

**BIOREDUCTION AND FORMATION OF GOLD NANOPARTICLES BY
SOLANUM TORVUM LEAVES EXTRACT****B.S. NAVEEN PRASAD*¹ AND T.V.N. PADMESH ²**¹*Department of Chemical Engineering, Sathyabama University, Chennai – 600119, India*²*Department of Chemical Engineering, Manipal International University, Nilai, Malaysia***ABSTRACT**

Development of biologically inspired experimental processes for the synthesis of nanoparticles might be important in biomedical applications. *Solanum torvum* extract when challenged with chloroauric acid leads to formation of gold nanoparticles. The formations of gold nanoparticles were studied by UV-vis spectra and their size of gold nanoparticles in the colloidal solutions was investigated by HRTEM, the average size of gold nanoparticles was found to be 8.4 nm. The rapid reduction of AuCl_4^- ions highlights the possibility of green pathways to produce technologically important nanomaterials.

KEYWORDS: *Solanum torvum*; biomaterials; gold nanoparticles; electron microscopy**B.S. NAVEEN PRASAD**

Department of Chemical Engineering, Sathyabama University, Chennai – 600119, India

*Corresponding author

INTRODUCTION

Synthesis of nanoparticles with well-defined shape, size and composition is a big challenge in nanotechnology¹. Metal nanoparticles have received considerable attention in recent years due to their enormous applications in different fields of science. Further, there is a growing need to develop an eco-friendly process for nanoparticles synthesis and hence the focus turned towards “green” chemistry. The synthesis of metal nanoparticles has in particular, received considerable attention and potential applications in single electron tunneling², non-linear optical devices³, DNA sequencing⁴, biomedical devices⁵, detection of genetic disorders^{6,7}, gene therapy⁸, biological labeling^{9,10}, biosensing^{5,10}, antimicrobial activity^{11,12}, anticancer¹³ and effective drug delivery¹⁴. Various chemical and physical synthetic protocols, aimed at controlling the physical properties of the particles, are presently employed in the production of metal nanoparticles. Biological synthesis of nanoparticles using plants, seaweeds and microorganisms proves to be biocompatible and hence used for biomedical applications. Plant based synthesis of gold nanoparticles is more advantageous over other biological processes as it is best suited for large-scale synthesis of nanoparticles¹⁵. Gold nanoparticles from the medicinally useful plant *Adhatoda vasica* Nees which has anti-allergic properties and can be used for various therapeutic applications¹⁶. Extracellular shape selective formation of single crystalline triangular gold nanoparticles was observed using the extract of the lemongrass plant (*Cymbopogon flexosus*)¹⁵. A number of different microorganisms including bacteria, yeast and fungi are involved in the reduction of gold ions and formation of gold nanoparticles intra and extracellularly^{1,17}. Seaweed mediated synthesis of gold nanoparticles has also played a major role in therapeutic applications. Recently, *Sargassum myriocystum*, pharmaceutically important seaweed has been used in the synthesis of AuNPs which possess heparin like compounds used in the treatment of cardiovascular diseases. Size controlled

synthesis of gold nanoparticles is challenging as it is one of the major strategies in biological applications. In this consideration, cyanobacteria *Spirulina platensis* have been used for the synthesis of Au nanoparticles which resulted in particles of size in the range of 6–12 nm¹⁸. *S. torvum* is used vastly in traditional medicine to cure diseases like cough, liver and spleen enlargement¹⁹. It is intensively used worldwide in the treatment of fever, wounds, tooth decay, arterial hypertension etc²⁰. It also possesses antimicrobial, antiviral, anti-inflammatory, anti-diabetic and nephroprotective properties. Compounds isolated from *Solanum* species have anticancer potential and is used in the treatment of eye diseases^{21, 22}. As traditional herb possesses various pharmacological applications, it has been chosen for the study to synthesize AuNPs for biomedical applications. In the present study, AuNPs have been synthesized from the potent plant *S. torvum* and the physico-chemical characteristics have been studied.

MATERIALS AND METHODS

Solanum torvum (Solanaceae) leaves collected from Chennai (latitude 13.0839° N and longitude 80.2700° E). 100g of fresh leaves of *Solanum torvum* were washed three times with deionized water. It was grinded to juice in a glass mortar. The extracts were filtered using whatman filter paper No.1 and the filtrates of 1 mL extract was added to 100 mL of deionized water incubated at room temperature. For reduction of chloroaurate ions 1 mL of aqueous AuCl₄⁻ ions in solution was monitored by periodic sampling of aliquots (2 mL) of the aqueous component and measuring the UV-vis spectrum of the solution. UV-vis spectra of these aliquots were monitored as a function of time of reaction on a Shimadzu spectrophotometer (model 1601) operated at a resolution of 2 nm. TEM analysis was performed using a JEOL 3010 instrument operated at an accelerating voltage at 120 kV. Samples for high resolution transmission electron microscopy analysis were prepared by drop coating Au nanoparticles solutions

onto carbon-coated copper TEM grids. The films on the TEM grids were allowed to stand for 2 min, following which the extra solution was removed using a blotting and the grid was allowed to dry prior to measurement.

