



EFFECT OF ETHANOLIC EXTRACTS OF *CYAMOPSIS TETRAGONOLOBUS* AND *CYPERUS ROTUNDUS* ON BIOCHEMICAL PARAMETERS OF DIABETIC CATARACT INDUCED WISTAR ALBINO RATS

SEEMA SURENDRAN¹ AND VIJAYALAKSHMI KRISHNAMOORTHY*²

¹Research Scholar, Research and Development Centre, Bharathiar University, Coimbatore, Tamil Nadu, India

²Associate Professor, Department of Biochemistry, Bharathi Women's College (Autonomous), North Chennai, Tamil Nadu, India

ABSTRACT

The present study evaluated the effect of ethanolic extracts of *Cyamopsis tetragonolobus* rhizomes and *Cyperus rotundus* pods on the biochemical parameters of diabetic cataract induced Wistar albino rats. Healthy rats were grouped into 4: Group I – Normal, Group II – Diabetic cataract induced animals (STZ-induced diabetic rats which developed cataract), Group III – *Cyamopsis tetragonolobus* extract co-treated animals and Group IV – *Cyperus rotundus* extract co-treated animals (Treatment for 19 weeks after STZ induction). Body weight and organ weight of cataract animals decreased while plant treatment prevented this loss. Increase in values of glucose, glycated haemoglobin, insulin, creatinine, urea and uric acid levels were observed in diabetic cataract induced rats, while plant co-treated groups restored these levels. The changes in lipid and protein profile in Group II rats were prevented by the plant treatment. SGOT, SGPT, ALP and LDH increased in Group II but plant treatment retained enzymatic activities. The results signify the plant's protective nature.

KEYWORDS: Diabetic cataract, *Cyamopsis tetragonolobus*, *Cyperus rotundus*, Streptozotocin, Biochemical parameter



VIJAYALAKSHMI KRISHNAMOORTHY

Associate Professor, Department of Biochemistry, Bharathi Women's College (Autonomous), North Chennai, Tamil Nadu, India

*Corresponding author

INTRODUCTION

Diabetic cataract is one of the major complications of diabetes which affects most of the patients. Cataractogenesis is one of the early secondary obstacle of diabetes mellitus which is a severe metabolic disorder characterized by hyperglycemia. Cataracts which occur in diabetic patients may cause blindness¹. Though cataract surgery provides the possibility of regaining vision, it has some disadvantages. It will not resolve all the vision problems². Medicinal plants are proven to be the resources of new drugs. They constitute a rich source of bioactive compounds and are found free from adverse effects but have exceptional pharmacological actions³. *Cyamopsis tetragonolobus*, commonly referred as cluster bean or guar bean is the source of guar gum. The guar bears pods which contain many seeds⁴. *Cyamopsis tetragonolobus* is traditionally used to cure inflammation, sprains and arthritis. They are also used as an antioxidant, antibilious and laxative agent⁵. *Cyperus rotundus* also known as nutgrass or nut sedge is a major weed. It has tuberous roots or rhizomes⁶. It is used for treating a wide range of diseases such as scabies, eczema, itching, fever, cough and asthma⁷. Many natural products and herbal medicines have been well recommended for the treatment of diabetes. Most of the plant extracts exhibited hypoglycaemic, hypolipidemic and antioxidant effects in experimental animals as well as in humans, which are helpful in treating diabetes and its associated complications³. In our present study, we have investigated the effect of the ethanolic extracts of *Cyamopsis tetragonolobus* pod and *Cyperus rotundus* rhizome on the biochemical parameters of diabetic cataract induced Wistar albino rats.

MATERIALS AND METHODS

(i) Plant Collection

The rhizomes of *Cyamopsis tetragonolobus* and pod (fruit) of *Cyperus rotundus* were obtained from medicinal plant vendor in Chennai, Tamil Nadu, India without any external defects.

(ii) Authentication of Plants

The selected plant materials were identified and authenticated as PRAC/2010/495 for *Cyperus rotundus* and as PRAC/2010/494 for *Cyamopsis tetragonolobus* by a Botanist Prof. P. Jayaraman, Plant Anatomy Research Center, West Tambaram,

Chennai, Tamil Nadu, India. The rhizome and pods were shade dried at room temperature for a week and made into coarse powder.

(iii) Extraction of Plants

20gm coarse powder of rhizomes of *Cyperus rotundus* and 20gm coarse powder of pods of *Cyamopsis tetragonolobus* were homogenized in 100ml of 90% ethanol separately using Waring blender. The extraction procedure was carried out as mentioned in our earlier study⁸. The ethanolic extracts were concentrated under reduced pressure and preserved at 5°C in airtight bottle until further use.

