



## HEPATOPROTECTIVE ACTIVITY OF PLANT BASED DRUGS AGAINST D-GALN.

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### ABSTRACT

Liver disorder may cause liver inflammation or tissue injury and affects liver physiologic condition. Natural products that are found in the form of vegetables, fruits, plant extract, herbs, and animals, have been traditionally and clinically used for treating liver disorders. They are specific chemical compounds that usually have biological activities for use in drug evaluation and design. Many herbal products have been clinically available as potent anti-hepatotoxic agents against commonly occurring liver disorders. This review summarizes the current progress in the basic, clinical, and traditional research on herbal products in treatment of various liver disorders. Also, we will rivet on the discovery and biological evaluation of the herbal products, which shows potential as a new anti-hepatotoxic agent of liver disorders.

**KEYWORDS:** Antioxidant enzyme, Hepatotoxicity, D-Galactosamine, Flavanoide, Free radical, Hepatoprotective activity, Herbal medicine, Ant inflammation.



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## INTRODUCTION

A liver in injury is closely related to human hepatitis; D-Galactosamine is well defined hepatotoxicant & plays an important role in evolution of hepatoprotection. D Galn hepatotoxicant is an experimental model which mainly causes acute hepatitis & it not affect other organs. D-Galn has a greater affinity with liver because liver cell have high affinity with galactokinase & galactose 1-uridytransferase&also it inhibit synthesis of uridylylate nucleotide<sup>1</sup>. D Galn induced hepatotoxicity by inhibiting the synthesis of RNA & protein via reduction of cellular U.T.P a substrate for RNA polymerase<sup>2</sup> D-Galn breaks the enzyme that transfers the substrate to mitochondria & modifies the composition of phospholipids of membrane this bring about change in mitochondria also<sup>3</sup>. A single injection with d-galactosamine can decrease the uracil nucleotides in the liver and heart.<sup>4</sup> It alter the synthesis of essential uridylylate nucleotide, depletion of these nucleotide impaired the synthesis of glycoprotein lead to change of cell membrane resulting change in permeability of membrane & finally leakage of enzyme from cell<sup>5</sup>.It cause metabolic disturbance. The main source of damaging cell membrane & stimulation of lipid peroxidase is due to formation of highly reactive hydroxyl radical (OH\*)<sup>6</sup>. Oxidative stress is one major problem of D Galn induction due to excessive formation of free radical resulting from oxidative stress damage macromolecule as lipid & D-Gal injection decreased GSH, SOD,& CAT who show that that D Galn intoxicant increased TBRS level which are a typical parameter for lipid peroxidase<sup>7</sup> Oxidation is common procedure of body, free radical are formed when body breakdown food into energy it is highly unstable molecule that alter many vital function of body. Free radical causes damage known as "oxidative stress" & key role in production of state like alzimer, cancer, heart disease, Parkinson, rheumatoid arthritis<sup>8</sup>

### **HEDYOTIS CORYMBOSA**

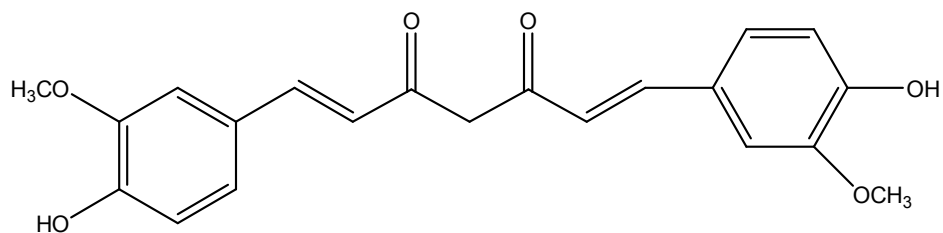
Hedyotis corymbosa (L.) also known as Oldenlandia corymbosa Lam, belonging to family Rubiaceae, is a weed which distributed throughout India. In traditional medicine of Kerala It is commonly known as 'Parppatakapullu'. There are approx 180 species in which 35 species found in Malaysia. Plant extract given in liver deasese, jaundice, heat eruptions, and vitiated conditions. Increased level of AST, ALT, ALP and GT in serum Caused by D-Galn alters by the treatment with HC extract at 100 mg & 200 mg/kg does. Recovery getting normalisation that extract of HC causes parenchyma cell regeneration in liver thus protecting membrane fragility, decreasing enzyme leakage and extract mediated decreased level of increased bilirubin. Suggested HC extract able to stabilize billiary dysfunction. D-Galn treatment causes decrease in level of antioxidative enzyme activities. Extract of HC arises the antioxidant parameter and decreased oxidative stress which appeared in decreasing the mean value of liver TBARS and compared to D-Galn. The level of GSH, SOD & CAT decreased out the hepatic damage with D-Galn administration but treatment with 100 & 200 mg/kg of treatment show significant increase in these enzymes and controls the level of LPO which indicate antioxidant activity of HC. Histopathology examination of HCE at Different Dose levels offers hepatoprotection, but 200mg/kg is more effective than other lower dose.

