



**A STUDY ON PHYTOCHEMICAL AND ANTIFUNGAL ACTIVITY
OF LEAF EXTRACTS OF *TERMINALIA CATAPPA***

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ABSTRACT

The antifungal activity of *Terminalia catappa* leaves was tested against five allergenic and pathogenic moulds. An initial screening of effectivity of different solvent extracts was done by phytochemical analysis and *in-vitro* antifungal activity. The antifungal activity was determined by both food poison technique and by cup-well diffusion method for five test organisms viz *Aspergillus niger*, *Alternaria alternata*, *Curvularia lunata*, *Penicillium chrysogenum*, and *Trichophyton tonsurans*. Methanol extract showed significant antifungal activity. The MIC evaluated at three different concentrations, it was not concentration dependent activity. Most susceptible mould was *Curvularia lunata* and least susceptible was *Alternaria alternata* as compared with standard antifungals. Hplc analysis revealed presence of a known flavonoid component identified as quercetin.

KEYWORDS: *Terminalia catappa*, phytochemical analysis, medicinal plants, bioactive compounds, antifungal activity



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INTRODUCTION

The reduced number of herbal drugs available against pathogenic fungi makes it necessary to discover new classes of antifungals and compounds that inhibit their resistant mechanisms. This has led to the search for therapeutic alternatives, particularly among medicinal plants and compounds isolated from them that are used for their empirically antifungal properties¹. Ethnopharmacological surveys performed around the world have mentioned that among the plant species belonging to combretaceae family, *Terminalia catappa* is the most requested medicinal plant. *Terminalia catappa* family Combricitaceae (Tropical almond) has a vast natural distribution in near coastal areas of the Indian Ocean, through Tropical Asia and into Pacific Ocean. It is fast growing and easily propagated from seed that flourishes with minimal maintenance in suitable environments.² The decoction of leaves is used in Nigeria, as medicine against malaria and abdominal pains. In Togo & Benin root bark decoction is used in the treatment of various dermatosis. Similarly in Phillipines, leaves extract is used against Leprosis³. The leaf, bark and fruit of this plant have long been used in folk medicine for antidiarrheic, antipyretic and haemostatic purpose in India, Phillipines, Malaysia and Indonesia. *T.catappa* leaf has been reported to possess antioxidative, hepatoprotective, antidiabetic, anti-inflammatory and anti-HIV reverse transcriptase activity. The presence of wide number of bioactive compounds with variety of biological activities in *T. catappa* attracted us to select this plant for the present study.⁴ Phytochemical analysis of active plant extracts for qualitative identification of the major phyto constituents and anti fungal activity is also reported here.

MATERIALS AND METHODS

COLLECTION OF PLANT MATERIAL

Twigs of *Terminalia catappa* Linn were collected from Bhopal. The plant was identified from Govt. M.V.M. college with the help of the project

report of Dr.Madhuri Modak, who has confirmed the identification from Botanical Survey of India, Allahabad. The leaves were separated washed and dried at room temperature. The dried leaves were powdered in a mixer blender and stored at room temperature in close containers in dark until used. A herbarium of the same was deposited at SNGGPG autonomous Girls college, Bhopal (M.P.).

SOURCE OF MICRO-ORGANISMS

Fungi *Aspergillus niger*, *Alternaria alternata*, *Curvularia lunata*, *Trychophyton tonsurans* and *Penicillium chrysogenum* selected were isolated from aeromycoflora of Bhopal based on their allergenic and pathogenic nature. Fungal mycelia from the culture were stained with *lactophenolcottonblue* and identified by using compound microscope at 40x objective (Olympus make). The species were initially identified with reference to standard literature. For confirmation of identification cultures were sent to NCFT (National Centre of Fungal Taxonomy), Delhi.

