

**MODULATORY EFFECT OF *ALOE VERA* LEAF EXTRACT AGAINST SULPHUR DIOXIDE INDUCED LIVER INJURY IN ALBINO RAT****MADHURI YADAV*, ASHA AGARWAL AND PREETI KUMARI***Department of Zoology, School of Life Sciences, Khandari Campus, Dr. B.R. Ambedkar University, Agra***ABSTRACT**

The present study was carried out to evaluate the antioxidative effect of *Aloe vera* (*A. vera*) aqueous leaf extract on hepatotoxicity induced by SO₂ inhalation in male albino rats. Hepatotoxicity was assessed by estimating the Serum Alkaline Phosphatase (ALP), Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), Gamma Glutamyl Transferase (GGT) and histopathological changes in liver of rats. Experimental rats were exposed to 80ppm hrd-1, SO₂ gas per day for 30 and 60 days. A significant increase in Serum ALP (P<0.001), AST (P<0.01), ALT (P<0.001), GGT (P<0.05) and histopathological changes include hepatocytes necrosis, degeneration of hepatocytes, centrilobular necrosis, ballooning degeneration and cellular debris have observed after SO₂ exposure in comparison to control rats. A marked reduction in liver injury is reported after exposure with oral administration of *A. vera* leaf extract (200mgkg⁻¹ body weight d⁻¹) in comparison to SO₂ exposed rats. Antioxidative and anti-inflammatory properties of *A. vera* shows potential to modulate the hepatocellular damage induced by SO₂ gas in rats.

KEY WORDS: Male albino rat, SO₂, *A. vera* aqueous leaf extract, Liver injury**Dr. MADHURI YADAV**Department of Zoology, School of Life Sciences, Khandari
Campus, Dr. B.R. Ambedkar University, Agra** Corresponding author*

INTRODUCTION

Sulphur dioxide (SO₂) is the most toxic air pollutant and detrimental to many organs¹. The major sources of SO₂ emission are combustion of fossil fuels, burning of coal in thermal power plants, petroleum refineries, fertilizers and paper and pulp industries etc. Sulphur dioxide inhalation causes production of reactive oxygen species (ROS) in the tissues. An imbalance between the production of ROS and biological system's ability to readily detoxify the reactive intermediate causes oxidative stress². The oxidative stress caused by SO₂ leads to damage or death of tissue, causing the leakage of enzymes in the affected tissue(s) into the blood stream³. Hepatotoxic effects of any toxic substance is mainly diagnosed by markers of liver enzymes such as ALP, ALT, AST, and GGT are employed in assessing liver injury⁴. *Aloe vera* (L.) belongs to the family Liliaceae commonly known as "Gheekunwar". It has therapeutic properties such as – anti-oxidative, anti-inflammatory effects, wound healing, anti-bacterial, anti-viral, anti-fungal, anti-diabetic and stimulation of hematopoiesis⁵. The aloesin derivatives of *Aloe vera* possess strong OPPH radical and superoxide anion scavenging activities^{6,7}. With this in view, present study is carried out to investigate the protective role of *A. vera* leaf extract on SO₂ induced serum enzymes and histopathological change in liver of albino rat.

MATERIALS AND METHODS

Animals

Thirty male wistar albino rats (2 months old, body weight 120-135g) were used in present study. The animals were maintained as per the norms of institutional Animal Ethics committee. Experimental rats were kept in polypropylene cages, and maintained at temperature (25±2°C), humidity (60±5%) and photoperiod of 12 hrs d⁻¹. The rats were acclimated for two weeks prior to experiment. During this period rats were fed on commercial food pellets (Golden feed, New Delhi) and the water *ad libitum*.

Generation Of SO₂

SO₂ gas (80ppm) was generated by controlled action of 5% sulphuric acid and 22% sodium sulphite solution in SO₂ generator⁸. To produce 1ppm of Sulphur dioxide/ m³ air, the strength of sodium sulphite in the vessel was maintained to 2.7 mgml⁻¹ and 5% sulphuric acid was used in the reaction.

Exposure of SO₂ gas

Rats were exposed in fumigation chamber (AP 07 model SFC 120), Standard Appliances, Varanasi.