### **CURCUMIN**

Curcumine is main compound found in plant curcuma longa was first isolated two centuries ago. It is used from long time in Indian tradition to treat various disorders like body ache, skin diseases rheumatism, intestinal worms, diarrhea, hepatic disorders, biliousness, urinary discharges, intermittent fevers, leukoderma, amenorrhoea, and colic. Viewing its medicinal importance, interest has been focused on curcumine due to its treatment of wide variety of disease such as Alzheimer's disease,

psoriasis, cardiovascular, arthritis, cancer, diabetes & so many by modulation of various

molecular target<sup>9</sup>. Curcumin inhibit many factors like nuclear factor-kappa.



Chemical structure of various natural products: Curcumin from the plant Curcuma Longa.

Curcumin control many pro-inflammatory and profibrotic cytokines and its antioxidant properties proved a rational use in liver disease. Curcumin could attenuate liver injury induced by ethanol, thioacetamide, iron overdose, cholestasis<sup>10</sup>

### **ENICOSTEMMA AXILLARE**

*Enicostemma axillare* Raynal Belonging from Family Gentianaceae is a perennial glabrous herb found throughout India common in coastal areas. In Indian tradition it is used in the treatment of hepatic diseases and a blood purifier. Swertiamarin is a main content found in *Enicostemma axillare* in the form of alkaloids, steroids, saponins, triterpenoids, flavanoids, phenolic acids and xanthenes were isolated from *Enicostemma axillare*. Swertiamarin Treatment at 100 and 200 mg/kg show a significant reversed the level of liver enzyme, produced by D-Galn & recover towards normalize. Extract of *Enicostemma* caused increase level in CAT, SOD, & GSH & Minimize the level of TBARS in serum, kidney and liver show a significant role that it restored these antioxidant action in the problem caused by D-Galn hence the mechanism of hepatoprotection of Swertiamarin is mainly by its antioxidant property. Swertiamarin is one of the treatments at 200 mg/kg body wt administrated orally for 8 day to normalize animal and also show better hepatoprotective action<sup>11</sup>. This result given by histological studies. Extracted Swertiamarin is 100% pure, which can be evaluated by HPTLC. In Acute toxicity evaluation Swertiamarin up to 2000 mg/kg show no any toxicity & mortality when

given orally. Swertiamarin (200 mg/kg) dose to d-Galn intoxicated rats showed a maximum hepatoprotection.

### **GLYCYRRHIZIN**

Glycyrrhizin is an aqueous extract of the liquorice root (*Glycyrrhiza glabra*) and used from ancient time in traditional medicine to treat bronchitis, jaundice and gastritis. The main chemical constituents are flavanoids, hydroxycoumarins, glycyrrhetic acid, and beta-sitosterol, with some extent with glucocorticoid and mineral corticoid properties. A standard extract containing, cysteine and glycine, glycyrrhizin, used to treat chronic hepatitis as Stronger Neominophagen C (SNMC) in Japan. Daily treatment with Glycyrrhizin with 80 mg/kg/day administration iv for 2 week lowered the elevated level of alanine transaminase<sup>12</sup>. SNMC mainly available in the united state as tablet & liquid powder. In cell culture experiments, glycyrrhizin modifies glycosylation and blocks sialylation of hepatitis B surface antigen (HBsAg), leading to its retention in the trans-Golgi apparatus<sup>13</sup>. There is no any experimental evidence for antiviral activity of Glycyrrhizin, but it is due to membrane-stabilizing effect proposed by in vitro hepatotoxicity model study<sup>15</sup>.

### **MELOCHIA CORCHORIFOLIA**

The extract of Aerial part of plant *Melochia Corchorifolia* shows presence of different phytochemicals constituents like steroid, flavanoids, alkaloid, tannins, and carbohydrates. The extract gives negative due to its potent pharmacological activity, economic

viability and toxicity. The main antioxidant property is due to its reflecting electron donation capacity of bioactive compound<sup>15</sup>. By donating electrons, antioxidant substances are able to block radical chain reaction by converting reactive oxygen species to more stable products. The antioxidant properties of plant that promote health protection by counteracting reactive oxygen species (ROS)<sup>16</sup>. The phenolic content in Arial part of *M.*

*corchorifolia* were found to be from (16.28+<sub>-</sub>0.52) to (34.22+<sub>-</sub>0.43) mg/g. Methanol extract contains more phenolic content i.e. (34.22+<sub>-</sub>0.43) mg/g than other extracts and the alkaloid content was ranging from (18.46+<sub>-</sub>0.34) to (26.37+<sub>-</sub>0.16) mg/g. Also methanolic content have more alkaloid content (26.37+<sub>-</sub>0.16) mg/g than other extracts<sup>17</sup>.