EXTRACT PREPARATION

The powdered leaves (25g) of *T.catappa* was extracted separately to exhaustion in a soxhlet apparatus using ethanol, methanol, petroleum ether and acetone as solvent system. All the extracts were filtered through Whatman filter paper no.1 and then concentrated in hot air oven at low temperature 40⁰-50⁰C. A yield of 11.6% from ethanol, 12.2% from methanol, 1.6% petroleum ether and 1.8% acetone solvent system were obtained.⁵

PHYTOCHEMICAL ANALYSIS OF LEAF EXTRACT

Specific qualitative tests were performed to identify bioactive compounds of pharmacological importance through standard methods as described by Harbone⁶.

Test for alkaloids (Mayer's test):- To 2.3 ml. of filtrate a few drops of Mayer's reagent was

added, appearance of ppt. indicates the presence of alkaloids. Wagner's test:- To 2-3 ml. filtrate was treated with few drops Wagner's reagent, reddish brown precipitate appeared.

Test for glycosides (Keller Killiani test) :- To the 2ml extract, a few drops of glacial acetic acid, one drop 5 % FeCl₃ and conc. H₂SO₄. were added, reddish brown color appeared at the junction of the two liquid layer and upper layer appeared bluish green.

Test for tannic and phenolic compound:- To 2-3 ml. of aqueous or alcoholic extract, added few drops of following reagent.

a. 5% FeCl₃ solution --- deep blue black colour.

b. Lead acetate solution --- white ppt.

Dilute Iodine solution --- red colour

Test for Saponin:- To 2 ml extract 5 ml. distilled water was added and heated to boil. Frothing appeared, showed the presence of saponin.

Test for Flavonoids:- To small quantity of residue, (0.1%) lead acetate solution was added. Yellow colour ppt. formed. Addition of increasing amount of sodium hydroxide to the residue showed decoloration.

Test for Steroid Salkowski reactions – To 2 ml. of extract add 2 ml. chloroform and 2 ml conc. H₂SO₄ and shaken well. Chloroform layer appeared red and acid layer showed greenish yellow fluorescence.

Test For Carbohydrate Molish test:- To 2-3 ml aqueous extract, add a few drops of α naphthol solution in alcohol. Mix was shaken and conc. H₂SO₄ was added from sides of the test tube. Violet ring formed at the junction of two liquids showed the presence of carbohydrate in the extract.

Test for non- reducing polysaccharides (Starch) Iodine test : Mixed 3 ml test solution and few drop of dilute iodine solution. Blue colour appeared.

Tannic acid test for Starch : 20% tannic acid was added in the test solution. Precipitate appeared.

Test for Protein Biuret test: To 3 ml test solution added 4% NaOH and few drop of 1% CuSO₄ solution violet colour appeared. Millon's test: To 2 ml. of test solution, Millon's reagent was added, white precipitate appeared.

ANTIFUNGAL ACTIVITY STUDY

The antimicrobial activities of all the extracts were determined by Food poison technique (The fungitoxicity of the extracts in terms of percentage inhibition of mycelial growth was calculated by using the formula

$$\% \text{ inhibition} = \frac{dc-dt}{dc} \times 100$$

Where:

Dc = Average increase in mycelial growth in control.

Dt = Average increase in mycelial growth in treatment.)⁷ and Agar well diffusion method⁸

SCREENING OF SUSCEPTIBLE FUNGI

One gram of each of the dried evaporated solvent extract of all the solvents was dissolved in 10ml of the respective solvent. 500 μ l of each solvent extract was amended with 15 μ l of PDA medium before solidification. The medium having only solvent served as control. Test fungi were inoculated and after incubation at 25 \pm 2^oC for two days the percent inhibition of mycelia growth was determined. The colony diameter was measured in millimeter. For each treatment three replicates were maintained,

MIC

Minimum inhibitory concentration of effective solvent extract was determined by cup well method⁹. Three different concentrations of effective extract 100mg/ml, 250mg/ml and 500mg/ml and 10mg/ml of standard were taken for the assay. The wells were filled with 50 μ l each of controls and test concentrations using a micro titre pipette. The plates were allowed to diffuse at room temperature for two hour. The plates were then incubated at 25^oC for two days and the zone of inhibition was measured.

