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ISOLATION, CHARACTERIZATION AND SCREENING OF COMPETENT FUNGAL ISOLATES FROM TEXTILE SLUDGE FOR DECOLORIZATION OF SELECTED SYNTHETIC DYES

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ABSTRACT

Textile industry is predominant in employing dyes and chemicals, cause for environmental degradation and human maladies. Synthetic dyes have an adverse effect on the aquatic ecosystem and their toxic substance has to be removed from the effluent before their exoneration. This research paper aimed, to isolate and screen the competent fungi from textile sludge capable of decolorizing broad spectrum of dyes. Twelve fungal isolates from sludge were preliminary screened by using Czapek Dox agar with 100 mg/ml of dyes. Six fungal isolates have shown decolorization of all three synthetic dyes. Among fungal isolates, *Aspergillus flavus* showed maximum growth (7.33±1.30) and in plate assay, *Aspergillus flavus* showed maximum decolorization zone (6.20 ±1.58) in Reactive violet-2RL followed by Acidic Blue-G and Direct Green-B, *Aspergillus fumigatus* showed 5.33±0.42 and 4.73±0.23. This research work reports that fungi offer an easy, cost effective method and could be used in development of an effective biotreatment method for the removal of dyes from textile effluent.

KEYWORDS: Competent fungal strains, Sludge, Dye agar plate assay, Decolorization and Linear regression.



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INTRODUCTION

Textile industries are economic, sustainable and consume large volumes of water, and employ diverse classes of dyes and chemicals like organic and inorganic complexes¹. Among the industrial wastewaters, textile effluent is one of the most stringent to treat and comprised of intricate mixture of carbon-based pollutants including dye. Synthetic dyes are classified as heterocyclic, polymeric triphenylmethane based on the chemical structure of chromogenic groups². In india, synthetic dyes accounts for around 5% of the global production. They are increasingly been used in the textile and dyeing industries because of their ease and cost effectiveness in synthesis, firmness, high stability to light, temperature, detergent and microbial attack and variety in color compared with natural dyes3. Discharge of textile effluent containing synthetic dyes especially poly-aromatics and pose threat to health issues like respiratory (coughing, wheezing, itchy or blocked noses and sneezing) and skin sensitization (itching, watery and sore eyes)4-6. The textile industry accounts for the largest consumption of dyestuffs. Reactive, Vat and Azo dyes are mainly required for dyeing and printing of cotton fibers. Disperse dyes constitute the largest market with about 21% share followed by direct dyes and reactive dyes with 16% and 11% respectively. Such dyes are recalcitrant to biodegradation, thereby creating an aesthetic problem in receiving waters. Effluents from textile industries, treated before discharged into the environment. Many trials made to develop effective technique decolorize these effluents.

methodologies Treatment for dve containing wastewater are categorized into three types: physical, chemical and biological. Most of the physical methods, however, simply accumulate and concentrate dves and create sludge and the problem of disposal exists. Chemical oxidation with either peroxide or ozone can destroy dyestuffs but this approach is costly⁷. Owing to their lower cost and ecofriendliness, biological methods have proved various physico-chemical superior over

methods. Various microorganisms like bacteria, fungi, yeasts and algae have been used to remove dyes⁸⁻¹². General mechanisms for the removal of dyes by microorganisms biosorption, bioaccumulation and biodegradation. Biosorption involves the entrapment of dyes in the matrix of the biomass by processes that do not involve metabolic energy or transport, although such processes may also occur simultaneously where live biomass is used. The process of biosorption can occur in either living or dead biomass^{13, 14} whereas bioaccumulation is the accumulation of pollutants by actively growing cells through metabolism and temperature-independent and metabolism-dependent mechanisms. biodegradation, the dye is fragmented into its metabolites 15-18.

The objectives of this study were:

- a) To isolate fungi from textile effluent,
- b) To identify the fungal isolates by LCB staining,
- c) To investigate the role of fungal isolates for decolorization efficiencies of three chemically different textile dyes namely Reactive Violet-2RL, Acidic Blue-G, Direct Green-B by Dyeagar plate assay.