(iv) Animal Study

Eight week old healthy Wistar albino rats of either sex weighing 120±30gm were purchased from King Institute, Guindy, Chennai, Tamil Nadu, India. The animals were maintained at Saveetha University, Chennai, Tamil Nadu, India under standard conditions of humidity (45-55%), temperature (25±2°C) and light (12hr light/12hr dark) (IAEC No. Biochem BWC 010/10). They were fed with standard pelleted diet and given free access to water *ad libitum*. Experimental animals were handled according to the University and Institutional legislation, regulated by the committee for the purpose of Control and Supervision of Experiments on Animals (CPCSEA), Ministry of Social Justice and Empowerment, Government of India.

(v) Experimental Design

The experimental animals were divided into 4 groups as given below:

Group I – Normal control (6 animals)

Group II – Diabetic cataract induced rats (6 animals)
Induction of Diabetic Cataract: Diabetes was induced by intraperitoneal injection of streptozotocin (STZ) (purchased from Sigma, USA) (60mg/kg body weight) dissolved in citrate buffer pH 4.5⁹ and maintained for 20 weeks to develop cataract and the presence of cataract was confirmed using slit lamp biomicroscope.

Group III – After STZ induction for one week, co-treatment with *Cyamopsis tetragonolobus* pod extract was done for 19 weeks [200mg/kg body weight of the plant extract were orally fed to experimental animals] (6 animals)

Group IV - After STZ induction for one week, co-treatment with *Cyperus rotundus* rhizome extract was done for 19 weeks [200mg/kg body weight of

the plant extract were orally fed to experimental animals] (6 animals)

(vi) Collection of Blood and Organs

All the animals were killed by cervical decapitation after the experimental period. The blood was collected without EDTA for the separation of serum. The lens was removed from the eyes and homogenized with motor driven Teflon coated homogenizer in ice-cold 0.1M Tris-HCl buffer pH 7.4 to obtain 10% homogenate.

(vii) Body weight and Organ weight

The body weight of rats was checked using a top loader weighing balance. The body weight is expressed in grams. The vital organs such as eyes, liver, kidney, spleen, brain, pancreas and heart were collected. Blood was removed from the organs with the filter paper and their weights were assessed with an electric weighing balance. The organ weights were expressed in grams or milligrams.

(viii) Assessment of biochemical parameters in blood and serum

Blood glucose¹⁰, serum glycated hemoglobin¹¹, total protein¹² and albumin¹³ were estimated. Globulin present in serum can be calculated by subtracting the measured albumin from total protein. Serum insulin levels were assayed using a standard Mercodia Rat Insulin ELISA enzyme immunoassay kit from Mercodia, Sweden (cat.no-10-1124-01). The

assay was performed according to manufacturer's protocol. Blood urea¹⁴, serum uric acid¹⁵ and creatinine¹⁶ were also estimated. Lipid was extracted¹⁷ from serum and cholesterol¹⁸, triglyceride¹⁹, free fatty acid^{20,21}, lipoprotein²², and HDL²³ were estimated. LDL and VLDL levels were calculated by Friedewald formula. Activities of SGOT²⁴, SGPT²⁴, ALP²⁵ and LDH²⁶ in serum were also determined.

(ix) Statistical Analysis

Statistical analysis was done by using SPSS 16.0.1. All results were presented as mean value \pm standard deviation (SD) for six samples in each group. Within group comparisons were performed with Tukey's ANOVA test. Significant differences between the control and experimental groups were assessed by the Student's t-test using Graph pad prism 6.0.

RESULTS

1. Cataract Incidence

The occurrence of cataract was observed after 20 weeks of STZ administration and confirmed by slit-lamp biomicroscope to investigate lenticular opacity. Diabetes-induced cataract gradually progressed in STZ-administered rats alone and the cataract was graded as cortical cataract (Figure 1).

Figure 1
Diabetic Cataract Rat with clouding of eyes

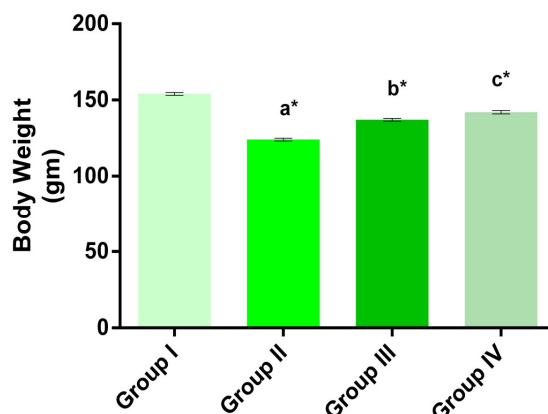


The experimental animals co-treated with the ethanolic extract of *Cyamopsis tetragonolobus* pod and *Cyperus rotundus* rhizome for 19 weeks after STZ administration (after 1 week period) did not show the development of cataract.