S.no.	Name of the extract	Total phenolic content (mg/g)	Total alkaloid content (mg/g)
1	Hydro alcoholic (Ethanol70%)	29.34±0.69	21.86±0.39
2	Ethyl acetate	24.69±0.47	20.58±0.43
3	Hexane	16.28±0.52	18.46±0.34
4	Methanol	34.22±0.43	26.37±0.16

On comparing with the silymarin plant extract at 500mg/kg produce normalize the level of sGOT, sGPT, ALP and TBRS level. The main protective effect is arises due to stabilizing of membrane of liver and scavenging the free radical or by both mechanism. Methanolic extract of *M. corchorifolia* show better antioxidant and hepatoprotective properties due to the presence of important antioxidative factors like phenolic, alkaloid and flavanoids compounds<sup>18-24</sup>.

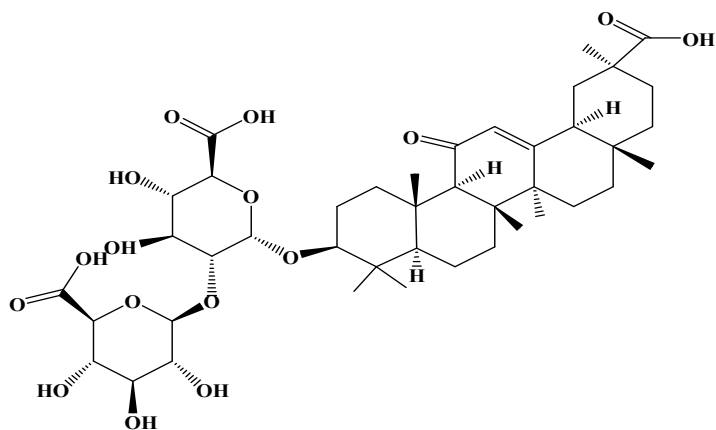
#### **IN VITRO ANTI OXIDANT ACTIVITY**

In this studied the hydro alcoholic, methanol, ethyl acetate extract of *M.corchorifolia* were found concentration dependent scavenging activity on tested free radicals (superoxide, hydroxyl and DPPH). The mean IC50 values for superoxide radical of hydro alcoholic, methanol, ethyl acetate and hexane extracts of *M. corchorifolia* were found to be 206 µg, 127µg,

530µg and 901 µg respectively. The mean IC50 values for hydroxyl radical of methanol extract is 240µg, 384µg for hydroalcoholic, ethyl acetate was found to be 490µg and 501µg respectively. The IC50 value for DPPH radical of methanol, hydro alcoholic, ethyl acetate and hexane was found to be 179µg, 286µg, and 470µg and 971µg respectively<sup>8</sup>.

#### **POLYGONUM CUSPIDATUM**

The main chemical constituents of *Polygonum cuspidatum* is resveratrol, a polyphenolic phyto chemicals mainly found in grapes wine and berries, several study show that its potent treatment in the liver ailments. The hepatoprotection of *Resveratrol* is due to inflammatory cytokines and free radicals induce antioxidant enzyme and increase the level of Glutathione and modulate varied signal transduction pathway of hepatotoxicity.

Chemical structure of Resveratrol from *Polygonum cuspidatum*

Many studies such as cultured cell, purified enzymes and laboratory animal show that its antioxidant, anti-inflammatory, anti-carcinogenic and anti-aging properties that might be relevant to chronic diseases and/or longevity in humans<sup>25</sup>. Resveratrol reduces the elevated level of TNF- $\alpha$  and IL-6, mRNA and normalizes the number of kuffer cells in the damaged liver. It also promotes liver cell regenerations and minimizes fibrosis that enhances the survival of BDL mice. Resveratrol is also used in cholestasis liver injury<sup>26</sup>. Another hepatoprotection of this plant derived by suppressing oxidative stress and apoptosis. Either pretreatment or advanced stage of hepatocarcinogenesis is treatment show equally effective by activation of the apoptotic pathway in rodent<sup>27</sup>. The activity of Resveratrol on vascular endothelial growth factor and angiogenesis in hepatocellular carcinoma caused by suppression of the activation of NF- $\kappa$ B in HepG2 cell<sup>28</sup>. It also protects liver from methotrexate liver injury<sup>29</sup>. Oral treatment with Resveratrol (20 mg/kg daily for 4 weeks) remarkably inhibits the DMN-induced loss in liver, body weight, and inhibits the elevated level of serum alanine transaminase, aspartate transaminase, alkaline phosphatase and bilirubin levels<sup>30</sup>.

### ***Bei Chaihu (Bupleurum Chinese)***

D-GalN is an important hepatotoxicant producing hepatic injury model. It causes inhibition of RNA & protein synthesis through loss of uridine nucleotide and aggregation of UDP hexosamines in liver cells. In a study it was found that D-GalN induced hepatotoxicity increases sensitivity to TNF which causes hepatic injury during inflammation. Aminotransferase catalysis of amino transfer reaction is a major diagnosis of hepatotoxicity. Serum AST, ALT, and TNF in blood are main points for evaluation of hepatoprotection. In estimation of the hepatoprotective effect of B. Chinese polysaccharide rat's treated with different concentrations of WBCP after D-GalN injection. As in table D-GalN significantly increased serum AST and ALT levels compared with control group, and treatment with WBCP decreases this elevated level in a dose-dependent manner. When treated with WBCP, AST levels were found to be decreased by 11.9%, 33.4% and 40.3% at the different doses of 100, 200 and 400 mg/kg, respectively, and ALT activity by 8.0%, 19.4% and 30.6%, and LDH activity by 14.9%, 26.9% and 37.9%, respectively in a dose-dependent manner. ALP is a main agent for removing phosphate groups from nucleotides and proteins which are formed in the liver and play a key role in hepatic function's. D-GalN increased the level of ALP<sup>31</sup>.