THIN LAYER CHROMATOGRAPHY (TLC) OF LEAF EXTRACTS.

TLC for methanolic leaf extract that showed strong antimicrobial activity was carried out on prepared TLC microslides .¹⁰ with Silica gel G. About 20 μ l of plant extract was spotted on TLC chromatogram. A solvent system consisting of Chloroform : Ethyl Acetate (6 : 4 v/v) was used for separation of compounds. Individual R_f for

each spot was measured. TLC spots were visualized under UV light.

HPLC analysis

The HPLC analysis of leaf extract was carried out using Waters(make) HPLC system, having Waters 486 UV visible detectors, equipped with rehodyne injector, HPLC cartridge system, Thermo C18 (250x4.6mm), 5µm column. A UV visible detector with variable wavelength was used for the separation of different bioactive compounds. Sample preparation for HPLC analysis :- Methanol extract of *T. catappa* was separated using a mobile phase of 50Mm KH₂PO₄ Buffer (pH-3 with OPA): Acetonitrile in the ratio of (30:70 v/v) to a flow rate of 1.0ml/min, with column temperature 25°C. Injection volume was 20ul and detection was carried out at 360nm.

STATISTICAL ANALYSIS

PHYTOCHEMICAL ANALYSIS

PHYTOCHEMICALS	METHANOL	ETHANOL	PET. ETHER	ACETONE
Alkaloids	+	-	-	+
Glycoside	+	+	-	-
Saponins	+	+	-	+
Tannic & Phenolic	+	+	-	+
Flavonoid	+	+	-	+
Steroid	+	+	-	-
Terpenoid	-	-	-	-
Sugars	-	-	-	-
Proteins	+	-	-	-

Table 1
Showing results of phytochemical analysis.

SUSCEPTIBILITY TEST AGAINST DIFFERENT ORGANIC SOLVENT EXTRACTS

The antifungal activity of *Terminalia catappa* was evaluated against five allergenic and pathogenic moulds. An initial screening of effectivity of ethanol, acetone, methanol and petroleum ether fraction on fungi was performed by food poison technique. Among the four extracts used, methanol extract for all five fungi showed significant antifungal activity. The percentage of inhibition of methanol extract was more than 50% and it was dose independent activity. Among the five fungi tested, *Curvularia* was found to be most sensitive and *Alternaria* to

The results were statistically analysed by Tukeys method using ANOVA software.

RESULTS AND DISCUSSION

RESULTS

Phytochemical characteristics of leaf extracts of *Terminalia catappa* in four different solvent tested are summarized in Table 1. The leaf extracts in different solvents were screened for the presence of various bioactive phytochemical compounds. The analysis revealed presence of bioactive compounds such as terpenes, saponins, steroids, flavonoids, phenols and glycosides in ethanol fraction. The methanolic fraction contained alkaloid also. The therapeutic qualities of this plant can be attributed to the presence of chemical compounds like terpenes, phenols, saponins and flavonoids.

be least sensitive. It has been observed that ethanolic and methanolic fractions were effective against all the organisms in varying levels of sensitivity. However the extracts in acetone and petroleum ether fraction did not showed any remarkable antimicrobial activity. *Curvularia lunata* was found to be most susceptible to ethanol fractions of *Terminalia catappa*. With an inhibition of 74% of growth where as *Trychophyton* and *Alternaria* were least susceptible. While comparing the solvents, methanol fraction was found to be more effective than ethanol fraction on all the fungi. In methanolic extract, *Aspergillus niger*, was most susceptible with 68.7% growth inhibition,

followed by *Penicillium*, *Trychophyton*, analysis are shown in table 2 and fig 1. *Curvalaria* and *Alternaria*. Results of antifungal

Table 2
Showing results of screening of effective leaf extract of
***Terminalia catappa* in various solvents.**

