



**QUALITATIVE PHYTOCHEMICAL SCREENING OF
GNIDIA GLAUCA (FRESEN) GILG. PLANT EXTRACT**

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ABSTRACT

The present study was undertaken to find out the phytochemicals present in plant *Gnidia glauca* belonging to the family Thymelaeaceae. Plant extract of dried leaves, bark and stem was prepared and qualitative phytochemical analysis was performed for the presence of Flavonoids, Quinone, Steroids, Tannin, Alkaloids, Phlobatanins, Anthraquinones, Glycosides, Terpenoides, Saponins using standard procedures. Phytochemicals from this plant, *Gnidia glauca* shows more potential in drug design and it requires further study.

KEYWORDS: *Gnidia glauca*, plant extract, phytochemicals, analysis.



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INTRODUCTION

The evaluation of all the drugs is based on phytochemical and pharmacological approaches which lead to drug discovery referred as natural product screening ^[1]. Any part of the plant may contain active components like bark, leaves, flowers, roots, fruits, seeds, etc. ^[2]. The effect of the plant materials result when the secondary products such as phytochemicals get combined. In recent time focus on plant research has increased all over the world. Plant have played a significant role in maintaining human health and improving the quality of human life for thousands of years and have served humans as well as valuable components of medicines, seasonings, beverages, cosmetics and dyes, ^[3]. Phytochemicals are the chemicals produces by plants, of these bioactive constituents of plants are steroids, terpenoids, carotenoids, flavanoids, alkaloids, tannins and glycosides. These compounds have various activities like antimicrobial or antibiotic, some have been reported to exhibit hemolytic and foaming activity, ^[4], Anti-inflammatory, ^[5], fungistatic, ^[6] and molluscidal, ^[7]. Thus the plants have played important role in drug development, ^[8]. *Gnidia glauca*, commonly called "Rametha", vernacular name *Datpadi*, is a semi- woody herb of Thymelaeaceae family, an inhabitant along margin of evergreen forests found in the Western Ghats- throughout India, Sri Lanka and also in Africa. Different parts (Stem and bark) of the herb are used in fishing techniques followed by some tribes in Maharashtra. The present investigation was undertaken to study qualitative chemical tests for establishing profile of *Gnidia glauca* with different solvents such as aqueous, chloroform, methanol and buffer extract.

MATERIALS AND METHODS

Plant materials

Fresh leaves stem and bark of *Gnidia glauca* were collected in bulk from Tamhini region of Western Ghats, Maharashtra, 18°27' latitude and 73°25' Longitude, during the month of

October- November in the year 2012. The plant was identified and authenticated from Botanical Survey of India, Western Regional Centre, 7, Koregaon Road, Pune and herbarium was kept at Department of Zoology, Modern College of Arts, Science and Commerce, Ganeshkhind, Pune as PK-1 specimen. Cleaned leaves, stem and bark were then dried under shade and were ground into a fine powder formed using mortar pestle and domestic mixer grinder machine. The fine powder of the plant leaves and aerial parts obtained was stored in sterile plastic bags at room temperature for further processing.

Chemicals and instruments

Chloroform, Methanol, Sodium Phosphate buffer and distilled water, hydrochloric acid, Wagner's reagent, Fehling's solution, ferric chloride reagent, sulphuric acid, benzene, ammonia solution, Conical flask, Test tubes, Bumper tubes, Stirrer, Burner, Water bath, pH meter, etc.

Preparation of extracts

Plant tissue homogenization

Plant tissue homogenization in solvent has been used. Dried fresh plant parts are grinded in a blender to fine particles, put in a certain quantity of solvent and shaken vigorously for 5 min and left for 24 h after which the extract is filtered and used for phytochemical screening ^[9].

Qualitative phytochemical screening

A number of phytochemical studies have demonstrated the presence of several classes of chemical compounds. Following different qualitative chemical tests were performed to investigate the chemical composition of *Gnidia glauca* plant extracts. Chemical test were carried out for alkaloids ^[10], glycosides ^[11] and quinines ^[12], flavonoids ^[13], tannins ^[14], saponins ^[15] and anthroquinones ^[16], terpenoids ^[8] and phlobatannins ^[8] and for Steroids ^[13].

Test for Alkaloids

5 ml of the extract was added to 2 ml of HCl. To this acidic medium, 1 ml of Wagners

reagent was added. A reddish precipitate brown produced immediately indicates the presence of alkaloids.

Test for Glycosides

To a small amount of extract, 1ml of Fehling's solution was added and heated, orange precipitate indicates the presence of glycosides.

Test for Flavonoids

4ml of extract solution was treated with 1.5 ml of 50% methanol solution. The solution was warmed and metal magnesium was added. To this solution, 5-6 drops of concentrated hydrochloric acid was added and red color was observed for flavonoids and orange color for flavones.