Group	Dose (µmol/mg protein)	AST (µmol/mg protein)	ALT (µmol/mg protein)	ALP (µmol/mg protein)	LDH (µmol/mg protein)
Normal		107.44±9.25	74.41±7.44	501.23±69.42	710.61±69.72
Control		932.23±86.46	635.21±79.32	821.59±110.55	1929.45±242.46
WBCP	100	821.34±77.52	584.44±95.24	753.43±99.19	1641.48±196.42
	200	621.03±92.42*	512.74±63.48*	671.56±90.26*	1411.53±241.28
	400	556.22±86.42**	441.06±84.41**	592.46±94.99**	1197.46±243.98**

Values are expressed as mean ±SD (n = 10).

\* p < 0.05.

\*\* p < 0.01 vs. control

### ***Hedyotis corymbosa***

The effect on various doses of HCE was studied on serum marker enzymes and total bilirubin in D-Galn intoxicated animal. It showed no any toxicological effect at 100 and 200 mg/kg body wt. therefore 200 mg/kg dose of plant extract is effective for its pharmacological effect's D-Galn injury cause significantly change in marker enzyme like, AST - 172.35%, ALT by 188.04%, ALP by 119.9%, γ-GT by 112.33% and total bilirubin by 57.73% on comparing with control group. The percentage protection in marker enzyme of treated group at 100 mg/kg as AST 7.83 (P<

0.01), ALT 12.47 (P< 0.05), ALP 8.4 (P< 0.01), γ-GT 7.03 and total bilirubin 10.67(P<0.01) on comparing with toxic group while maximum protection percentage in marker enzyme at the dose of 200 mg/kg and silymarin (100mg/kg) as AST 38.38 (P< 0.001), 56.34 (P< 0.001),ALT 32.65 (P< 0.001), 46.76 (P< 0.001), ALP 36.93 (P< 0.001), 45.94 (P< 0.001), γ-GT 26.3 (P< 0.01), 38.53 (P<.001), and total bilirubin 24.27 (P< 0.001), 32.03 (P< 0.001) which is almost comparable to the group treated with silymarin. Silymarin is a potent hepatoprotective which used as standard drug<sup>32</sup>.

**Table**

***Effect of HCE on serum AST (U/L), ALT (U/L), ALP (U/L), Total Bilirubin (TBL) level (mg/dl), and γ-GT (U/L) against D-galactosamine induced liver.***

Group	ALT	AST	ALP	TBL	γ-GT
Control	32.38 ±2.7	72.2 ±3.2	98.27 ± 2.2	0.67 ± 0.02	1.54 ± 0.09
D-Galn	93.27 ± 4.6*	213.20± 5.24*	216.1 ± 5.8*	1.03 ± 0.04*	3.27 ± 0.23*
HCE100	81.67 ± 4.2a	196.50 ± 4.2b	197.8 ± 4.1b	0.92 ± 0.03b	3.04 ± 0.2n
HCE 200	62.81 ± 3.8c	131.35 ± 3.8c	136.28 ± 3.2c	0.78 ± 0.02c	2.41 ± 0.19b
Silymarin	49.65 ± 3.1c	93.08 ± 3.5c	116.81 ± 2.9c	0.7 ± 0.02c	2.01± 0.17c

Mean ± S.E.M. of 6 rats in each group

n: no significant.

P values: \*<0.001 compared with respective control group me.

P values: a<0.05, b<0.01, c<0.001 compared with group II (D-Gal).

### ***Allium sativum***

Allicin is main active constitute of freshly *Allium sativum* (garlic) cloves. In D-Galn induced hepatotoxicity in rat resulting increase in level of lipid per oxidation and deacresd antioxidant property of liver. Treatment with allicin prevents these alterations. The concentration of sGOT level decreased that found in liver cell, pancreas, lungs, leukocytes, skeletal muscle, kidneys, brain, and erythrocytes. In tissue sGOT occurs in two locations, the cytosol and mitochondria but sGPT show highest concentration in liver cell.<sup>33</sup> sGPT show more

specific test for assessing hepatocellular damage then test of sGOT. On comparing both sGOT as well as sGPT level that assessed the hepatic damage caused by D-Galn. Hepatotoxicant (D-Galn) gp B compared with Control gp A, Through treatment gp, with the allicin 0.3 and 0.4 mg/kg for 15 and 22 days respectively show highest efficacy and potency in decreasing the sGOT and sGPT levels, the dose with low concentration and less number of days (0.3 mg/kg for 15 days) showing better protection for the liver against d-Galn induced hepatotoxicity as shown in table.