S. No.	Leaf extracts	% of mycelial growth inhibition of various Fungi				
		<i>Aspergillus niger</i>	<i>Alternaria alternata</i>	<i>Curvalaria lunata</i>	<i>Penicillium chysogenum</i>	<i>Trychophyton tonsurans</i>
1	Ethanol Extract	57.33 ± 0.33	24.66 ± 0.33	74.7 ± 0.28	50.8 ± 0.27	43.7±0.31
2	Methanol extract	68.7 ± .33	35.3 ± 1.45	60 ±0 .51	64 ± .15	61±0.25
3	Petrolium ether	64 ± 1.15	27.33 ± 0.18	36.67 ± 0.85	51 ± 0.57	61.67±1.53
4	Acetone	43.7 ± 0.31	31.7 ± 0.15	24.6 ± 0.33	43.7 ± 0.31	27.33±.18
5	Control Ketoconazole (10 mg/ ml)	63 ± 0.33	ND	ND	45 ± 0.57	71±0.33
6	Carbendazim (10 mg/ ml)	ND	75 ± .18	60 ± 0.33	ND	ND

Replicates = three

Result were expressed as mean ± SEM

All the results were found to be significant at $p < 0.01$ and $p < 0.05$

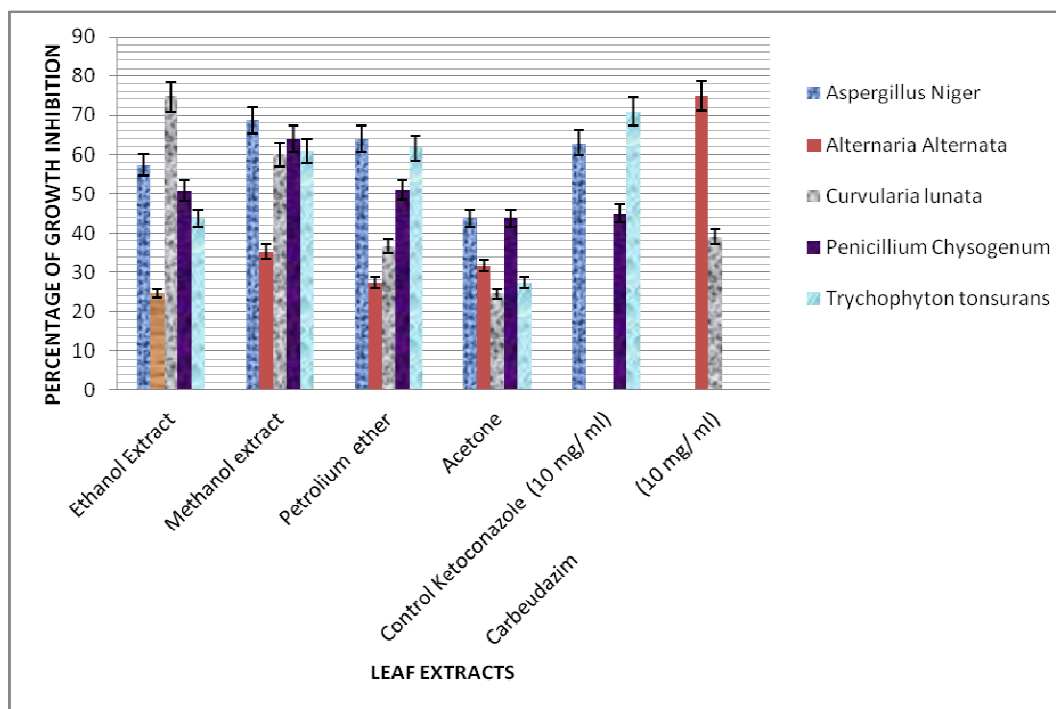


Figure 1

shows antifungal activity of leaf extracts of *Terminalia catappa* against various fungi.

MIC

Since, *Curvalaria* was found to be more susceptible to ethanol and methanol fractions, MIC was determined by cup well method against it. Three concentrations of 100mg/ml, 250mg/ml and 500mg/ml were tested against the fungi. Significant antifungal activity was recorded in 250mg/ml fraction of ethanol and 100mg/ml of methanol of *T. catappa*. Table 3.