Test for Tannins

Ferric chloride reagent added to the filtrate. A blueblack, green, or blue-green precipitate was taken as evidence for the presence of tannins.

Test for Saponins

Frothing test was used to detect the presence of saponins. 0.5 gm of extract from each part was taken in test tube containing water. Solutions were warmed, resulted in to mass of bubbles indicates presence of saponins.

Test for Quinones

To a small amount of extract, concentration of sulphuric acid is added. Appearance of red color indicates the presence of quinones.

Test for Terpenoids

5 ml of each extract was mixed in 2 ml of chloroform. 3 ml of concentrated H₂SO₄ was then added to form a layer. A reddish brown precipitate colouration at the interface formed indicated the presence of terpenoids.

Test for Phlobatannins

Deposition of a red precipitate when an aqueous extract of each plant sample was boiled with 1 % aqueous hydrochloric acid was taken as evidence for the phlobatannins.

Test for Anthraquinones

Approximately 5g of each herbal extract was boiled with 10ml aqueous sulphuric acid and filtered while hot. The filtrate was shaken with 5 ml of benzene. The benzene layer was then separated and 10% ammonia solution was added to half of its volume. A pink, red or violet coloration in the ammonia phase (lower layer) indicated the presence of anthraquinone derivatives in the extracts.

Test for Steroid

Extract was treated with 2.5 ml of acetic anhydride and 2.5 ml of chloroform. Then concentrated solution of sulphuric acid was added slowly and red violet color was observed for terpenoid and green bluish color for steroids.

RESULTS

Phytochemical studies

Qualitative analysis of phytochemicals was carried out in four extracts of stem, leaves and bark of *Gnidia glauca*. Maximum extraction of phytochemicals were found to be present in Methanolic extract when compared with chloroform, aqueous and buffer extracts of *Gnidia glauca*. The phytochemical screening of all extracts of leaf showed that it contains flavonoids, steroids, tannins, terpenoids. The phytochemical screening of all extracts of stem showed that it contains steroids, quinones, alkaloids, terpenoids and the extract of bark showed that it contains steroids, terpenoids, quinones. The phytochemical screening of all extracts showed that the whole plant was rich in quinones, terpenoids, Tannins and Steroids. These compounds may used for several medicinal activities.

DISCUSSION

A number of phytochemical screening of plant material have been carried out^[18,19] and demonstrated the presence of several classes of chemical compounds. The various studies carried out on *Gnidia glauca* and recorded as piscicidal, arrow poison, insecticidal and molluscidal,^[20] Andelhave further reported that rotenone is chemical

compound present in *Gnidia glauca* for ichthyotoxic^[21], reported *Gnidia glauca* as magical plant that makes the painful tooth drop^[22]. It seems that various phytochemicals present in *Gnidia glauca* are showing the effects, as illustrated by various workers. Efficiency of *Gnidia glauca* inhibit amylase and

glucosidase activity^[23] and are showing positive results. Present study is supported by the above finding of various authors as the *Gnidia glauca* showing constituents as listed in the table. Further study is genuinely required to find out the importance of *Gnidia glauca* in drug design.

Table 1
Qualitative phytochemical analysis of extracts of Leaves, stem and bark of *Gnidia glauca* in different solvent system.

| No | Solvents Phytoconstituent | Leaf Extract | | | | Stem Extract | | | | Bark Extract | | | |
|----|------------------------------|--------------|----------|--------|---------|--------------|----------|--------|---------|--------------|----------|--------|---------|
| | | Chloroform | Methanol | Buffer | Aqueous | Chloroform | Methanol | Buffer | Aqueous | Chloroform | Methanol | Buffer | Aqueous |
| 1 | Flavonoids | +ve | +ve | +ve | +ve | -ve | +ve | +ve | -ve | -ve | +ve | -ve | -ve |
| 2 | Quinones | -ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve |
| 3 | Steroids | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve |
| 4 | Tannins | +ve | +ve | +ve | +ve | +ve | +ve | +ve | -ve | +ve | +ve | +ve | -ve |
| 5 | Alkaloids | +ve | +ve | -ve | -ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | -ve |
| 6 | Phlobatanins | -ve | -ve | -ve | -ve | +ve | +ve | -ve | -ve | +ve | +ve | -ve | -ve |
| 7 | Anthraquinones | -ve | +ve | +ve | +ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve |
| 8 | Glycosides | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve | +ve | +ve | -ve | -ve |
| 9 | Terpenoides | -ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve | +ve |
| 10 | Saponin | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve | -ve |