**Effect of different doses of allicin and the reference drug silymarin on sGOT and sGPT activities in d-Galn-treated rats.**

Group	sGOT			sGPT		
	8day	15day	22day	8day	15day	22day
Control (A)	110.6 ± 3.1	107.5 ± 3.4	109.0 ± 3.9	60.0 ± 1.8	58.2 ± 1.7	61.7 ± 2.1
GalN-intoxicated (B)	340.6 ± 25.4 □□	329.0 ± 23.2 □□	333.2 ± 26.4 □□	215.8 ± 22.1 □□	199.4 ± 21.6 □□	204.4 ± 24.2 □□
Silymarin (C)	140.3 ± 16.5 □	129.2 ± 9.3 □	136.1 ± 8.7 □	102.5 ± 8.3 □	59.1 ± 2.2 □	58.7 ± 1.7 □
Allicin, 0.2 mg/kg (D)	163.0 ± 9.3 □□	149.0 ± 8.5 □□	141.0 ± 7.6 □□	143.2 ± 6.3 □□	74.4 ± 6.2 □□	68.2 ± 2.3 □□
Allicin, 0.3 mg/kg (E)	151.5 ± 7.2 □	134.0 ± 6.7 □	131.0 ± 5.7	123.7 ± 7.9 □	60.1 ± 3.1 □	59.7 ± 1.7 □
Allicin, 0.4 mg/kg (F)	149.3 ± 6.3 □	133.0 ± 6.1 □	127.3 ± 5.7 □	121.5 ± 8.2 □	59.9 ± 2.7 □	58.3 ± 4.2 □

Values are represented as mean ± S.E. (n = 6) and analyzed by one-way analysis of variance (ANOVA). Further analysis between the groups was statistically evaluated by the Newman-Keuls test. P-value <0.05 was regarded as significant. (A) Saline-treated control; (B) Galactosamine (300 mg/kg per day) and lip polysaccharide (30 µg/kg per day) i.p. on the last day of treatment protocol and the animals were sacrificed 24 h after administration.

□ P < 0.01

□□ P < 0.05

D-Galn hepatotoxicity significant increases the level of lipid per oxidation as well as increased the level of iron and ferritin as compared with control rats.

**Table**  
**Effect of allicin on lipid per oxidation and its antioxidant properties**

Parameters	Group I normal control	Group II allicin pretreated (A)	Group III GalN/LPS-intoxicated (B)	Group IV(A + B)
Iron	171 ± 16.70	169 ± 16.56	369.45 ± 17.60†	181.12 ± 16.87†
SOD	6.46 ± 0.75	7.01 ± 0.67	3.36 ± 0.47†	7.46 ± 0.69†
CAT	77.56 ± 7.38	79.12 ± 7.1	36.51 ± 2.97†	67.13 ± 7.81†
LPO	1.39 ± 0.16	1.41 ± 0.13	2.35 ± 0.24†	1.46 ± 0.17†
GSH	5.65 ± 0.49	5.13 ± 0.48	2.97 ± 0.21†	4.63 ± 0.39†
GST	11.16 ± 1.21	11.25 ± 1.31	5.26 ± 0.48†	9.59 ± 0.89†
GPX	81.12 ± 7.86	83.11 ± 8.14	52.56 ± 4.97†	77.63 ± 7.11†
Ferritin	27.71 ± 2.67	25.67 ± 2.49	79.16 ± 7.36†	31.54 ± 3.09†
Ceruloplasmin	39.46 ± 3.43	36.13 ± 3.52	17.12 ± 1.65†	31.23 ± 2.96†

Values expressed as mean ± S.D, n = 6; Group III vs. Group I; Group IV vs. Group III; Student's t-test.

Activity of tissue antioxidants is expressed as: nmol/g liver tissue for GSH; µmol of GSH oxidized/min/mg of protein for GPX; unit/min/mg of protein for GST; 50% inhibition of epinephrine auto-oxidation for SOD; µmoles of hydrogen peroxide decomposed/min/mg of protein for CAT. Activity of ceruloplasmin in serum is expressed as mg/dl. (A) Allicin (0.3 mg/kg per day) diluted in 1% of gum acacia; (B) Galactosamine (300 mg/kg per day); lip polysaccharide (30 µg/kg per day) i.p. on the last day. †P < 0.001. This studied show that oral pre treatment with allicin for 15 days with different dose show significantly normalizes the elevated level of lipid per oxidation and other. Most of the effect is due to its strong SH modifying and antioxidant properties<sup>33</sup>.