Preparation of Aloe vera leaf extract

Fresh leaves of *A. vera* were collected from the Diwan Farm, Barara, Agra. The thick epidermis was selectively removed and inner colourless mucilaginous pulp was taken. The fresh pulp (1kg) of *A. vera* grinded in spice grinder and extracted with one litre of distilled water and kept with magnetic stirrer in cold room temperature for 48hours. The suspension was filtered by Whatman's No. 1 filter paper and filtrate was used for experimentation. The dose of *A. vera* leaf extract (200mgkg⁻¹ body weight d⁻¹) was administered in rats by gavaging. The dose was selected as guidelines as per Traditional Medicine System⁹.

Experimental design

The rats were randomly divided into six groups (C₁, C₂, E₁, E₂, E₃ and E₄) of five rats each- two control group (C₁&C₂) were exposed to ambient air and four experimental groups (E₁ & E₂ and E₃ & E₄). Experimental groups (E₁ & E₂) were exposed to 80ppm SO₂ gas and Experimental groups (E₃ & E₄) were exposed to 80ppm SO₂ with oral administration of *A. vera* (200mgkg⁻¹ body weight d⁻¹) for one hour per day for 30 and 60 days respectively. The rats of control and experimental groups were dissected carefully under light anaesthesia and blood samples (4-5ml) were collected from the ventricle of heart in sterilized plain centrifuge tubes for separation of serum. Blood samples were centrifuged at 2500rpm for 30min and serum was separated for the estimation of enzyme

activities. Liver tissues were taken to assess the histopathological study.

Biochemical study

Serum enzymatic activities (ALP, ALT, AST and GGT) were measured by the International Federation of Clinical Chemistry (IFCC) kit methods described by Bradley¹⁰; Tietz¹¹.

Histopathological study

The small pieces of liver tissue of each rat was fixed in 10% formalin after washing in saline. Dehydrated tissue through a series of ethanol solution and paraffin blocks were prepared and sections at 5 micron were cut and stained with haematoxyline and eosin¹². The stained sections were examined and photomicrographs were taken.

Statistical analysis

The results were expressed as Mean \pm S.Em. were signified by using Student's 't' test. Statistical calculation was carried out by using one way ANOVA with the help of computer statistical program KpKy plot (version- 3.0).

RESULTS

Results of present study show a significant increase ($P < 0.05$) in Serum ALP, AST, ALT and GGT activities in SO₂ exposed rats in comparison to control group, while a significant decrease ($P < 0.05$) in SO₂ exposed rats with *A. vera* in comparison to 80ppm SO₂ gas exposed rats after 30days (Table-1). A highly significant increase ($P < 0.01$) in Serum ALP, AST, ALT and GGT activities in SO₂ exposed rats in comparison to control group,

while a very highly significant decrease ($P < 0.001$) in Serum ALP, ALT activities, highly significant decrease ($P < 0.01$) in AST and significant decrease ($P < 0.05$) in GGT activity in SO₂ exposed rats with *A. vera* in comparison to 80ppm SO₂ gas exposed rats after 60days (Table-2). Histopathological changes were observed in liver tissue of experimental rats in comparison to control rats include moderate hepatocytes necrosis, degeneration of hepatocytes, centrilobular necrosis, ballooning degeneration and cellular debris (Fig. 2a and 2b) after 30 days exposure to 80ppm SO₂ gas, while after 60days exposure to 80ppm SO₂ gas, severity in hepatocyte necrosis and degeneration of hepatocytes have been observed in liver section. Ballooning degeneration, cellular debris and centrilobular necrosis are more pronounced in comparison to 30 days (Fig. 3a and 3b). Photomicrographs of liver section shows an improvement in hepatotoxicity after 30days exposure to SO₂ with administration of *A. vera* in rats. Mild hepatocyte necrosis and degeneration of hepatocytes have been observed at some places. Occasional debris, cellular boundaries are slightly disintegrate and cells appear in their normal shape and architecture. Binucleated cells of hepatocytes are also seen (Fig. 4a and 4b). Photomicrographs of liver section after 60days exposure to SO₂ with administration of *A. vera* shows modulation of hepatic injury in rats, which is evidenced by the appearance of central vein in their normal shape and architecture and cellular boundaries are clearly identify. Binucleated cells of hepatocytes are also seen (Fig. 5a and 5b).