(-ve) Absent, (+ve) Present

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REFERENCES

1. Foye WO, Lemke TL, Williams DA, *Foye's Principles of Medicinal Chemistry*, 6th Ed Lippincott Williams and Wilkins. Philadelphia, 44-45, (2008).
2. Gordon MC, David JN. Natural product drug discovery in the next millennium. *Pharm Biol.* 39 Suppl 1:8-17, (2001).
3. Makhija I.K.*, Sharma I.P., Khamar D.; *Phytochemistry and Pharmacological properties of Ficus religiosa: an overview*; Scholars Research Library; Annals of Biological Research, 1 (4) : 171-180, (2010).
4. Feroz, M., R. Ahmad and S.T.A.K. Sindhu and A.M. Shahbaz, 1993. Antifungal activities of saponins from indigenous plant roots. *Pak. Vet. J.*, 13: 4-4, (1993).
5. Takagi, K., Hee P.E. and Histoshi K., Anti-inflammatory activities of hederagenin and crude saponin from *Sacchindus mukarassi*. *Chem. Bull.* 28(4): 1183 (Eng.) *Chem. Abst.* 93(21): 372384, (1980).
6. Zehavi, U.M.L. and R. Segel,. Fungistatic activity of saponin A from *Styrex officinalis* L. on plant pathogens. *J. Phytopathol.*, 116: 338-343. (1986).
7. Sati, O.P., Chandra C.D. and Usha R., Molluscidal saponins of xeromphis saponins. *Planta Medica*, 5: 981-983, (1987).
8. Edeoga HO, Okwa DE, Mbaebie BO. Phytochemical constituents of some Nigerian medicinal plants. *African Journal of Biotechnology*, 4(7): 685-688, (2005).

9. Das K, Tiwari RKS, Shrivastava DK. Techniques for evaluation of medicinal plant products as antimicrobial agent: Current methods and future trends. *Journal of Medicinal Plants Research*; 4(2): 104-111, (2010).
10. William C Evans, Trease and Evans Pharmacognosy, 16th Ed. Elsevier, 133-135,(2009).
11. Marco Brito-Arias , Synthesis and characterization of Glycosides; Springer, 304-313, (2006).
12. Priya D., Rajaram K., Suresh kumar P. Phytochemical studies and GC-MS analysis of *caralluma fimbriata* wall. *IJPRD*, VOL 3(10) (105- 110): (2011).
13. Siddiqui, A.A., and Ali , M. Practical Pharmaceutical chemistry. 1st ed. CBS publishers and distributors, New Delhi.pp.126-131, (1997).
14. Segelman, A.B., Farnsworth, N.R. and Quimby, M.D. False-negative saponins test results induced by the presence of tannins. *Lloydia*, 32: 52-58, (1969).
15. David S. Seigler, Plant Secondary Metabolism, Kluwer academic publishers,440-470, (2001).
16. Gokhale S. B., Kokate C. K. and A. P. Purohit., A textbook Pharmacognosy ,29th Ed. Nirali Prakashan, Pune, pp 192- 8-11, (2009).
17. Abba, D., Inabo, H. I., Yakubu, S. E. and Olonitola, O. S. Phytochemical analysis and antibacterial activity of some powdered herbal preparations marketed in Kaduna metropolis; *Science World Journal* Vol 4 (No 1) ISSN 1597-6343 (2009).
18. Manorenjitha M. S., Norita A. K., Norshisham S. and Asmauli M. Z., GC- Ms Analysis of Bioactive Components of *Ficus religiosa* (linn.) Stem., *Int. J. Pharm. Biosci.*, 4(2) (p) 99-103. (2013).
19. Tiwari Prashant, Bimlesh Kumar, Mandeep Kaur, Gurpreet Kaur, Harleen Kaur; Phytochemical screening and extraction: a review, *Internationale pharmaceutica sciencia* Vol. 1 | Issue 1 (2011).
20. Ajayi I.A, Ajibade O, Oderinde RA. Preliminary Phytochemical Analysis of some Plant Seeds. *Res. J. Chem. Sci.* 1(3): 58-62(2011).
21. Andel T. V., The diverse uses of fish-poison plants in Northwest Guyana ; *Economic Botony* 54(4) pp. 500-512. (2000).
22. Raghavendra Rao C V , A magic plant that makes your painful tooth drop *Times of India*, Mar 3, 2002. Page 3 March 3, (2002)
23. Ghosh, Sougata ; Ahire, Mehul ; Patil, Sumersing ; Jabgunde, Amit ; Dusane, Meenakshi Bhat ; Joshi, Bimba N. ; Pardesi, Karishma ; Jachak, Sanjay ; Dhavale, Dilip D. ; Chopade, Balu A. Antidiabetic activity of *Gnidia glauca* and *Dioscorea bulbifera*: potent amylase and glucosidase inhibitors, *Evidence-based Complementary and Alternative Medicine*, 2012 . 929051_1-929051_10 (2012).