**Tridax procumbens**

Tridax weed belonging from family Compositae found all over India and used as indigenous medicine for various hepatotoxicities. It also used from traditionally in Indian medicine as antifungal and insect repellent, anticoagulant, in bronchial catarrh, diarrhea and dysentery. Tridax procumbens also found in 'Bhringraj' which perform a key role in herbal medicine for hepatotoxicity.<sup>34</sup>

**Lygodium flexuosum**

Lygodium belonging to family lygodiaceae is a climbing fern. It also known as vallipanna and distributed all over India mainly in Western Ghats of Kerala. Lygodium flexuosum is very important medicinal plant in Indian ayurveda tradition some scholar state that plant may be 'Rudra jata' its medicinal properties show by all

part of the plant. 'Rudra jata' is an intermediate drug in Ayurveda system. The rhizomes' used in the treatment of jaundice. The root is also used to treat stomach pain by Mech, Oraon and Rabha tribes in Jalpaiguri district of West Bengal, India<sup>35</sup>. D-Galn at dose of 800mg/kg causes elevation in serum AST, ALT, and LDH. When n hexen extract of *Lygodium flexuosum* given at dose of 100mg/kg and 200mg/kg body wt it show remarkably decrease in elevated level of marker enzyme and also protect the live from increasing hepatic malondialdehyde [MDA] formation & normalize the level of

glutathione (GSH) in d-galactosamine treated rats. 200mg/kg dose show more effective than 100mg/kg. Histopathology report show its membrane stabilizing effect and hepatoprotection action is due to inhibiting UDP-sugar derivatives & biosynthesis of glycoprotein<sup>35</sup>. On treatment with *Lygodium flexuosum* for 2, 24 & 48h after D-Galn injection the free radical generated wear scavenged by the extract as evidence from normal liver MDA and GSH level. And its hepatoprotection action is due to inhibiting UDP-sugar derivatives<sup>35</sup>

**Table**  
**Effect of *Lygodium flexuosum* extracts and Silymarin on GSH and MDA in d-galactosamine intoxicated rats (post-treatment groups)**

Treatment gp	MDA(nmol/mg tissue)	GSH(nmol/mg Protein)
Normal control	0.635 ± 0.001	0.522 ± 0.002
<i>Lygodium flexuosum</i> (200 mg/kg) + d-galactosamine	0.637 ± 0.003†	0.520 ± 0.002†
D-galactosamine control (800 mg/kg; i.p.)	1.089 ± 0.006*	0.400 ± 0.003*
<i>Lygodium flexuosum</i> (100 mg/kg) + d-galactosamine	0.676 ± 0.002†	0.498 ± 0.001†
Silymarin (50 mg/kg) + d-galactosamine	0.658 ± 0.002†	0.503 ± 0.001†

Values are means's., n=6.

\*p≤0.05 vs. normal control.

† p≤0.05 vs. d-galactosamine control.

## CONCLUSION

In the above study we discussed about laboratory finding and clinical trial study of natural hepatoprotective plant. Main focused on the natural extract which contain bioactive molecule which show hepatoprotection. Herbal plant extract are used from ancient time to treat various disease associated with liver and now they become a promising therapy for pathological liver state. This time many natural plant and there potent extract are available which show hepatoprotection. These phytochemicals can be extracted and developed as separate drugs, with quality and standards of recent medicinal system. It's very critical time for the pharmaceutical industry in

drug discovery process which are going very expensive, inefficient and riskier. In Indian tradition natural plant play key & wide source of drug many pharmaceutical preparation contain these herbals to enhance their efficacy. Half of Indian medicine preparation contains these natural plants that of the modern concept of evidence-based medicinal evaluation, standardization of herbal products and randomized placebo-controlled clinical trials to support clinical efficacy. Herbal drug served as a materia medica of our living lifestyle, and becoming a promising treatment for pathological liver disorder.

## REFERENCES

- Ramesh Kr. Gupta, Rajnish Kr. Singh, Sudhansu Ranjan Swain, Talib Hussain, Chandana Venkateswara Rao, Anti-hepatotoxic potential of Hedyotis