Table-1
Protective effects of *A. vera* on Serum ALP, AST, ALT and GGT activities after 30 days exposure to SO₂ in albino rats

Groups	Exposure and pre-exposure supplementation	ALP (U/L)	AST (U/L)	ALT (U/L)	GGT (U/L)
		Mean \pm S.Em	Mean \pm S.Em	Mean \pm S.Em	Mean \pm S.Em
Control group-C ₁ (5)	Ambient air	160.6 \pm 2.18	112.6 \pm 2.66	48.2 \pm 3.02	15.18 \pm 0.87
Experimental group-E ₁ (5)	80ppm SO ₂ gas	181.6 \pm 6.86*	123.2 \pm 2.89*	58.6 \pm 2.34*	19.86 \pm 1.38*
Experimental group-E ₃ (5)	80ppm SO ₂ gas + <i>A. vera</i>	159.6 \pm 2.56 ^{●S}	113.2 \pm 1.85 ^{●S}	47.2 \pm 2.63 ^{●S}	15.32 \pm 0.87 ^{●S}

5 = Number of rat

S.Em. = Standard Error of Mean

Significantly different from control

● = ($p < 0.05$)

* = ($p < 0.01$)

Significantly different from SO₂ exposed group

S = ($p < 0.05$)

Table-2
Protective effects of *A. vera* on Serum ALP, AST, ALT and GGT Activities after 60 days exposure to SO₂ in albino rats

Groups	Exposure and pre-exposure supplementation	ALP (U/L)	AST (U/L)	ALT (U/L)	GGT (U/L)
		Mean±S.Em	Mean±S.Em	Mean±S.Em	Mean±S.Em
Control group-C ₂ (5)	Ambient air	161.2±2.50	113.8±3.12	49.0±3.21	14.85±0.80
Experimental group-E ₂ (5)	80ppm SO ₂ gas	196.4±7.05**	130.6±3.67**	62.6±1.56**	21.90±1.86**
Experimental group-E ₄ (5)	80ppm SO ₂ gas + <i>A. vera</i>	159.8±3.56 [•] VHS	112.6±1.69 [•] HS	47.0±1.87 [•] VHS	16.07±0.41 [•] S

5 = Number of rat
 S.Em. = Standard Error of Mean

Significantly different from control
 • = (p<0.05)
 ** = (p<0.001)

Significantly different from SO₂ exposed group
 S = (p<0.05)
 HS = (p<0.01)
 VHS = (p<0.00)

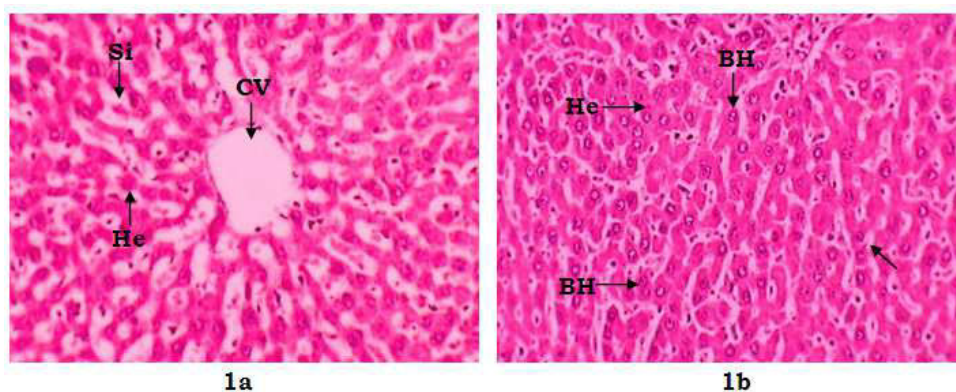


Figure. 1a and 1b
Liver section in control rats. Central Vein (CV), Hepatocytes (He), Sinusoids (Si) and Binucleated Hepatocyte (BH) (x400)

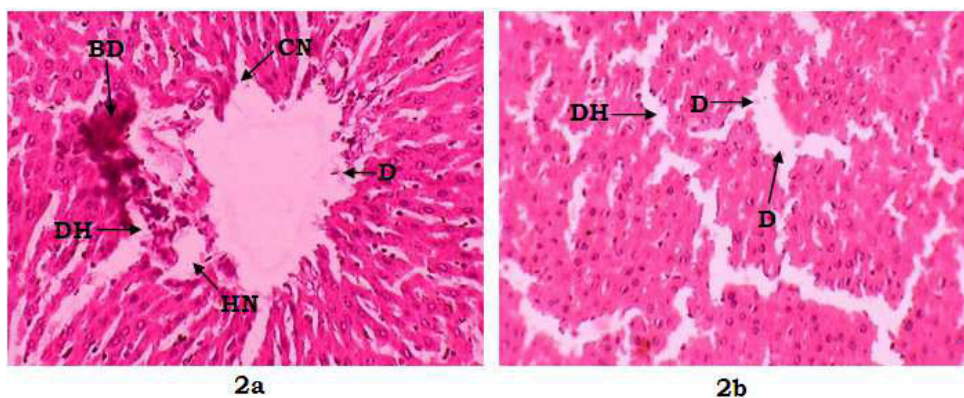


Figure. 2a and 2b
Liver section of SO₂ exposed rats after 30 days. Centrilobular Necrosis (CN), Degeneration of Hepatocyte (DH), Ballooning Degeneration (BD), Hepatocyte Necrosis (HN) and Cellular Debris (D) (x400)