2. Body Weight

Different groups of rats showing their body weight is depicted in Figure 2. Diabetes-induced cataract animals showed significant decrease ($p < 0.001$) in body weight when compared with control animals. The ethanolic extract of *Cyamopsis tetragonolobus* treated animals and *Cyperus rotundus* treated animals were found to be have significant increase ($p < 0.001$) in body weight when compared with diabetes-induced cataract animals.

Figure 2
Body weights of experimental animals



Values are expressed as mean \pm SD for 6 animals in each group

Statistical Significance: $p < 0.001$

Comparison: ^a – as compared with Group I

^b – as compared with Group II

^c – as compared with Group II

Group I - Control Animals

Group II - STZ-induced animals with Diabetic Cataract

Group III - Ethanolic extract of *Cyamopsis tetragonolobus* treated animals

Group IV - Ethanolic extract of *Cyperus rotundus* treated animals

Though, both plants were found to be effective in maintaining the body weight of STZ-induced animals, *Cyperus rotundus* treated animals showed slightly better results when compared with *Cyamopsis tetragonolobus* treated animals. Extract does not exert any side effects or toxic symptoms inferring its non-toxic nature. No death was observed.

3. Organ Weight

Different groups of rats with their organ weight is shown in Table 1.

Table 1
Organ weights of experimental animals

Groups	Eye Weight (gm)	Liver Weight (gm)	Kidney Weight (mg)	Pancreas Weight (mg)	Spleen Weight (mg)	Heart Weight (mg)	Lens Weight (mg)	Brain Weight (mg)
Group I	1.28 \pm 0.01	5.46 \pm 0.02	629 \pm 0.89	275 \pm 0.63	236 \pm 0.89	339 \pm 0.89	729 \pm 1.55	761 \pm 1.10
Group II	1.09 \pm 0.01 ^{a†}	5.24 \pm 0.01 ^{a†}	581 \pm 1.1 ^{a†}	207 \pm 1.79 ^{a†}	214 \pm 1.26 ^{a†}	348 \pm 1.26 ^{a†}	710 \pm 0.63 ^{a†}	734 \pm 0.89 ^{a†}
Group III	1.19 \pm 0.01 ^{b†}	5.43 \pm 0.02 ^{b†}	602 \pm 1.26 ^{b†}	246 \pm 0.89 ^{b†}	250 \pm 0.89 ^{b†}	338 \pm 1.41 ^{b†}	713 \pm 1.26 ^{b†}	740 \pm 1.41 ^{b†}
Group IV	1.18 \pm 0.01 ^{c†}	5.44 \pm 0.02 ^{c†}	610 \pm 1.79 ^{c†}	270 \pm 0.89 ^{c†}	225 \pm 1.10 ^{c†}	340 \pm 1.26 ^{c†}	723 \pm 1.41 ^{c†}	748 \pm 1.41 ^{c†}

Values are expressed as mean \pm SD for 6 animals in each group

Statistical Significance: $p < 0.001$

Comparison: ^a – as compared with Group I

^b – as compared with Group II

^c – as compared with Group II

There was a significant ($p < 0.001$) decrease in weights of vital organs such as eye, liver, kidney, pancreas, spleen, heart, lens and brain of diabetes-induced cataract animals when compared with control animals. The ethanolic extract of *Cyamopsis tetragonolobus* treated animals and *Cyperus rotundus* treated animals were found to have significant increase ($p < 0.001$) in the organ weights when compared with that of animals with diabetes-induced cataract.

4. Biochemical Parameters in Serum

4 (i). Biochemical profile in serum

Table 2 represents the changes of blood glucose, serum glycosylated haemoglobin, insulin, urea, uric acid and creatinine levels in samples of control and experimental animals. The levels of blood glucose, serum glycosylated haemoglobin, urea and uric acid were significantly increased ($p < 0.001$) in Group II animals when compared with Group I animals. In plant treated animals, the levels of glucose,

glycosylated haemoglobin, urea and uric acid were significantly less ($p < 0.001$) when compared to Group II animals. *Cyperus rotundus* treatment decreased the blood glucose levels of animals to a greater extent than when compared to *Cyamopsis tetragonolobus* treatment even after STZ injection. The levels of glycosylated haemoglobin, urea and uric acid were significantly less ($p < 0.001$) in Group III animals and Group IV animals. The levels of serum insulin and creatinine significantly decreased ($p < 0.001$) in Group II animals when compared with Group I animals. The levels of serum insulin and creatinine in both plant treated animals increased significantly ($p < 0.001$). *Cyperus rotundus* treatment was better in increasing the serum insulin levels when compared to that of *Cyamopsis tetragonolobus* treatment. It was noticed that the levels of glycosylated haemoglobin, creatinine, urea and uric acid were higher in *Cyamopsis tetragonolobus* treated plants than with *Cyperus rotundus* treatment.