Sr no.	Fungi	Ketoconazole /carbendazium diameter in mm	Ethanol mg/ml			Methanol mg/ml		
			100	250	500	100	250	500
1	A.niger	14.6±0.33	-	-	9.6±0.33	12.66±0.66	11.66±0.33	12.6±0.46
2	Alternaria alternate	23±0.12	-	-	13±0.57	-	-	19.66±0.33
3	Curvularia lunata	29.6±0.33	-	17.16±0.59	21±0.57	9.6±0.33	19.6±0.33	12.6±0.21
4	Trychophyton tonsurans	21.66±1.19	-	-	-	-	-	-

Table 3

Inhibition zone diameter of ethanol and methanol extracts of Terminalia catappa

RESULTS OF TLC AND HPLC

Quercetin was isolated from the methanol extract and then detected on Tlc plate in comparison with standard flavonoid, quercetin. The retention factor value of 0.42 was found to be similar to that of standard quercetin. Also, characterisation of methanolic extract through Hplc analysis, revealed that fraction of *T.catappa* leaves contain a bioactive

component quercetin, one of the flavonoid component. The retention time of standard quercetin was recorded 6.235 min while the retention time of the sample was found to be 6.159 min. The amount of quercetin was calculated by extrapolating the value of area from the calibration curve. The percentage assay of quercetin after calculation was found to be 3.940. Table 4 and table 5.

The Rf value of TLC of T.Cattapa were found as follows.

Extract	Chloroform : Ethyl Acetate (6 : 4 v/v)
Terminalia cattapa (Methanolic)	2.60/6 = 0.43

Table 4

Result of Assay of Quercetin in methanol Extract

Name of Extract	% Assay
Terminalia cattapa (Methanol)	3.940

Table 5

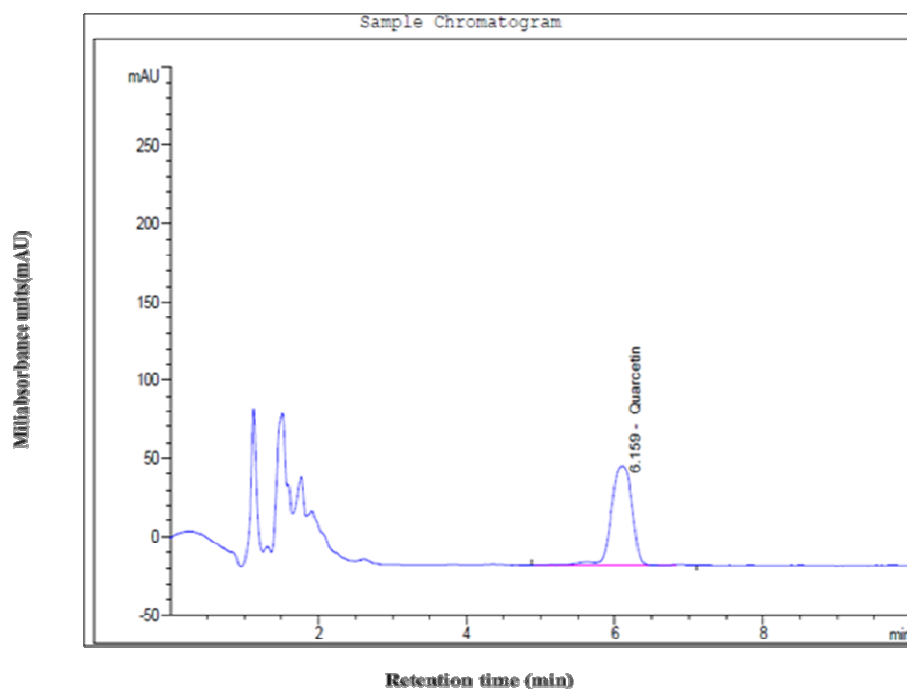


Figure 2
Sample Chromatogram of Extract Terminalia cattapa (Methanol)