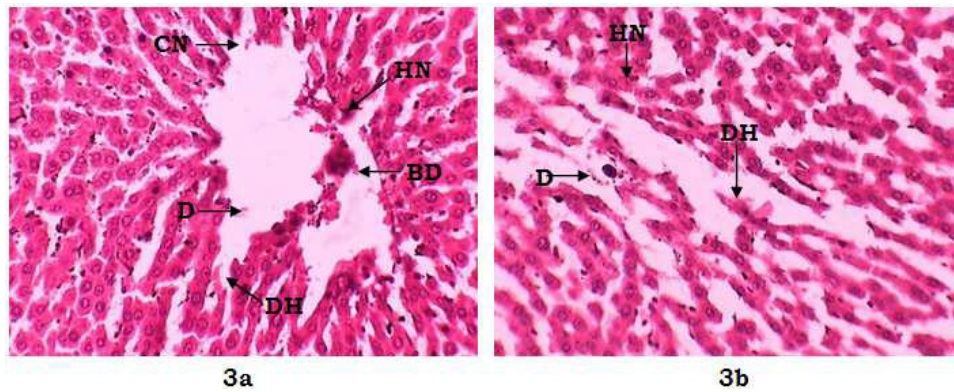


Figure. 3a and 3b
Liver section of SO₂ exposed rats after 60 days. Centrilobular Necrosis (CN), Degeneration of Hepatocyte (DH) Ballooning Degeneration (BD), Hepatocyte Necrosis (HN) and Cellular Debris (D) (x400)

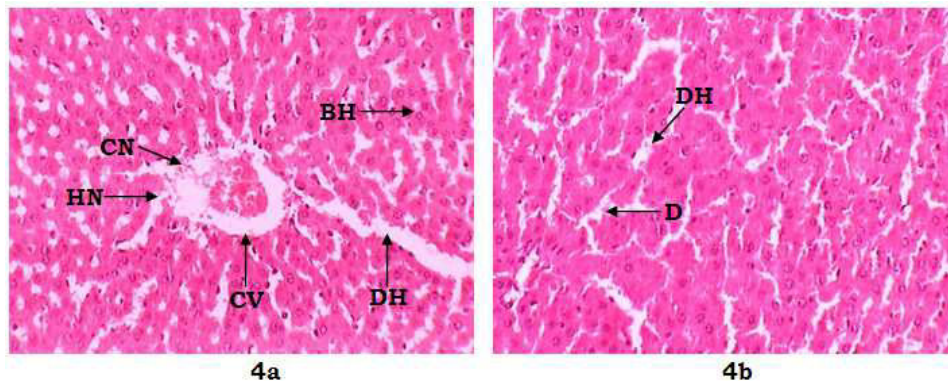


Figure. 4a and 4b
Liver section of SO₂ exposed rats with *A. vera* after 30 days. Centrilobular Necrosis (CN), Degeneration of Hepatocyte (DH), Hepatocyte Necrosis (HN) and Binucleated Hepatocyte (BH) (x400)

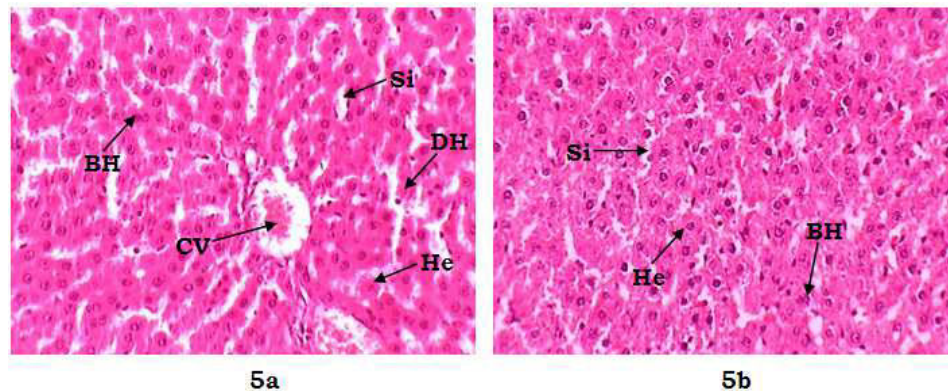


Figure. 5a and 5b
Liver section of SO₂ exposed rats with *A. vera* after 60 days. Central Vein (CV), Polygonal Hepatocyte (He), Sinusoids (Si) and Binucleated Hepatocyte (BH) (x400)