MATERIALS AND METHODS

(i) Source of sludge

Sludge sample was collected (Mid off - April 2010) from a textile-dyeing mill in perundurai, Erode district, Tamil Nadu, India, in aseptic airtight plastic containers and filtered to remove large suspended particles and stored at 4±1°C in refrigerator until use.

(ii) Isolation and enrichment of fungi

Fungal strains were isolated from textile sludge by dilution plate method (10⁻¹ - 10⁻¹⁵) and the plates were observed for the growth of fungi. The fungal strains were sub-cultured by point inoculation method on Czapek Dox agar plates. Pure fungal isolates were maintained on agar slants, stored in 4°C and served as stock cultures. Sub-cultures were consistently made every 10 days. The fungal isolates were selected for further screening.

(iii)Dyes

A broad spectrum of dyes namely Reactive Violet-2RL, Direct Green-B and Acidic Blue-G listed in Table 1 and their structure is depicted in Figure 1. Further, to achieve high

decolorization ability of the fungal isolates was screened and treated in the laboratory-scale dye-agar plate assay with structurally different dyes. The dyes were procured from dye suppliers, Coimbatore, and were of analytical grade and used further without purification. The dye stock solutions were, prepared accurately by weighed 1gm/l of dyes in double distilled water.

Table 1
Synthetic Dyes used in the Study

DYE [CAS No.]	SYNONYMS	MOLECULAR WEIGHT (KDa)	λ _{max} (nm)
Reactive Violet-2RL [12218-07-4]	Reactive Red 83; Drimarene Violet X-2RL.	969.45	568nm
Acidic Blue-G [6104-58-1]	CBB G-250; Cyanine G; Blue G 250; Acid Blue 90; Serva Blue G; Polar Blue G; Page Blue G90; Brilliant Blue.	854.02	549nm
Direct Green-B [5422-17-3]	Diamine Green; Renol Green B; Dianil Green B; Direct Green G; Direct Green B	812.7	614nm

Figure 1
Structure of the Dyes used in the Study

(a) Reactive Violet-2RL

(b) Acidic Blue-G

(c) Direct Green-B

(iv) Preliminary screening

The fungal isolates were screened on relevant solid media containing 100mg/ml and incubated for 3 - 7 days in dark. The fungal colonies, which exhibit zone of clearance was chosen for further study.

(v) Characterization of competent fungal isolates

Fungi were identified according to their morphological characters by Agharkar Research Institute, Pune, India.

Screening

(vi) Fungal growth assay

The textile sludge contains organic matter, chemical nutrients and relative amount of heavy In spite of metals and aromatic dves. decolorization. maximum fungal biomass production was monitored by fungal growth assay. The mycelial agar plugs (~2 mm) of oneweek-old culture grown on Czapek Dox Agar at 28°C was cut from the colony margin under sterile conditions and transferred onto the centre of the experimental plates for each replicate and un-inoculated plate as control. These plates were incubated at room temperature (28°C) in dark and observed for 7days. The experiments were performed in triplicate for each culture.

(vii) Dye-agar plate assay

Dye - agar plate assay was carried out to ensure the potency of decolorization for

competent fungal isolates on Czapek Dox Agar medium containing 100mg/ml of dyes. Plates were inoculated with ~2 mm of fungal disc (actively growing region) was transferred onto the centre of the experimental plates for each replicate under aseptic conditions and incubated at room temperature (~28°C) in dark and un-inoculated agar plates were served as control. The clear zone formed around the mycelial disc indicates decolorization; the size of the decolorized halo was measured for every 24hrs.

(viii) Statistical analysis

Fungal growth and dye - agar plate assay were performed in triplicate and the results are reported as mean ± SD (Standard Deviation) by using statistical analysis tool Graph Pad Prism Version 5.04 and association of fungi with decolorization was assessed by linear regression model.

RESULTS AND DISCUSSION

Twelve fungal strains (Figure 2) were isolated from textile sludge and primarily screened by dye-agar plate assay for the dye was chosen for the study. Some dyes inhibit fungal growth because of their chemical structure. Among twelve fungal strains, six fungal strains showed clear zone. Table 2 illustrates the efficiency of fungal isolates for decolorization of selected synthetic dyes.