Table 2

Levels of glucose, glycosylated hemoglobin, insulin, urea, uric acid and creatinine of experimental animals

Groups	Glucose (mg/dl)	Glycosylated Hemoglobin (%)	Insulin (μ U/ml)	Urea (mg/dl)	Uric acid (mg/dl)	Creatinine (mg/dl)
Group I	82.57 \pm 0.10	4.5 \pm 0.22	16.44 \pm 0.03	24.35 \pm 0.04	1.13 \pm 0.01	0.96 \pm 0.01
Group II	476.29 \pm 0.04 ^{a*}	8.4 \pm 0.21 ^{a*}	7.72 \pm 0.04 ^{a*}	38.61 \pm 0.04 ^{a*}	2.51 \pm 0.01 ^{a*}	0.34 \pm 0.01 ^{a*}
Group III	257.80 \pm 0.15 ^{b*}	5.0 \pm 0.63 ^{b*}	12.27 \pm 0.04 ^{b*}	25.48 \pm 0.08 ^{b*}	1.52 \pm 0.01 ^{b*}	1.10 \pm 0.02 ^{b*}
Group IV	132.11 \pm 0.01 ^{c*}	4.1 \pm 0.06 ^{c*}	13.75 \pm 0.05 ^{c*}	25.20 \pm 0.05 ^{c*}	1.47 \pm 0.02 ^{c*}	1.00 \pm 0.13 ^{c*}

Values are expressed as mean \pm SD for 6 animals in each group

Statistical Significance: $p < 0.001$

Comparison: ^a – as compared with Group I

^b – as compared with Group II

^c – as compared with Group III

4 (ii). Protein profile in serum

Table 3 represents the changes of total protein, albumin and globulin levels in serum samples of control and experimental animals.

Table 3

Levels of total protein, albumin and globulin in serum of experimental animals

Groups	Total Protein (g/dl)	Albumin (g/dl)	Globulin (g/dl)
Group I	6.57 \pm 0.04	4.25 \pm 0.03	3.39 \pm 0.05
Group II	5.44 \pm 0.07 ^{a*}	2.64 \pm 0.09 ^{a*}	2.29 \pm 0.08 ^{a*}
Group III	5.70 \pm 0.04 ^{b*}	3.30 \pm 0.07 ^{b*}	3.38 \pm 0.03 ^{b*}
Group IV	6.55 \pm 0.04 ^{c*}	3.39 \pm 0.09 ^{c*}	3.89 \pm 0.06 ^{c*}

Values are expressed as mean \pm SD for 6 animals in each group Statistical Significance: $p < 0.001$

Comparison: ^a – as compared with Group I, ^b – as compared with Group II, ^c – as compared with Group III

The levels of total protein, albumin and globulin were significantly decreased ($p < 0.001$) in Group II animals when compared with Group I animals. In both plant treated animals, the total protein, albumin and globulin levels increased significantly ($p < 0.001$) and reached near normal when compared to that of Group II animals. The levels of total protein, albumin and globulin were significantly more in Group IV animals when compared with that of Group III animals. Thus, *Cyperus rotundus* treatment was better in restoring the total protein, albumin and globulin levels.

4 (iii). Enzymatic markers in serum

Table 4 represents the activities of marker enzymes such as SGOT, SGPT, ALP and LDH in serum samples of control and experimental animals.

Table 4
Activities of enzymatic markers such as SGOT, SGPT, ALP and LDH in serum of experimental animals

Groups	SGOT (IU/L)	SGPT (IU/L)	ALP (IU/L)	LDH (IU/L)
Group I	84.1 ± 0.06	48.30 ± 0.11	153.25 ± 0.14	169.1 ± 0.11
Group II	110.2 ± 0.35 ^{a*}	67.69 ± 0.19 ^{a*}	298.77 ± 0.09 ^{a*}	278.0 ± 1.67 ^{a*}
Group III	93.0 ± 1.90 ^{b*}	34.10 ± 0.06 ^{b*}	232.02 ± 0.51 ^{b*}	185.0 ± 1.10 ^{b*}
Group IV	87.0 ± 2.28 ^{c*}	43.00 ± 1.41 ^{c*}	245.37 ± 0.02 ^{c*}	173.0 ± 2.10 ^{c*}

Values are expressed as mean ± SD for 6 animals in each group

Statistical Significance: $p < 0.001$

Comparison: ^a – as compared with Group I
^b – as compared with Group II
^c – as compared with Group II

The activities of SGOT, SGPT, ALP and LDH increased significantly ($p < 0.001$) in Group II animals when compared to that of Group I animals. The activities of these marker enzymes decreased significantly ($p < 0.001$) in both plant treated animals as compared to Group II animals.

4 (iv). Lipid profile in serum

Table 5 represents the changes in lipid profile in serum samples of control and experimental animals.