RESULTS AND DISCUSSION

It is known that the physical properties of biologically synthesized nanoparticles may vary according to the bioreductive natural resources. In the present investigation polydispersed gold nanoparticles were observed to occur when the concentration of HAuCl_4 increased. (Fig. 1a) shows the before immersion HAuCl_4 solution the green colour of the plant extract (*S. torvum*) can clearly be seen. (Fig.1b) shows the formation of ruby red color by the increased concentration of aqueous AuCl_4^- ions indicates the formation of gold nanoparticles in solution. Gold

nanoparticles absorb radiation in the visible region of the electromagnetic spectrum (ca. 520 nm) because of excitation of surface plasmon vibrations giving gold nanoparticles the striking ruby red colours in various media²³. UV-vis spectra show no evidence of absorption in the range of 400-800 nm for the plant extract, the plant extract solution exposed to AuCl_4^- ions show a distinct absorption at around 528 nm of nanoparticles obtained (Fig.2). The absorbance is close to that observed for thin films of gold nanoparticles grown by different techniques²⁴. It is observed that the gold surface plasmon resonance band occurs initially at ca 523 nm after completion of the reaction, the wavelength of the surface plasmon band stabilizes at 528 nm. Reduction of the AuCl_4^- ions are completed after 45 min of reaction, in both nanocrystals and nanoparticles formation.



Figure 1

(a) *S.torvum* leaf extract, (b) Ruby red color indicating the formation of gold nanoparticles by reduction of the aqueous metal ions during exposure to the *S.torvum*.

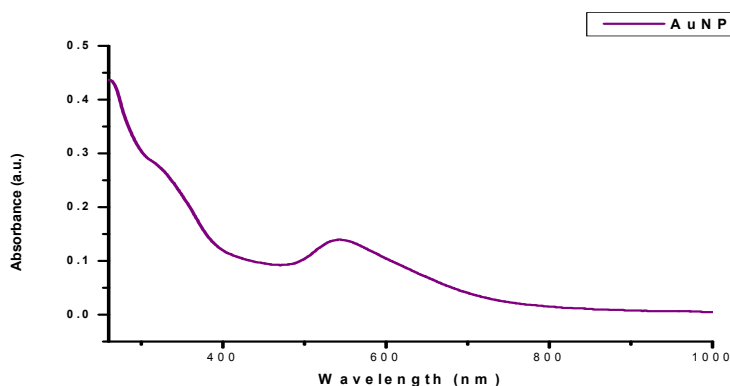


Figure 2

UV-vis spectra recorded as a function of time of reaction of aqueous solutions of chlorauric acid with *S. torvum*.

High resolution transmission electron microscopy has provides further insight in the morphology and size details of the gold nanoparticles. A representative HR-TEM image recorded from the gold nanoparticles is shown in (Fig 3a). In general, the particles are isotropic (i.e low aspect ratio) in shape and reasonably spherical in structure. All the nanoparticles are well separated and no agglomeration was noticed. From the HR-TEM images we obtained the average size of gold nanoparticles of 8.4 nm. The histogram of the nanoparticles size distribution (Fig 3b) is obtained by measuring the size of about 10 nanoparticles. Synthesis and characterization of nanocrystals of materials have become an area of intense research over the last few ears. Several materials scientist have been reported for the preparation of nanocrystals of

metals such as Au, Ag, CdS and CdSe using chemical and physical methods²⁵⁻²⁹. Presently, nanocrystals of metals are finding a variety of applications starting from biological tagging to electronic devices³⁰. A key challenge in application of these materials is to prevent agglomeration of nanomaterials to overcome in the present study and it may be due to the surface function utilization/ stabilization of the *S. torvum* extract. Recently, biosynthetic methods employing either biological microorganisms or plant extracts have emerged as a simple and viable alternative to chemical synthetic procedures and physical methods. Many reports on synthesis of metals (Au, Ag) and semiconductor nanoparticles (CdS, Pt and Pd) using various microorganisms³¹⁻³⁴ have been appeared.