DISCUSSION

The leaf, bark and fruit of this plant have long been used in folk medicines for antidiarrheic, antipyretic, haemostatic purposes in India. Phytochemical screening of ethanolic extracts of *T. catappa* shows the presence of glycosides, steroids, flavonoid, tanins, phenolic compounds and saponins. The earlier work on methanolic leaf extracts of *T. catappa* has also reported the presence of these components.^{11,12} Earlier report on phytochemical analysis of pericarp of *T. chebula* fruit has also shown alcohol to be best extractant of bioactive components.¹³ The presence of antifungal activity in alcoholic fraction of *Terminalia* leaf extracts finds agreement with the work of several other workers.¹⁴⁻¹⁹ Hydroalcoholic extracts of *T. catappa* and *T. mantaly* have been reported to inhibit the *in vitro* growth of *Aspergillus fumigatus*.²⁰ Several studies have reported the presence of bioactive compounds that are responsible for the medicinal properties of the plant that is used for treatment of different ailments. Some important antimicrobial phytoconstituents of *T. catappa* leaf reported

are tannins (terflavins, tergalagin), flavonoids (rutin) and triterpenoids.^{21,22} TLC and HPLC of methanolic leaf extracts showed the presence of quercetin (flavonoid) as active constituent. These findings correlate with the other findings where an average proportion of flavonoids was reported from various fractions of same plant.²³ In yet another work monomeric flavonoids were detected as active constituents in *E. officinalis*.²⁴ The present study on antifungal screening also justifies the traditional uses of *T. catappa* in treatment of various ailments. Study of the synergistic interaction of active phytocompounds with antibiotics is required to exploit the use of this plant extracts in the combination therapy of infectious diseases caused by multidrug resistant organism.

CONCLUSION

Since growth of *Curvularia lunata*, *Aspergillus niger*, *Penicillium* and *Trichophyton tonsurans* was suppressed by more than 50%, the

alcoholic extracts are effective against them. This study has highlighted *T. catappa* as a potential antifungal agent against *Curvularia lunata* under *in vitro* conditions. It is concluded

that antifungal activity of methanolic leaves extract of *T. catappa* and its active constituents would be helpful in interacting various kinds of human allergies and plant diseases

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REFERENCES

1. Abad, M.J., Ansuategui, M. and Bermejo, P. Active antifungal substances from natural sources. *Arkivoc.*, 7: 116-145. (2007).
2. Thomson, Lex A.J, Evans, Berry. Species profiles for Pacific Island Agroforestry www.traditionaltree.org. (2006)
3. Noel Z.G, Koffi, N.G., Justin, K.N., Kiyinima, C. and Joseph, D.A Evaluation and comparison of antifungal activities of *Terminalia Catappa* and *Terminalia mantaly* (combretaceae) on the *in vitro* growth of *Aspergillus fumigatus*. *Journal of Medicinal Plants Research* Vol 6(12)pp. 2299-2308. (2012).
4. Annegowda H.V, Mordi, M.N., Ramanathan, S. and Mansor, S.M Analgesic and antioxidant properties of Ethanolic extract of *Terminalia Catappa* L. leaves. *International Journal of pharmacology*, 6:910-915. (2010).
5. Elumali, E. K., Chandrasekaran, N., Thirumalai, Sivakumar, T.C., Viviyani, Therasa, S., David E *Achyranthes Aspera* leaf extracts inhibited fungal growth. *International journal of Pharmtech Research* Vol. No.4pp 1576-1579. (2009).
6. Harborne, J.B., *Phytochemical method, A guide to modern technique of plant Analysis*, 3rd Edition Chapman and Hall. New York. pp 1-198. (1998)
7. Satish, S., Mohana, D.C., Raghvendra, M.P. and Raveesha, K.A. Antifungal activity of some plant extracts against important seed borne pathogens of *Aspergillus* spp. *Jr. of Agriculture Technology* p.109-119.
8. Omenka, C.A. and Osuoha, J.O. Antimicrobial potency of grape fruit seed extract on five selected pathogens. *Nigerian journal of Microbiology*. (14)2: 39-42. (2000)
9. Kareem, S. O., Akpan, I. and Ojo, O.P. Antimicrobial activities of calotropis *Procera* on selected pathogenic biomedical research, vol 11p105-110. (2008)
10. Al-Bayati, F.A., Isolation and identification of antimicrobial compound from *Mentha longifolia* (L) leaves grown wild in Iraq. *Annals of clinical Microbiology and Antimicrobials* 8:20 (2009).
11. Babayi H., Kolo I., Okugun J.I. and Ijah, U.J.J., The antimicrobial extracts of *Eucalyptus camaldulensis* and *Terminalia catappa* against some pathogenic microorganisms. *Biokemistri*, vol 16, no.2:106-111 (2004)
12. Akharaiyi, F., C., Ilori, R.M. and Adesida J., A., Antibacterial effect of *Terminalia catappa* on some selected pathogenic bacteria. *Int jr. Pharm Biomed Res.* 2 (2) :64-67 (2011).
13. Rooplata U.C. and Vijay Mala Nair. The Phytochemical screening of pericarp of fruits of *Terminalia chebula* Retz. *Int.J Pharm. Bio Sci* July ; 4 (3) (p)550-559
14. Parekh, J. and Chanda, S. *In vitro* antifungal activity of methanol extracts of some Indian medicinal plants against pathogenic yeast and moulds. *African journal of Biotechnology* vol 7(23), pp 4349-4353; (2008).