Fungal Strains Isolated from Textile Sludge

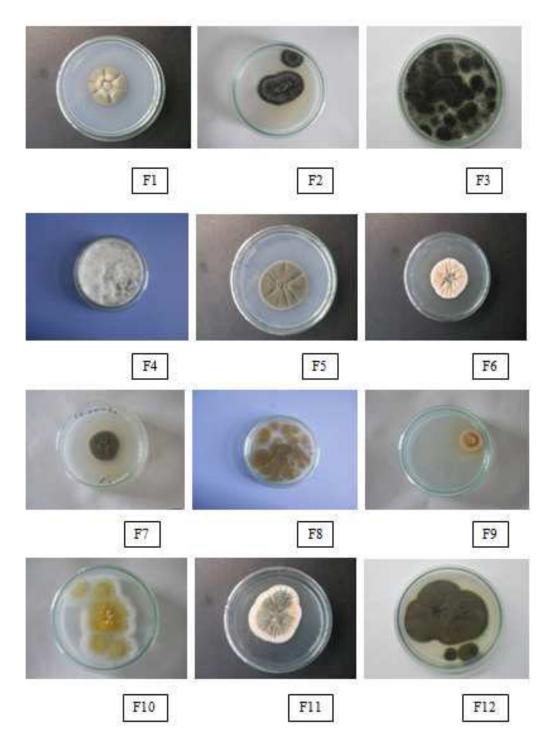


Figure 2
Twelve fungal strains were isolated from textile sludge and cultured in Czapek Dox Agar.

Table 2

Preliminary Screening: Decolorizing efficiency of fungal isolates

FUNGAL NO.	ISOLATE.	REACTIVE VIOLET-2RL	DIRECT GREEN-B	ACIDIC BLUE-G
F1		+	+	+
F2		+	+	+
F3		_	+	_
F4		+	+	+
F5		+	+	+
F6		+	_	_
F7		+	+	+
F8		+	+	+
F9	•	_	_	+
F10		_	_	_
F11		_	+	_
F12		_	_	_

(+) - Clear Zone observed, (-) - No Clear Zone.

Decolorization efficiency of six fungal isolates are identified as *Aspergillus flavus > Aspergillus fumigatus > Penicillium purpurogenum > Penicillium aurantiogriseum Dierckx > Lecanicillium lecanii > Cladosporium sphaerospermum penz.*, by Agharkar Research Institute, Pune, are listed in Table 3 and the fungal isolates are showed in Figure 3.

Table 3
Identification of Fungal Strains based on Microscopic
Observations and Cultural Characteristics

STRAIN NO.	ORGANISMS IDENTIFIED	ACCESSION NUMBER
F1	Aspergillus flavus	NFCCI -1957
F2	Aspergillus fumigatus	NFCCI - 1993
F4	Penicillium purpurogenum	NFCCI - 2823
F5	Penicillium aurantiogriesum	NFCCI - 942
F7	Lecanicillium lecanii	NFCCI - 2824
F8	Cladosporium sphaerospermum	NFCCI - 2730

Competent Fungal Isolates for Selected Synthetic Dye Decolorization

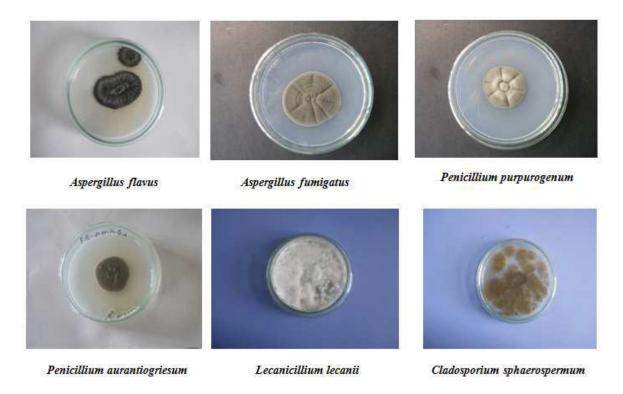
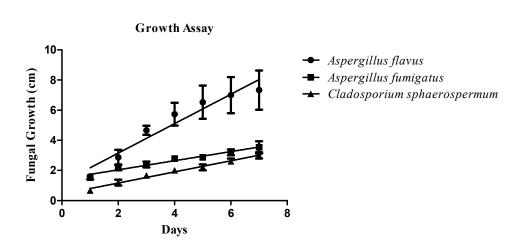


Figure 3
Six fungal isolates have Decolorizing potential for three selected synthetic dyes.