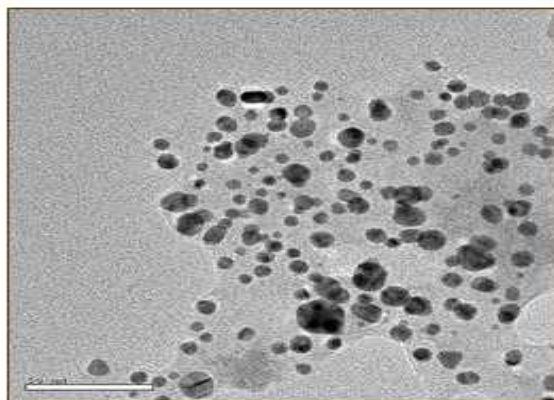


Figure 3a
HR-TEM image of gold nanoparticles synthesized using Solanum torvum

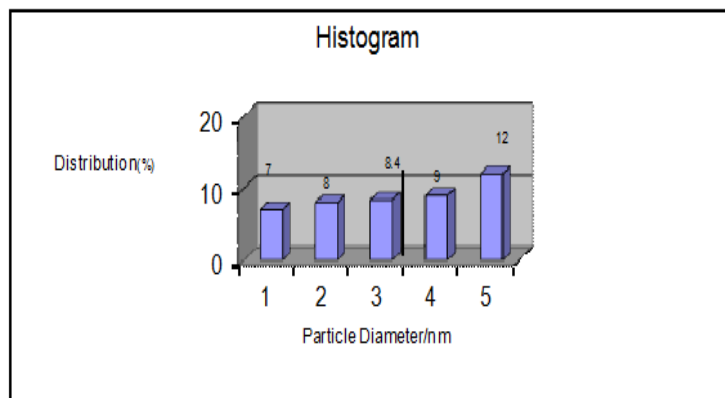


Figure 3b
Shows the histogram of gold nanoparticles size distribution

CONCLUSION

A process for the rapid synthesis of stable gold nanoparticles using *S. torvum* leaf extract and to develop a simple and efficient route for the synthesis of nanomaterials, within 45 min has been proposed. The ability to synthesis gold nanoparticles rapidly with morphology

control by eco-friendly biological methods is exciting and represents an important advance in making them a viable alternative to the more popular chemical methods. The production of nanoparticles highlights the possibility of green pathway to produce nanomaterials also easily amenable for large-scale production.

REFERENCES

1. Gericke M and Pinches A. Biological Synthesis of metal nanoparticles. Hydrometallurgy, 83 (1-4): 132-140, (2006).
2. Andres R P, Bein T, Dorogi M, Feng S, Henderson J I, Kubiak C P, Mahoney W, Osifchin R G, Reifenger R, Coulomb staircase at room temperature in a self-assembled molecular nanostructure. Science, 272 (5266): 1323-1325, (1996).
3. Galletto P, Brevet P F, Girault H H, Antoine R, Broyer M, Enhancement of the second harmonic response by adsorbates on gold colloids: The effect of aggregation. J. Phys.Chem.B, 103 (41): 8706-8710, (1999).
4. Mirkin C A, Letsinger R L, Mucic R C, Storhoff J J, A DNA-based method for rationally assembling nanoparticles into macroscopic materials. Nature, 382 (6592): 607-609, (1996).
5. See the Feb.28, 2000 issue of Chem. Eng. News and articles by R.Dagani therein for coverage of the new applications envisaged for nanomaterials.
6. Taton T A, Mirkin C A, Letsinger R L, Scanometric DNA array detection with nanoparticle probes. Science, 289 (5485): 1757-1760, (2000).
7. Cao Y C, Jin R, Mirkin C A, Nanoparticles with Raman Spectroscopic Fingerprints for DNA and RNA detection. Science 297 (5586): 1536-1540, (2002).
8. Sandhu K K, McIntosh C M, Simard J M, Smith S W, Rotello V M, Gold nanoparticle mediated transfection of mammalian cells. Bioconjugate chem. B. 13(1): 3-6, (2002).
9. Nicewarner-Pena S R, Freeman R G, Reiss B D, He L, Pena D J, Walton I D, Cromer R, Keating C D, Natan M J, Submicrometer metallic barcodes. Science 294 (5540): 137-141, (2001).
10. Han M, Gao X, Su J Z, Nie S, Quantum dot Tagged microbeads for multiplexed optical coding of Biomolecules. Nat. Biotechnol, 19: 631-635, (2001).
11. Zhilong Shi, Neoh K G, Kang E T, Surface grafted viologen for precipitatin of silver nanoparticles and their combined bactericidal activities. Langmuir, 20 (16): 6847 – 6852, (2004).
12. Elechiguerra J L, Burt J L, Morones R J, Camacho A, Gao X, Lara H H, Yacaman M J, Interaction of silver nanoparticles with HIV-1. J.Nanobiotechnol, 3: 1-10, (2005).
13. Jain P K, El-Sayed I H, El-Sayed M A, Au nanoparticles target cancer. Nanotoday, 2 (1): 18-29, (2007).
14. Joshi H M, Bhumkar D R, Kalpana Joshi, Varsha Pokharkar, Murali Sastry, Gold nanoparticles as carriers for efficient transmucosal insuling delivery. Langmuir, 22 (1): 300 – 305, (2006).
15. Shankar S S, Rai A, Ankamwar B, Singh A, Ahmad A, Sastry M, Biological synthesis of triangular gold nanoprisms. Nat.Mater, 3 :482-488, (2004).
16. Karthick V, Kumar V G, Maiyalagan T, Deepa R, Govindaraju K, Rajeswari A, Dhas T S, Green Synthesis of Well Dispersed Nanoparticles using leaf extract of medicinally useful *Adhatoda Vasica Nees*, Micro and Nanosystems 4 (3):192–198 (2012).
17. Mandal D, Bolander M E, Mukhopadhyay D, Sarker G, Mukherjee P, The use of microorganisms for the

