1977; p. 138.
33. Deshpande SG, Joseph M, Sharma RN, Insect growth and development inhibition properties of *Catharanthus roseus*. *International Journal of Tropical Agriculture*, 1988; 6 (34): 287-290.
34. Noble RL, The discovery of the vinca alkaloids-chemotherapeutic agents against cancer. *Biochemistry and Cell Biology*, 1990; 68: 1344-1351.
35. Verpoorte R, Contin A, Memelink, J, Biotechnology for the production of plant secondary metabolites. *Phytochemical Review*, 2002; 1: 13-25.
36. De Luca V, Cutler AJ, Subcellular localization of enzymes involved in indole alkaloid biosynthesis in *Catharanthus roseus*. *Plant Physiology*, 1987; 85: 1099-1102.
37. De Luca V, St. Pierre B, The cell and developmental biology of alkaloid biosynthesis. *Trends in Plant Science*, 2000; 5: 168-173.
38. Goodbody AE, Vukovic J, Production of alkaloid dimers using ferric ion. *PCT Pat. WO88/02002*, 1988.
39. Kutney JP, Choi LSL, Nakano J, Tsukamoto H, Boulet CA, McHugh M, Process of synthesis of vinblastine and vincristine. *U.S. Pat. 5047528*, 1991.
40. Kuehne ME, Matson PA, Bornmann WG, Enantioselective synthesis of Vinblastine, Leurosidine, Vincovaline and 20' -epi-Vincovaline. *Journal of Organic Chemistry*, 1991; 56: 513-528.
41. Moreno PRH, Van der Heijden R, Verpoorte R, Cell and tissue cultures of *Catharanthus roseus*: A literature survey II. Updating from 1998 to 1993. *Plant Cell Tissue and Organ Culture*. 1995; 42: 1-25.
42. Pennanen S, Huhtikangas A, Photochemical one pot synthesis of vinblastine and vincristine. *Photochemical and Photobiological*. 1990; 51: 515-518.
43. Cheruth Abdul Jaleel, Soil Salinity Regimes Alters Antioxidant Enzyme Activities in Two Varieties of *Catharanthus*

- roseus*. Botany Research International, 2009; 2 (2): 64-68, 009
44. Swati Agarwal, Simi Jacob, Nikkita Chettri, Saloni Bisoyi, Ayesha Tazeen¹, VedamurthyAB, Krishna V, Joy Hoskeri H, Evaluation of *In-vitro* Anthelmintic Activity of *Catharanthus roseus* Extract. International Journal of Pharmaceutical Sciences and Drug Research, 2011; 3(3): 211-213
 45. J ayanthi M, Sowbala¹ N, Rajalakshmi G, Kanagavalli U, Sivakumar V, Study Of Anti Hyperglycemic Effect Of *Catharanthus Roseus* In Alloxan Induced Diabetic Rats, International Journal of Pharmacy and Pharmaceutical Sciences, 2009; 4: 19-25.
 46. Ueda JY, Tezuka Y, Banskota AH, Antiproliferative activity of Vietnamese medicinal plants. Biological Pharmaceutical Bulletin, 2002; 25(6):753-60
 47. Wang S, Zheng Z, Weng Y, Angiogenesis and anti-angiogenesis activity of Chinese medicinal herbal extracts, Life Science, 2004; 74(20): 2467-78
 48. Ghosh RK, Gupta I, Effect of *Vinca rosea* and *Ficus racemososus* on hyperglycemia in rats. Indian Journal of Animal Health, 1982; 19:145-8
 49. Chattopadhyay RR, Sarkar SK, Ganguli S, Hypoglycemic and antihyperglycemic effect of leaves of *Vinca rosea* Linn. Indian Journal of Physiology and Pharmacology, 1991; 35: 145-51.
 50. Singh SN, Vats P, Suri S, Effect of an antidiabetic extract of *Catharanthus roseus* on enzymic activities in streptozotocin induced diabetic rats. Journal of Ethnopharmacology, 2001; 76: 269-77
 51. Chattopadhyay RR, A comparative evaluation of some blood sugar lowering agents of plant origin. Journal of Ethnopharmacology, 1991; 67:367-72
 52. Kyakulaga A, Hassan, Alinda T, Brenda, Vudriko Patrick, Ogwang E, Patrick, *In vivo* antidiarrheal activity of the ethanolic leaf extract of *Catharanthus roseus* Linn. (Apocyanaceae) in Wistar rats, African Journal of Pharmacy and Pharmacology, 2011; 5(15) 1797-1800.
 53. Prajakta J. Patil, Jai S. Ghosh, Antimicrobial Activity of *Catharanthus roseus* – A Detailed Study. British Journal of Pharmacology and Toxicology, 2010; 1(1): 40-44.
 54. Ashutosh K. Shukla, Ajit K. Shasany, Madan M. Gupta, Suman PS Khanuja, Transcriptome analysis in *Catharanthus roseus* leaves and roots for comparative terpenoid indole alkaloid profiles. Journal of Experimental Botany, 2006; 57(14): 3921-3932.
 55. Veena Prajapati, Tripathi AK, Jain DC, Sharma S, Khanuja SPS, Sensitivity of *Spilarctia obliqua* to Root Extracts of *Catharanthus roseus*. Phytotherapy Research, 1998; 12: 270-274.
 56. Hisae Maki, Satoshi Ando, Hiroaki Kodama, Atsushi Komamine, Polyamines and the Cell Cycle of *Catharanthus roseus*. Cells in Culture and Plant Physiology, 1991; 96: 1008-1 013
 57. Alba Bhutkar MA, Bhise SB, Comparative Studies on Antioxidant Properties of *Catharanthus Rosea* and *Catharanthus*. International Journal of Pharmaceutical Techniques, 2011; 3(3): 1551-1556.
 58. Som Nath Singh, Praveen Vats, Shoba Suri, Radhey Shyam, Kumria MML, Ranganathan S Sridharan K, Effect of an antidiabetic extract of *Catharanthus roseus* on enzymic activities in streptozotocin induced diabetic rats. Journal of Ethnopharmacology, 2001; 76: 269-277