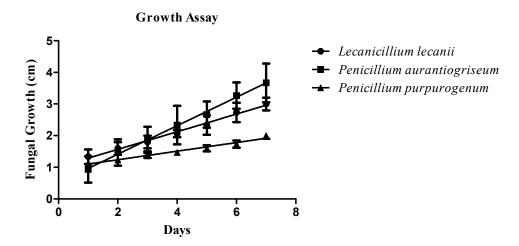
Selected fungal strains were tested for their growth and decolorization efficiency are shown in Graph 1. Maximum fungal growth was seen in *Aspergillus flavus* of diameter 7.30±1.30. Dye decolorization by fungi during growth on solid medium has been widely employed to identify

the degradation potential of dyes. In plate assay, maximum decolorization was showed by *Aspergillus flavus* in Reactive Violet-2RL (6.20±1.58) and *Aspergillus fumigatus* in Acidic Blue-G (5.33±0.42) and Direct Green-B (4.73±0.23).

Graph 1
Growth Assay for Six Isolates in Czapek Dox Agar



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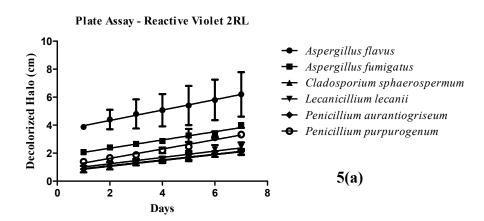


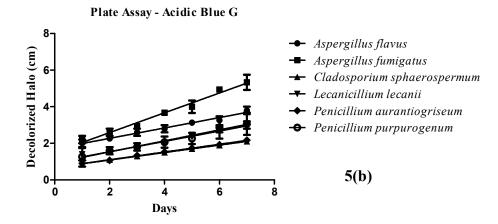
In order to verify the relationship between the fungal growth, and decolorization, a model of linear regression was evaluated for three dyes (Graph 2). The regression analysis reveals, the correlation coefficient (R²) for fungal growth, R² was found to be 0.8334, which is better than the acceptance criteria of 0.9177. The fungal growth and decolorized halo was compared with the fungal isolates by the measurement of the diameter. The linear relationship between the fungal growth and decolorization produces a

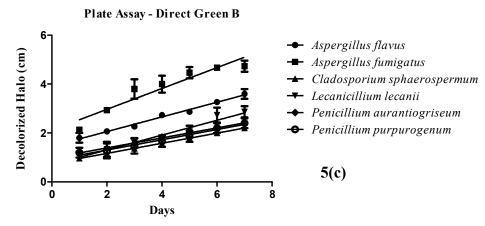
correlation coefficient for Reactive Violet-2RL (*Aspergillus flavus*), R² was found to be 0.3787, Acidic Blue-G and Direct Green-B (*Aspergillus fumigatus*), R² was found to be 0.9516 and 0.8595 and p - value is significant (<0.0001) and confirm the goodness of this approach. The development of plate assay holds a great promise and paves a way for new approach for treatment of synthetic dyes in textile industries by submerged fermentation.

Graph 2

Plate Assay for Decolorization of Synthetic Dyes







5(a): Reactive Violet-2RL, 5(b): Acidic Blue-G, 5(c): Direct Green-B

CONCLUSION

This research paper has demonstrated that isolated fungal strains had excellent potential for decolorization of selected synthetic dyes viz., Reactive Violet-2RL, Acidic Blue-G and Direct potential Green-B. Fungi have role persistent mineralizing many composites. Aspergillus flavus and Aspergillus fumigatus showed the greatest extent of dve decolorization in agar plates. This study demonstrate the decolorization of dyes with efficient strains like Penicillium purpurogenum, Penicillium aurantiogriseum Dierckx. Lecanicillium lecanii and Cladosporium sphaerospermum penz., revealed the decolorization of wide range of dyes. These results highlighted the potential strains for use in the treatment of dye-contaminated effluent.