Table 5
Changes in lipid profile in serum of experimental animals

Groups	Free fatty acid (mg/dl)	Triglyceride (mg/dl)	Total Cholesterol (mg/dl)	LDL Cholesterol (mg/dl)	HDL Cholesterol (mg/dl)	VLDL Cholesterol (mg/dl)
Group I	84.71 ± 0.01	69.1 ± 0.06	53.58 ± 0.05	29.88 ± 0.05	40.5 ± 0.01	16.41 ± 0.01
Group II	193.63 ± 0.02 ^{a*}	125.2 ± 0.14 ^{a*}	94.49 ± 0.03 ^{a*}	85.10 ± 0.09 ^{a*}	30.2 ± 0.06 ^{a*}	26.64 ± 0.02 ^{a*}
Group III	107.19 ± 0.01 ^{b*}	105.0 ± 1.67 ^{b*}	67.24 ± 0.03 ^{b*}	40.00 ± 2.83 ^{b*}	33.1 ± 0.09 ^{b*}	21.79 ± 0.03 ^{b*}
Group IV	97.07 ± 0.03 ^{c*}	90.0 ± 2.68 ^{c*}	58.30 ± 0.04 ^{c*}	35.00 ± 2.00 ^{c*}	38.4 ± 0.20 ^{c*}	20.29 ± 0.02 ^{c*}

Values are expressed as mean ± SD for 6 animals in each group

Statistical Significance: $p < 0.001$

Comparison: ^a – as compared with Group I
^b – as compared with Group II
^c – as compared with Group II

The levels of free fatty acids, total cholesterol, triglyceride, LDL-cholesterol and VLDL-cholesterol significantly increased ($p < 0.001$) in Group II animals, while HDL-cholesterol level decreased significantly ($p < 0.001$) when compared to Group I animals. The levels of free fatty acids, total cholesterol, triglyceride, LDL-cholesterol and VLDL-cholesterol lowered significantly ($p < 0.001$) in Group III and Group IV animals when compared to that of Group II animals. The levels of HDL-cholesterol rose significantly ($p < 0.001$) in both plant treated animals compared to Group II animals. From the results, it should be noticed that *Cyperus rotundus* treatment was fairly better in altering the lipid profile of experimental rats when compared with that of *Cyamopsis tetragonolobus* treatment.

DISCUSSION

Oral administration of ethanol extracts of *Cyperus rotundus* rhizomes did not lead to any changes in general behavior, mortality, weight gain, hematological and clinical blood chemistry parameters. Hence, these extracts did not cause oral acute (single dose of 5000 mg/kg) and sub-acute toxicity (repeated oral dose of 1000 mg/kg) in rats²⁷. Thus, oral administration of ethanol extracts of *Cyperus rotundus* rhizomes can be considered safer. Hydro-alcoholic extract of *Cyamopsis tetragonolobus* possess hepato-protective activity against CCl₄-induced hepatotoxicity in rats and suggested that the effect is preventive in nature at oral dose of 250 mg/kg and 500 mg/kg²⁸. Oral administration (dose of 500mg/kg body weight) of an ethanolic extract of *Cyamopsis tetragonoloba* (Guar) produced significant anti-ulcer, anti-secretory and cytoprotective effect in rats²⁹. It is well-known that diabetes is very closely associated with weight loss. The body weights of diabetic cataract rats are lowered³⁰. The same has been confirmed in a rat model during evaluation of early diabetes-induced biochemical changes in retina³¹. The body weight of STZ-induced diabetic rats improved on treatment with varied medicinal plants similar to our results. The oral feeding of water extract of fresh leaves of *Azadirachta indica* improved body weight of STZ-induced diabetic rats and reversed diabetic retinopathy³². Similarly, from our study also, it is observed that the treatment with both the plants prevented the loss of body weight induced by STZ. The relative weights of kidney and liver increased in STZ-induced diabetic albino rats, while the weight of pancreas remained unaffected³³. The weight of the pancreas and

kidney increased in STZ-diabetic rats, but treatment with fermented red ginseng seemed to decrease the overall weights with no significant difference in spleen weight of diseased and treated rats³⁴. The relative organ weights of brain, liver, lungs, heart, kidney and spleen of *Lepidium sativum* seed powder treated Wistar rats showed comparable results with that of the normal rats³⁵. The lens of diabetic cataract rats decreased, while treatment with *Trigonella* (Fabaceae) and vanadate resumed the lens weight to that of normal rats³⁰. Our study also indicates the beneficial aspects of administering *Cyamopsis tetragonolobus* and *Cyperus rotundus* to prevent loss of body weight or organ weight that occur due to diabetic complications.