DISCUSSION

SO₂ gas is an oxidant and produces free radicals that can harm our body including disruption of living cells. The damage or death of the tissue usually the leakage of enzymes in the effective tissue and subsequent release in the blood stream¹³. In the present study, an increase in serum enzyme activity and histological alterations in the rat liver is the indication of hepatocellular injury accompanied with inflammatory responses of SO₂ gas¹⁴⁻¹⁸. Accumulation of free radicals in hepatic cellular component increases cellular degeneration of liver causing elevation in serum enzyme activity^{19,20}. The injury to hepatocytes increases the bile acid concentration responsible for leakage of ALP from rat liver^{21,22}. The elevated level of serum enzymes is indicative of impaired liver function, which is associated with the hepatic cell damage caused by SO₂^{3,23-25}. In the present study, a significant reduction in hepatotoxicity in albino rat after oral administration of aqueous extract of *A. vera* is the result of antioxidative and anti-inflammatory action of *A. vera* against SO₂ toxicity. It contains three malic acid acylated carbohydrates: veracylglycans A, B, and C,

REFERENCES

1. Wang XB, Hong-Fang J, Chao-Shu, T, Jun-Bao D, The biological effect of endogenous sulfur dioxide in the cardiovascular system. *Eur J Pharmacol*, 670(1): 1-6 (2011).
2. Kelly FJ, Oxidative stress: its role in air pollution and adverse health effects. *Occup Environ Med*, 60: 612 – 612 (2003).
3. Meng Z, Oxidative damage of sulphur dioxide on various organs of mice: Sulphur dioxide is a systemic oxidative damage agent. *Inhal Toxicol*, 15(2): 181-195 (2003).
4. Hukkeri VI, Jaiprakash B, Lavhale MS, Karadi RV and Kuppast IJ, Hepatoprotective activity of *Anthus exceles* Roxb. Leaf extracts on experimental liver damage in rats. *J Pharmacogn*, 11: 120 – 128 (2002).
5. Hamman J H, Composition and applications of *Aloe vera* leaf gel. *Molecules*, 13: 1599-1616 (2008).
6. Yagi A, Kabash A, Okamurs N, Haraguchi H, Moustafa SM and Khalifa TI, Antioxidant, free radical scavenging and anti-inflammatory effects of aloesin derivatives in *Aloe vera*. *Planta Med*, 68: 957-960 (2002).
7. Yagi A and Takeo S, Anti-inflammatory constituents, aloesin and aloemannan in *Aloe* species and effects of tanshinon VI in *salvia miltiorrhiza* on heart. *Yakugaku Zasshi*, 74: 517-532 (2003).
8. Singh N and Rao DN, Studies on the effect of sulphur dioxide on alfa alfa plants specially under conditions of natural precipitation. *Ind J Air Poll Contr*, 2(2): 55-69 (1979).

which are responsible for anti-inflammatory effects^{26,27}. Superoxide dismutase enzymes are responsible for antioxidant effects in reducing hepatic damage by the restoration of lipid peroxidation⁵. The administration of *A. vera* exhibits a significant reduction in Serum ALP, AST, ALT and GGT in SO₂ exposed rats. *A. vera* protect the tissues and enhance cellular regeneration by suppressing the oxidative damage resulting reduction in hepatocellular changes in rats. Aloe-emodin protects the liver of rats by reduction in the elevated ALT and AST activity²⁸. The shrunken nuclei of hepatocytes, granular cytoplasm, dilution of sinusoids and inflammation were reduced after administration of *A. vera* extract due to beneficial and protective effect in rats and mice^{29,30}. Present study reveals that *A. vera* aqueous leaf extract can use in modulating the liver injury against toxic effects induced by air pollutants.

ACKNOWLEDGEMENT

The authors are thankful to Dr. R.R. Singh, Department of Botany, Dr. B.R. Ambedkar University, Agra to provide *Aloe vera* leaf.