15. Manjur,A.,Raju and S.Rahman Antimicrobial activity of Terminalia catappa extracts against some pathogenic microbial strains .Pharmacology and Pharmacy,2 (4) 299-305 (2011)
16. Shinde, S.L, Junne, S.B, Wadje, S.S, Baig, M.M. The diversity of antibacterial compounds of Terminalia species (combretaceae).Pak J Biol Sci 15;12(22):1483-6 (2009)
17. Elizabeth, K.M. Antimicrobial activity of Terminalia bellerica . Indian journal of clinical biochemistry: 20(2): 150-153. (2005)
18. Lee, S.H., Chang K.S., Sums, Huang, Y.S. and Jang, H.D. Effects of some Chinese medicinal plant extracts on five different fungi .Food control 2006:18:1547-1554.
19. Parekh, J. and Chanda, S. Screening of aqueous and alcoholic extracts of some Indian medicinal plants for antibacterial activity. Indian journal of pharmaceutical science 68 (6): 835-838. (2006)
20. NOEL, ZIRIHI G.; NGUESSAN Koffi,KASSY N`dja Justin ,COULIBALY kiyinlma and Djaman Allico Joseph.Evaluation and comparision of antifungal activities of Terminalia catappa and Terminalia mantaly (combretaceae) on the in vitro growth of Aspergillus fumigatus. Jr. of medicinal plant research vol.6 (12) 2299-2308 (2012)
21. Tanaka, T., Morita, A., Nanaka, G. Tanins and related compounds CIII isolation and characterization of new monomeric, dimeric and trimeric ellagitanins ,calamanisanin and calaminins A, B, and C from Terminallia camanisani. 38,60 (1991)
22. Lin,Y.L., Kuo, H., Shiao, M.S. Chen C.C. and J.J.ouflavonoids glycosides of terminellia catappal.,J. chinese chem. Soc.47 : 253-256 (2000)
23. Venkatachalam, T.,V. K. Kumar,P.K.Selvi,A.O.Maske, S.Kumar).physicochemical and preliminary phytochemical studies on the Lantana camara(L.) fruits.International jr. of pharmacy and pharmaceutical sciences.3(1)52-54. (2011)
24. Ahmed,I., and A.Z. Beg.(2001). Antimicrobial and phytochemical studies on 45 Indian medicinal plants against multidrug resistant human pathogens. Jr. of ethanopharmacology 4: 113-123