The aqueous extract of leaves and flowers of *Clitoria ternatea* (Fabaceae) exerted a hypoglycaemic effect in alloxan-induced diabetic rats as evidenced by reduction in serum glucose, glycated haemoglobin along with increased serum insulin³⁶. Oral administration of *Annona squamosa* aqueous extract to diabetic rats significantly reduced blood glucose, urea, uric acid and creatinine³⁷. The ethanol extract of *Chromolaena odorata* leaves reduced glucose, lipid profile and improved glucose and insulin tolerance as well as serum insulin and HDL-cholesterol levels when administered to STZ-induced diabetes and cataract in rats³⁸. The aqueous root extract of *Pseudarthria viscida* (Fabaceae) restored the levels of protein and albumin in STZ-diabetic rats³⁹. The levels of protein reduced in diabetic rats while treatment with *Annona squamosa* increased the protein levels and reached near normal. The levels of albumin and albumin/globulin ratio also decreased in diabetic animals while treatment restored these levels significantly³⁷. Thus, in our study the reduction in protein profiles induced by STZ is restored by treatment with both the plants. The activity of ALP and aminotransferases ALT and AST lowered in STZ-induced diabetic rats while treatment of *Aspalathus linearis* (rooibos tea) restored their activities⁴⁰. The increase in serum lactate dehydrogenase activity in streptozotocin-induced diabetic rats is reversed by aqueous extract of *Embelia ribes* Burm⁴¹. The ethanolic extracts of *Rheum emodi* rhizome treated for 30 days restored the activities of marker enzymes such as ALT, AST, ALP and LDH in serum to near control levels which rose due to induction of diabetes by alloxan in rats⁴². Hence, it can be noticed that the enzymatic marker levels were increased by STZ in diabetes-induced groups. But the plant treated groups

retained their enzyme activities despite induction of diabetes during the experimental period. Treatment with *Tinospora cordifolia* extract decreased serum total cholesterol, LDL-cholesterol and triglyceride significantly, while increase in serum HDL-cholesterol levels was observed. The ethanolic leaf extract of *Mucuna pruriens* (Fabaceae) improved the lipid profile of alloxan-induced diabetic Wistar rats⁴³. Dietary supplementation with *Cyperus esculentus* (Cyperaceae) significantly improved the altered biochemical parameters such as changes in

serum total lipids, total cholesterol, triglycerides, LDL-cholesterol and HDL-cholesterol levels in STZ-diabetic rats after four weeks of treatment⁴⁴. The elevated levels of total cholesterol, triglyceride and LDL-cholesterol and a decrease in HDL-cholesterol levels in STZ-induced diabetic rats are modified as better lipid profile by administering aqueous extract perfusion of *Rosmarinus officinalis*⁴⁵. Similarly, ethanol extract of *Wattakaka volubilis* possess significant antihyperlipidemic effect in alloxan-induced diabetic rats⁴⁶.

CONCLUSION

From the above results, it could be inferred that the biochemical changes signify that the ethanolic extract of *Cyamopsis tetragonoloba* and *Cyperus rotundus* has definite anti-cataract effect against STZ-induced diabetic cataract by delaying the cataractogenesis.

ACKNOWLEDGEMENT

We would like to thank Dr. R. Selvaraj (Assistant Professor) for his help in carrying out the animal experiments in Biomedical Research and Lab Animal Centre (BRULAC) in Saveetha University, Chennai, Tamil Nadu, India.

CONFLICT OF INTEREST

Conflict of interest declared none

REFERENCES

1. Klein R and Klein BEK, Diabetic eye disease, *The Lancet*, 350(9072): 197-204, (1997).
2. Crawford JS, Conservative management of cataracts, *International Ophthalmology Clinics*, 17(4): 31-35, (1977).
3. Mankil Jung, Moonsoo Park, Hyun Chul Lee, Yoon-Ho Kang, Eun Seok Kang and Sang Ki Kim, Antidiabetic agents from medicinal plants, *Current Medicinal Chemistry*, 13(10): 1203-1218, (2006).
4. Overbeeke N, Termorshuizen GH, Giuseppin ML, Underwood DR and Verrips CT, Secretion of the alpha-galactosidase from *Cyamopsis tetragonoloba* (guar) by *Bacillus subtilis*, *Applied and Environmental Microbiology*, 56(5): 1429-1434, (1990).
5. Khare CP, *Indian herbal remedies: rational Western therapy, ayurvedic and other traditional usage*, botany, Springer, Berlin & New York: 171-172, (2004).
6. Holm LeRoy G, Plucknett Donald L, Pancho Juan V and James P Herberger, *World's worst weeds: distribution and biology*, Honolulu, University Press of Hawaii: p. 609, (1977).
7. Asif Bin Rehman, Pharmacological studies on traditional medicine (*Cyperus rotundus*) used in Pakistan, <http://eprints.hec.gov.pk/2672/>, (2008).
8. Ramya P, Seema S and Vijayalakshmi K, Impact of hydroalcoholic extract of *Cyperus rotundus* on glucose induced cataract – An *in vitro* study, *International Journal of Pharmacy and Biological Sciences*, 2(4): 320-331, (2012).
9. Gajdosik A, Gajdosikova A, Stefek M, Navarova J and Hozova R, Streptozotocin-induced experimental diabetes in male Wistar rats, *General Physiology Biophysics*, 18 Spec No: 54-62, (1999).
10. Sasaki T, Matsy S and Sonae A, Effect of acetic acid concentration on the color reaction in the O-toluidine boric acid method for blood glucose estimation, *Rinshobokagaku*, 1: 346-353, (1972).