9. World Health Organization (WHO), Quality control methods for medicinal plants. WHO, Geneva, Switzerland 115-129 (2001).
10. Bradley DW, Maynard JE, Emery G and Webster H, Clin Chem, 18: pp1442 (1972).
11. Tietz NW, Clinical Chemistry. W.B. Saunders Co., Philadelphia, 29(5): pp751 (1983).
12. Humason GL, Animal tissue techniques. IVth ed. W.H. Feeman and Company, San Francisco, USA (1979).
13. Pant MC, Essentials of Biochemistry (9th ed.). Kedarnath Ramnath and Co. Meerut, pp 670 (2004).
14. Meng Z, Qin G, Zhang B and Bai J, DNA damaging effects of sulfur dioxide derivatives in cells from various organs of mice. Mutagenesis, 19: 465-468 (2004).
15. Bai J and Meng Z, Expression of apoptosis related genes in liver from rats exposed to sulphur dioxide. Toxicology, 216: 253-260 (2005).
16. Meng Z and Liu Y, Cell morphological ultra-structural changes in various organs from mice exposed by inhalation to sulphur dioxide. Inhal Toxicol, 19: 543-551 (2007).
17. Rajaii F, Khaki AA, Khaki A, Knorshid F, Borhani N, Jfrai H, Hagh dust H and Gneibi N, Histopathological effects of sulfur dioxide in mouse liver following the chronic and acute exposure. J Bio Sci, 8(7): 1241-1245 (2008).
18. Zhao H, Xu X, Na J, Hao L, Huang L, Li G and Xu Q, Protective effects of salicylic acid and vitamin C on sulfur dioxide induced lipid peroxidation in mice. Inhalation Toxicology, 20(9): 865-871 (2008).
19. Gopal DV and Rosen HR, Abnormal findings in liver function tests. Postgrad Med, 107(2): 100-114 (2000).
20. Karakilcik, AZ, Hayat A, Aydilek N, Zerim M and Cay M, Effects of vitamin C on liver enzymes and biochemical parameters in rats anesthetized with halothane. Gen Physiol Biophys, 24: 47-55 (2005).
21. Kaplan MM, Serum Alkaline phosphatase: Another piece is added to the puzzle. Hepatology, 6: 526-528 (1996).
22. Moss DW, Physiochemical and pathophysiological factors in the release of membrane bound alkaline phosphatase from cells. Clin Chim Acta, 257(1): 133-140 (1997).
23. Gumuslu S, Akbas H, Aliciguzel Y, Agar A, Kucukatay V, Yargicoglu P, Effects of sulphur dioxide inhalation on antioxidant enzyme activities in rat erythrocytes. Ind Health, 36: 70-73 (1998).
24. Al-Malki AL, Rezaq AM and Al-Saedy MH, Effect of fire smoke on some biochemical parameters in firefighters of Saudi Arabia. J Occup Med Toxicol, 3: 33-46 (2008).
25. Agarwal, A, Yadav M and Goyal PK, Sulphur dioxide gas induced histopathological changes in liver of albino rat. J. Ecophysiol. Hlth., 9: 55-58 (2009).
26. Esua MF and Ravwald JW, Novel bioactive methyl glucans from *Aloe vera* gel: isolation, structure elucidation and in vitro bioassay. Carbohydr Res, 341: 355-364 (2006).
27. Steenkamp V and Stewart MJ, Medicinal applications and toxicological activities of *Aloe* products. Openup-October 2: 1-13 (2007).
28. Alqasoumi, SI, Al-Howiring TA and Abdul-Kader MS, Evaluation of the Hepatoprotective effect of *Aloe-vera*, *Clematis hirsute*, *Cucumis prophetorum* and *Bee propolis* against experimentally induced liver injury in rats. Inter J Pharmacol, 4(3): 213-217 (2008).
29. Noor A, Gunasekaran S, Manickam AS and Vijayalakshmi MA, Antidiabetic activity of *Aloe vera* and histology of organs in streptozotocin-induced diabetic rats. Current Science, 94: 1070-1076 (2008).
30. Kim SH, Cheon HJ, Yun N, Oh ST, Shim E, Shim KS and Lee SM, Protective effect of a mixture of *Aloe vera* and *Silybeem marianum* against carbon tetrachloride induced acute hepatotoxicity and liver fibrosis. J Pharmacol Sci, 109(1): 119-127 (2009).