- formation of metal nanoparticles and their application. Appl. Microbiol. Biotechnol, 69 (5): 485-492, (2006).
18. Govindaraju K, Basha S K, Kumar V G, Singaravelu G, Silver, gold and bimetallic nanoparticles production using single-cell protein (*Spirulina platensis*) Geitler, J. Mater. Sci. 43: 5115–5122, (2008).
 19. Siemonsma J, Piluek K, Plant Resources of South-East Asia 8 (PROSEA), Bogor, Indonesia, 1994, pp. 412.
 20. Ndebia E J, Kamga R, Nchunga-Anye Nkeh B, Analgesic and anti-inflammatory properties of aqueous extract from leaves of *Solanum torvum* (Solanaceae), Afr J Tradit Complement Altern Med, 4 (2): 240 - 244 (2007).
 21. Oliver-Bever B, Medicinal plants in tropical West Africa, Cambridge University Press, Cambridge, 104, (1986).
 22. Watt J M , Breyer-Bradwijk, Gerdina m, Medicinal and poisonous plants of Southern and Eastern Africa, 1962 Edn, Vol 2, E. & S.Livingstone Ltd, Edinburgh : 1457, (1962).
 23. Mulvaney P, Surface Plasmon Spectroscopy of nanosized metal particles. Langmuir, 12(3): 788-800, (1996).
 24. Underwood S, Mulvaney P, Effect of the solution refractive index on the colour of gold colloids. Langmuir 10 (10): 3427-3430 (1994).
 25. Brust M, Bethell D, Schiffrin D J, Kiely C J, Novel Gold-dithiol nano networks with non-metallic electronic properties. Adv.mater, 7(9): 795-797, (1995).
 26. Rockenberger J, Scher E J, Alivisatos A P, A new non- hydrolytic single precursor approach to surfactant-capped nanocrystals of transition metal oxides J.Am.Chem.Soc, 121(49): 11595-11596, (1999).
 27. Murray C B, Norris D J, Bawendi M G, Synthesis and characterization of nearly monodisperse CdeE (E.sulfur, Selenium, Tellurium) Semiconductor nanocrystallites. J. Am. Chem.Soc.115 (19): 8706-8715, (1993).
 28. Sarathy K V, Kulkarni G U, Rao C N R, C.N.R. A novel method of preparing thiol-derivarised nanoparticles of gold, platinum and silver forming superstructures. J.Chem.Soc, Chem.Commun, (6): 537-538, (1997).
 29. Duff D G, Baiker A, Edwards P P, A new hydrosol of gold clusters. Formation and particle size variation. Langmuir, 9 (9): 2301-2309, (1993).
 30. Rao C N R, Kulkarni G U, John Thomas P, Agrawal V V, Gautam U K, Ghosh M, Nanocrystals of metals, Semiconductors and oxides: Novel synthesis and applications. Curr.Sci, 85 (7): 1041-1045, (2003).
 31. Nair B, Pradeep T, Coalescence of Nanoclusters and formation of submicron crystallites assisted by *Lactobacillus* strains. Crys. Growth & Design, 2 (4): 293-298, (2002).
 32. Ahmad A, Mukherjee P, Mandal D, Senapati S, Islamkhan M, Rajiv K, Sastry M, Enzyme mediated extracellular synthesis of CdS nanoparticles by the fungus, *Fusarium oxysporum*. J.Am.Chem.Soc, 124 (41): 12108-12109, (2002).
 33. Lengke M F, Fleet M E, Southam G, Synthesis of platinum nanoparticles by reaction of filamentous cyanobacteria with platinum (IV)-chloride complex. Langmuir 22 (17): 7318-7323, (2006).
 34. Ping Yong, Rowson N A, Farr J P G, Haris, I.R. and Macaskie, L.E, Bioreduction and Biocrystalization of palladium by *Desulfovibrio desulfuricans* NCIMB 8307. Biotechnol.Bioeng. 80 (4): 369-379 (2002).