11. Wang J and Yang G, Colorimetric detection of glycosylated hemoglobin, Hunan Yixeyuan Xuebao, 7(3): 325-328, (1982).
12. Okutucu B, Diner A, Habib O and Zihnioglu F, Comparison of five methods for determination of total plasma protein concentration, Journal of Biochemical and Biophysical Methods, 70(5): 709-711, (2007).
13. Doumas BT, Watson WA and Biggs HG, Albumin standards and the measurement of serum albumin with bromocresol green, Clinica Chimica Acta, 258(1): 21-30, (1997).
14. Natelson S, Scott ML and Beffa C, A rapid method for the estimation of urea in biological fluids, American Journal of Clinical Pathology, 21(3): 275-281, (1951).
15. Caraway WT, In: D. Seligron (Ed.), Standard Methods of Clinical Chemistry, Academic Press, New York: Vol.4: p.239, (1963).
16. Owen JA, Iggo B, Scandrett FJ and Stewart CP, The determination of creatinine in plasma or serum and in urine; a critical examination, Biochemical Journal, 58(3): 426-437, (1954).
17. Folch J, Lees M and Sloane Stanley GH, A simple method for the isolation and purification of total lipids from animal tissues, Journal of Biological Chemistry, 336(1): 497-509, (1957).
18. Parekh AC and Jung DH, Cholesterol determination with ferric acetate-uranium acetate and sulphuric acid-ferrous sulphate reagent, Analytical Chemistry, 42(12): 1423-1427, (1970).
19. Rice EW, In: Triglycerides ("neutral fats") in serum, Standard Methods of Clinical Chemistry, Roderick M P, editor, Vol.6. New York: Academic Press, p.215-22, (1970).
20. Hron WT and Menahan LA, A sensitive method for the determination of free fatty acids in plasma, Journal of Lipid Research, 22(2): 377-381, (1981).
21. Itaya K, A more sensitive and stable calorimetric determination of free fatty acids in blood, Journal of Lipid Research, 18(5): 663-665, (1977).
22. Wilson DE and Spiger MJ, A dual precipitation method of quantitative plasma lipoprotein measurement without ultracentrifugation, Journal of Laboratory Clinical Medicine, 82(3): 473-482, (1973).
23. Warnick GR and Albers JJ, A comprehensive evaluation of the heparin-manganese precipitation procedure for estimating high density lipoprotein cholesterol, Journal of Lipid Research, 19(1): 65-76, (1978).
24. Mohun AF and Cook IJY, Simple methods for measuring serum levels of the glutamic-oxalacetic and glutamic-pyruvic transaminases in routine laboratories, Journal of Clinical Pathology, 10(4): 394-399, (1957).
25. King EJ and Armstrong AR, A convenient method for determination of serum and bile phosphatase activity, Canadian Medical Association Journal, 31(4): 376-381, (1934).
26. King J, The dehydrogenase or oxidoreductase lactate dehydrogenase, In: Van. D. (Ed.), Practical clinical enzymology, Nostrand Co. Ltd., London. 191-2008, (1965).
27. Thanabhorn S, Jaijoy K, Thamaree S, Ingkanin K and Panthong A, Acute and subacute toxicities of the ethanol extract from the rhizomes of *Cyperus rotundus* Linn, Mahidol University Journal of Pharmaceutical Sciences, 32(1-2): 15-22, (2005).
28. Sujeet singh, Dinesh Mali, Yadunath M Joshi and Kadam VJ, Hepatoprotective effect of *Cyamopsis tetragonolobus* Linn. On carbon-tetrachloride (CCl₄)-induced acute hepatotoxicity on Albino wistar rats, Journal of Pharmacy Research, 4(3): 789, (2011).
29. Rafatullah S, Alyahya MA, Al-Said MS, Abdul Hameed Taragan KU and Mossa JS, Gastric anti-ulcer and cytoprotective effects of *Cyamopsis tetragonoloba* ('Guar') in rats, International Journal of Pharmacognosy, 32(2): 163-170, (1994).
30. Anju Preet, Bihari L Gupta, Pramod K Yadava and Najma Z Baquer, Efficacy of lower doses of vanadium in restoring altered glucose metabolism and antioxidant status in diabetic rat lenses, Journal of Biosciences, 30(2): 221-230, (2005).
31. Obrosova IG, Drel VR, Kumagai AK, Szabo C, Pacher P and Stevens MJ, Early diabetes-induced biochemical changes in the retina: comparison of rat and mouse models, Diabetologia, 49(10): 2525-2533, (2006).
32. Halim Eshrat M. Ali Hussain, Reversal of diabetic retinopathy in streptozotocin-induced diabetic rats using traditional Indian anti-diabetic plant, *Azadirachta indica* (L.), Indian Journal of Clinical Biochemistry, 17(2): 115-123, (2002).
33. Muhammad Zafar and Syed Naeem-ul-Hassan Naqvi, Effects of STZ-induced diabetes on the relative weights of kidney, liver and pancreas in Albino rats: a comparative study, International Journal of Morphology, 28(1): 135-142, (2010).