- corymbosa against D-galactosamine-induced hepatopathy in experimental rodents, Asian Pacific Journal of Tropical Biomedicine (2012)S1542-S1547.
2. F. Stickel, D. Schuppan , Herbal medicine in the treatment of liver diseases, Digestive and Liver Disease 39 (2007) 293–304
  3. Boigk G, Herbst H, Jia JD, Riecken EO, Schuppan D. Effect of antifibrotic agent silymarin on liver cell regeneration in a rat model of secondary biliary fibrosis: a morphometric analysis. J Phytother Res 1998;12(suppl.):S42–4.
  4. P.J. Wills, V.V. Asha, Protective effect of *Lygodium flexuosum* (L.) Sw. (Lygodiaceae) against d-galactosamine induced liver injury in rats. Journal of Ethnopharmacology 108 (2006) 116–123.
  5. Ravikumar V, Shivashangari KS, Devaki T. Hepatoprotective activity of *Tridax procumbens* against D-galactosamine/lipopolysaccharide-induced hepatitis in rats. J Ethnopharmacol 2005; 101: 55-60.
  6. Veereshwarayya Vimal, Thiruvengadam Devaki Hepatoprotective effect of allicin on tissue defense system in galactosamine/endotoxin challenged rats. Journal of Ethnopharmacology 90 (2004) 151–154.
  7. Najmi AK, Pillai KK, Pal SN, Aqil M. Free radical scavenging and hepatoprotective activity of jigrine against D-GalN induced hepatopathy in rats. J Ethnopharmacol 2005; 97: 521-525.
  8. B Ganga Rao, Y Venkateswara Rao, T Mallikarjuna Rao, Hepatoprotective and antioxidant capacity of *Melochia corchorifolia* extracts, Asian Pacific Journal of Tropical Medicine (2013)537-543.
  9. Ramesh Kr. Gupta, Rajnish Kr. Singh, Sudhansu Ranjan Swain, Talib Hussain, Chandana Venkateswara Rao, Anti-hepatotoxic potential of *Hedyotis corymbosa* against D-galactosamine-induced hepatopathy in experimental rodents, Asian Pacific Journal of Tropical Biomedicine (2012)S1542-S1547.
  10. L. Pari, D. Tewas, J. Eckel, Role of curcumin in health and disease, Arch. Physiol.Biochem. 114 (2) (2008) 127e449.
  11. Aihua Zhang, Hui Sun, Xijun Wang, Recent advances in natural products from plants for treatment of liver diseases, European Journal of Medicinal Chemistry 63 (2013) 570-577.
  12. V. Jaishree, Shrishailappa Badami, Antioxidant and hepatoprotective effect of swertiamarin from *Enicostemma axillare* against d-galactosamine induced acute liver damage in rats. Journal of Ethnopharmacology 130 (2010) 103–106.
  13. Yamamura Y, Kotaki H, Tanaka N, Aikawa T, Sawada Y, Iga T. The pharmacokinetics of glycyrrhizin and its restorative effect on hepatic function in patients with chronic hepatitis and in chronically carbon-tetrachloride-intoxicated rats. Biopharm Drug Dispos 1997;18:717–25.
  14. Takahara T, Watanabe A, Shiraki K. Effects of glycyrrhizin on hepatitis B surface antigen: a biochemical and morphological study. J Hepatol 1994;21:601–9.
  15. Wang JY, Guo JS, Li H, Liu SL, Zern MA. Inhibitory effect of glycyrrhizin on NF-kappa B binding activity in CCL4-plus ethanol-induced liver cirrhosis in rats. Liver 1998; 18:180–5.
  16. Arabshahi-Delouee S, Urooj A. Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves. Food Chem 2007; 102: 1223-1240.
  17. Wong C, Li H, Cheng K, Chen F. A systematic survey of antioxidant activity of 30 Chinese medicinal plants using the ferric reducing antioxidant power assay. Food Chem 2006; 97: 705-711.
  18. B Ganga Rao, Y Venkateswara Rao, T Mallikarjuna Rao, Hepatoprotective and antioxidant capacity of *Melochia corchorifolia* extracts, Asian Pacific Journal of Tropical Medicine (2013)537-543.
  19. Lotito SB, Frei B. Consumption of flavonoid-rich foods and increased plasma antioxidant capacity in humans:

- Cause, consequence, or epiphenomenon. *Free Radical Res* 2006; 41:1727-1746.
20. Rao PV, Sujana P, Vijayakanth T, Naidu MD. *Rhinacanthus nasutus* – Its protective role in oxidative stress and antioxidant status in streptozotocin induced diabetic rats. *Asian Pac J Trop Dis* 2012; 2(4): 327-330.
  21. Kumbhare MR, Guleha V, Sivakumar T. Estimation of total phenolic content, cytotoxicity and in vitro antioxidant activity of stem bark of *Moringa oleifera*. *Asian Pac J Trop Dis* 2012; 2(2):144-150.
  22. Kumar RS, Raj Kapoor B, Perumal P. Antioxidant activities of *Indigofera cassioides* Rottl. Ex. DC. using various in vitro assay models. *Asian Pac J Trop Biomed* 2012; 2(4): 256-261.
  23. Shahwar D, Raza MA. Antioxidant potential of phenolic extracts of *Mimusops elengi*. *Asian Pac J Trop Biomed* 2012; 2(7): 547-550.
  24. Sara Tulipani, Bruno Mezzetti, Franco Capocasa, Stefano Bompadre, Jules Beekwilder CH, Ric de Vos, et al. Antioxidants, phenolic compounds, and nutritional quality of different strawberry genotypes. *J Agric Food Chem* 2008; 56 (3): 696-704.
  25. N Niciforovic, V Mihailovic, P Maskovic, S Solujic, A Stojkovic, D, Pavlovic Muratspahic. Antioxidant activity of selected plant species; potential new sources of natural antioxidants. *Food & Chem Toxicol* 2010; 48(11): 3125-3130.
  26. J.M. Smoliga, J.A. Baur, H.A. Hausenblas, Resveratrol and health: a comprehensive review of human clinical trials, *Mol. Nutr. Food. Res.* 55 (8) (2011) 1129e1141.
  27. C.C. Chan, L.Y. Cheng, C.L. Lin, Y.H. Huang, H.C. Lin, F.Y. Lee, The protective role of natural phytoalexin resveratrol on inflammation, fibrosis and regeneration in cholestatic liver injury, *Mol. Nutr. Food. Res.* 55 (12) (2011) 1841e1849.
  28. D. Rajasekaran, J. Elavarasan, M. Sivalingam, E. Ganapathy, A. Kumar, K. Kalpana, D. Sakthisekaran, Resveratrol interferes with N-nitrosodiethylamine-induced hepatocellular carcinoma at early and advanced stages in male Wistar rats, *Mol. Med. Rep.* 4 (6) (2011) 1211e1217.
  29. H.B. Yu, H.F. Zhang, X. Zhang, D.Y. Li, H.Z. Xue, C.E. Pan, S.H. Zhao, Resveratrol inhibits VEGF expression of human hepatocellular carcinoma cells through a NF-kappa B-mediated mechanism, *Hepatogastroenterology* 57 (102e103) (2010) 1241e1246.
  30. T. Tunali-Akbay, O. Sehirli, F. Ercan, G. Sener, Resveratrol protects against methotrexate-induced hepatic injury in rats, *J. Pharm. Pharm. Sci.* 13 (2) (2010) 303e310.
  31. E.S. Lee, M.O. Shin, S. Yoon, J.O. Moon, Resveratrol inhibits dimethylnitrosamine-induced hepatic fibrosis in rats, *Arch. Pharm. Res.* 33 (6) (2010) 925e932.
  32. Wei Zhao, Jun-Jie Li, Shu-Qiang Yue, Lin-Ying Zhang, Ke-Feng Dou. Antioxidant activity and hepatoprotective effect of a polysaccharide from *BeiChaihu* (*Bupleurum chinense* DC). *Carbohydrate Polymers* 89 (2012) 448– 452.
  33. Ramesh Kr. Gupta, Rajnish Kr. Singh, Sudhansu Ranjan Swain, Talib Hussain, Chandana Venkateswara Rao. Anti-hepatotoxic potential of *Hedyotis corymbosa* against D-galactosamine-induced hepatopathy in experimental rodents. *Asian Pacific Journal of Tropical Biomedicine* (2012)S1542-S1547.
  34. Veereshwarayya Vimal, Thiruvengadam Devaki, Hepatoprotective effect of allicin on tissue defense system in galactosamine/endotoxin challenged rats. *Journal of Ethnopharmacology* 90 (2004) 151–154.
  35. Vilwanathan Ravikumar, Kanchi Subramanian Shivashangari, Thiruvengadam Devaki. Hepatoprotective activity of *Tridax procumbens* against d galactosamine/lipopolysaccharide-induced hepatitis in rats. *Journal of Ethnopharmacology* 101 (2005) 55–60.
  36. P.J. Wills, V.V. Asha, Protective effect of *Lygodium flexuosum* (L.) Sw.

- (Lygodiaceae) against d-galactosamine induced liver injury in rats *Journal of Ethnopharmacology* 108 (2006) 116–123.
37. Swarnalatha.L, P.Neelakanta Reddy, Hepatoprotective activity of *Sphaeranthus amaranthoides* on D-galactosamine induced hepatitis in albino rats, *Asian Pacific Journal of Tropical Biomedicine* (2012) S1900-S1905.
  38. Sire O, Mangenev M, Montagne J, Nordmann R, Nordmann J: Carnitine palmitoyltransferase. Inhibition by D-galactosamine and role of phospholipids. *Eur J Biochem* 1983, 136:371-375.
  39. Drury rva, Walligton, et al., *carltons histological techniques*, 5<sup>th</sup> ed, oxford university press, new york, 1980; 139-142.
  40. Van Rossum TG, Vulto AG, De Man RA, Brouwer JT, Schalm SW. Review article: glycyrrhizin as a potential treatment for chronic hepatitis C. *Aliment Pharmacol Ther* 1998;12:199–205.
  41. Y. Rivera-Espinoza, P. Muriel, Pharmacological actions of curcumin in liver diseases or damage, *Liver Int.* 29 (10) (2009) 1457-1466.
  42. Shiki Y, Shirai K, Saito Y, Yoshida S, Mori Y, Wakashin M. Effect of glycyrrhizin on lysis of hepatocytes membranes induced by anti-liver cell membrane antibody. *J Gastroenterology Hepatol* 1992; 7:12–6.
  43. R. Anandan, T. Devaki. Hepatoprotective effect of *Picrorrhiza kurroa* on tissue defence system in D-galactosamine-induced hepatitis in rats. *Fitoterapia* 70 [1999. 54].
  44. S. Roy, S. Sannigrahi, S. Majumdar, B. Ghosh, B. Sarkar, Resveratrol regulates antioxidant status, inhibits cytokine expression and restricts apoptosis in carbon tetrachloride induced rat hepatic injury, *Oxid. Med. Cell. Longev.* 2011 (2011) 703676.