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REFERENCES

- Jarosz-Wilkolazka A, Kochmanska-Rdest J, Malarczyk E, Wardas W and Leonowicz A, Fungi and their ability to decolorize azo and anthraquinonic dyes. Enzyme and Microbial Technology, 30: 566 – 572, (2002).
- 2. Hao OJ, Kim H and Chiang PC, Decolorization of wastewater. Critical Reviews in Environmental Science and Technology, 30: 449 505, (2000).
- 3. Couto SR, Dye removal by immobilised fungi. Biotechnology Advances, 27: 227 235, (2009).
- 4. Chung KT, The significance of azoreduction in the mutagenesis and carcinogenesis of azo dyes. Mutation Research, 114 (3): 269 281, (1983).
- 5. Achwal WB, Problems during analysis of textile as per eco-standards and the customer articles ordinance (Part I). Colourage, 44: 29 31, (1997).
- 6. Zhao X, Hardin IR and Hwang HM, Biodegradation of a model azo disperse dye by the white rot fungus *Pleurotus ostreatus*. International Biodeterioration and Biodegradation, 57: 1 6, (2006).
- 7. Robinson T, McMullan G, Marchant R and Nigam P, Remediation of dyes in textile effluent: a critical review on current treatment technologies with a proposed alternative. Bioresource Technology, 77(3): 247 255, (2001a).
- 8. Banat IM, Nigam P, Singh D and Marchant R, Microbial decolorization of textile-dye containing effluents: A review. Bioresource Technology, 58: 217 227, (1996).
- 9. Azmi W, Sani RK and Banerjee UC, Biodegradation of triphenylmethane dyes. Enzyme and Microbial Technology, 22: 185 191, (1998).
- Alhassani HA, Muhammad RA and Ashraf SS, Efficient Microbial Degradation of toluidine Blue dye by *Brevibacillus sp*. Dyes and Pigments, 755: 395 - 400, (2007).

- 11. Ncibi MC, Mahjoub B and Seffen M, Adsorptive removal of textile reactive dye using *Posidonia oceanica* (L.). International Journal of Environmental Science and Technology, 4 (4): 433 440, (2007).
- 12. Ghasemi F, Tabandeh F, Bambai B and Sambasiva Rao KRS, Decolorization of different azo dyes by *Phanerochaete chrysosporium* RP78 under optimal condition. International Journal for Environmental Science and Technology, 7 (3): 457 464, (2010).
- 13. Zhou W and Zimmermann W, Decolorization of industrial effluents containing reactive dyes by *Actinomycetes*. FEMS Microbiology Letters, 107 (2-3): 157 161, (1993).
- 14. Tobin JM, White C and Gadd GM, Metal accumulation by Fungi: Applications in Environmental Biotechnology. Journal of Industrial Microbiology, 13 (2): 126 130, (1994).
- 15. Sadettin S and Donmez G, Bioaccumulation of reactive dyes by thermophilic cyanobacteria. Process Biochemistry, 41 (4): 836 841, (2006).
- 16. Gupta VK and Rastogi A, Biosorption of lead from aqueous solutions by green algae *Spirogyra* species: kinetics and equilibrium studies. Journal of Hazardous Materials, 152 (1): 407 414, (2008).
- 17. Gupta VK and Rastogi A, Biosorption of hexavalent chromium by raw and acid-treated green alga *Oedogonium hatei* from aqueous solutions. Journal of Hazardous Materials, 163 (1): 396 402, (2009).
- Won SW, Choi SB and Yun YS, Interaction between protonated waste biomass of Corynebacterium glutamicum and anionic dye Reactive Red 4. Colloids and Surfaces A: Physicochemical and Engineering Aspects, 262 (1–3): 175 180, (2005).