34. Hyun-Jeong Kim, In-Gyeong Chae, Sung-Gyu Lee, Hyun-Jin Jeong, Eun-Ju Lee and In-Seon Lee, Effect of fermented red ginseng extracts on hyperglycemia in streptozotocin-induced diabetic rats, *Journal of ginseng research*, 34(2): 101-112 (2010).
35. Datta PK, Diwakar BT, Viswanatha S, Murthy KN, Naidu KA, Safety evaluation studies on Garden cress (*Lepidium sativum* L.) seeds in Wistar rats, *International Journal of Applied Research in Natural Products*, 4(1): 37-43 (2011).
36. Daisy P and Rajathi M, Hypoglycemic effects of *Clitoria ternatea* Linn. (Fabaceae) in alloxan-induced diabetes in rats, *Tropical Journal of Pharmaceutical Research*, 8(5): 393-398, (2009).
37. Kaleem M, Medha P, Ahmed QU, Asif M and Bano B, Beneficial effects of *Annona squamosa* extract in streptozotocin-induced diabetic rat, *Singapore Medical Journal*, 49(10): 800-804 (2008).
38. Onkaramurthy M, Veerapur VP, Thippeswamy BS, Madhusudana Reddy TN, Hunasagi Rayappa, and Badami S, Anti-diabetic and anti-cataract effects of *Chromolaena odorata* Linn. in streptozotocin-induced diabetic rats, *Journal of Ethnopharmacology*, 145(1): 363-372 (2013).
39. Rajendran Kuppusamy, Annie Shirwaikar, Kishore G Sam and Srinivasan K Kaitheri, Antidiabetic activity of *Pseuderthria viscida* aqueous root extract in neonatal streptozotocin-induced NIDDM rats, *Revista Brasileira de Farmacognosia Brazilian Journal of Pharmacognosy*, 22(5): 1079-1084 (2012).
40. Ulicna O, Vancova O, Bozek P, Carsky J, Sebekova K, Boor P, Nakano M and Greksak M, Roobos Tea (*Aspalathus linearis*) partially prevents oxidative stress in streptozotocin-induced diabetic rats, *Physiol. Res.*, 55(2): 157-164 (2006).
41. Uma Bhandari and Nazam Ansari M, Antihyperglycaemic activity of aqueous extract of *Embelia ribes* Burm in streptozotocin-induced diabetic rats. *Indian Journal of Experimental Biology*, 46(8): 607-613 (2008).
42. Radhika R, Ragavan B, Sharad Pawar D and Sudarsanam D, Action of marker enzymes of *Rheum emodi* in alloxan-induced diabetic rats, *Asian J Exp Biol Sci.*, 3(2): 420-423 (2012).
43. Eze ED, Mohammed A, Musa KY, Tanko Y and Isa AS, Effect of ethanolic leaf extract of *Mucuna pruriens* (Fabaceae) on lipid profile in alloxan-induced diabetic Wistar rats, *British Journal of Pharmacology and Toxicology*, 3(3): 102-109 (2012).
44. Hassan HA, Effect of dietary supplementation with tigernut tubers on streptozotocin-induced diabetic rats, *The Egyptian Journal of Hospital Medicine*, 29: 475-485 (2007).
45. Abdul-Rahim Al-Jamal and Taha Alqadi, Effects of rosemary (*Rosmarinus officinalis*) on lipid profile of diabetic rats, *Jordan Journal of Biological Sciences*, 4(4): 199-204 (2011).
46. Maruthupandian A, Mohan VR and Sampathraj R, Antidiabetic, antihyperlipidaemic and antioxidant activity of *Wattakaka volubilis* (L. F) Stapf leaves in alloxan-induced diabetic rats, *International Journal of Pharmaceutical Sciences and Research*, 1(11): 83-90 (2010).