



INFLUENCE OF NaCl MEDIATED SALINITY STRESS ON LIPID PEROXIDATION IN GERMINATING SEEDS OF SOYBEAN

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ABSTRACT

Salinity is one of the major abiotic stress that affects the growth and development of many important legume plants. Soybean is a most important oilseed legume cash crop of the world. It is also considered as an important source of high quality cheap protein and oil. In the present investigation, the effect of different concentrations of NaCl (50 mM, 100 mM, 150 mM and 200 mM) on lipid peroxidation during germination of soybean seeds was studied. The lipid peroxidase activity remarkably increases as concentration of salinity increases. The result revealed that, the activity of lipid peroxidation significantly increases up to 72 hrs in all treatments except 50 mM NaCl treatment.

KEYWORDS: Soybean, NaCl Salinity, Lipid peroxidase



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INTRODUCTION

Soybean plays a crucial role in world food trade. It has the highest level of protein (40%) in comparison with other leguminous plant. The oil content (18%) in the soybean is second only to groundnut among food legume. In worldwide, 20% of irrigated land and 2.1% of dry land agriculture suffers from the salt problem¹. Several biotic, abiotic and socio-economic constraints limit the productivity of soybean in India². Recently, Patil *et al.*,³ reported that NaCl mediated salinity stress adversely affects the germination percentage in soybean. Seed germination is much more complex process and sensitive life stage of plant that involves transition of metabolically quiescent to active and growing entity. According to Bailly⁴ seed germination and post-germination seedling development are well regulated process in plant physiology involving high metabolic activity and generation of reactive oxygen species (ROS) in the cell. Reactive oxygen species (ROS) play dual role in plants as they acts as signalling molecule in cell and simultaneously acts as toxic product accumulating under stress conditions⁵. Generally, reactive oxygen species are highly cytotoxic and can easily react with lipids, proteins, nucleic acids etc. causing lipid peroxidation, degradation of proteins and mutagenesis respectively^{6, 7, 8}. Oxidative degradation of lipid is termed as lipid peroxidation. In lipid peroxidation process polyunsaturated fatty acids in biological membrane system undergoes changes reaction as a result form lipid hydroperoxides^{9, 10}. MDA is the final product of lipid peroxidase. It may acts as marker for lipid

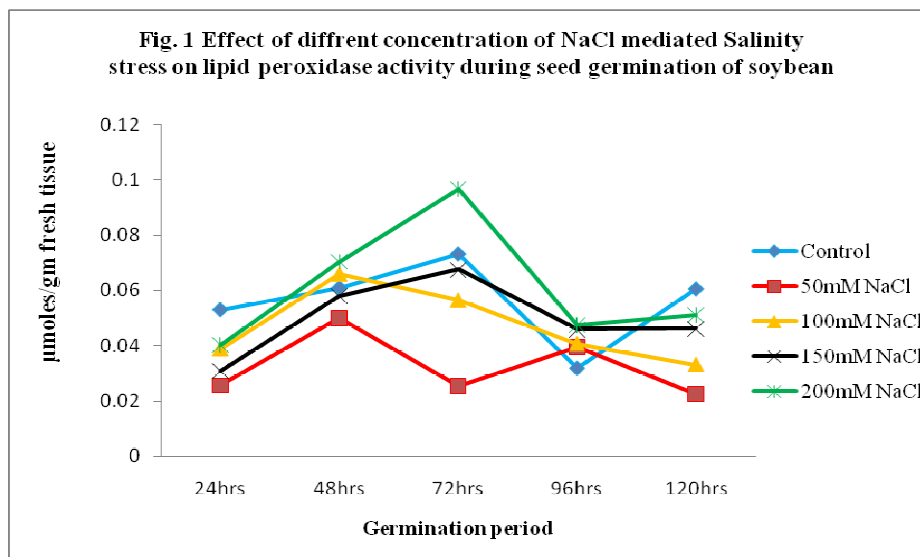
peroxidation and formed by the reaction of ROS with lipid molecules¹¹. No information can be deciphered from the literature on the influence of NaCl-salinity on lipid peroxidation during germinating soybean seeds.

MATERIALS AND METHODS

Morphologically healthy seeds of soybean (JS-335) were first surface sterilized with 1% of HgCl₂ for 2 mins and then washed with distilled water to remove toxic elements. Petri-plates were sterilized with absolute alcohol and lined with filter paper at bottom. Twenty uniform seeds were placed in each petri-plate. The desired treatments were given by adding 15cm³ of aqueous treatment solutions (water-control, 50 mM, 100 mM, 150 mM and 200 mM of NaCl). The petri-plates were incubated in a BOD incubator at 26±2°C in dark and investigations were covered at different stages of germination from 24 to 120 hrs. The activity of enzyme lipid peroxidase was studied by method of Cakmak and Hort¹² and expressed in term of µmoles/gm of fresh tissue.

RESULTS AND DISCUSSION

The results obtained in present study is shown in Fig.1. The results revealed that, the activity of lipid peroxidase was significantly increased by all the treatment upto 48hrs but noteworthy induction was found by 200 mM NaCl treatment up to 72 hours, however decline in activity in lipid peroxidation was reported to increase in germination period with all the treatment except 50 mM NaCl treatment.



The oxidative stress may be observed in germinating soybean seeds due to salinity stress. These results are also in agreement with reports of Anita and Usha¹³. They stated that salinity stress is enhancing lipid peroxidase activity in leaves of soybean cultivars. According to Neto *et al.*,¹⁴ the increase in lipid peroxidase activity is a one of the criteria to differentiate salt sensitive and salt tolerance variety. They reported that maize genotype increases different peroxidase activities due to salt treatment indicates salt tolerance nature in maize. Estandiari *et al.*,¹⁵ observed increase lipid peroxidase activity in wheat seedlings under salinity. Similarly, Increase in MDA contents under salt stress was also found in rice¹⁶, alfalfa¹⁷, cotton¹⁸. Sreenivasulu *et al.*,¹⁹ emphasized that the accumulation of MDA was more in the salt susceptible than in the salt tolerance cultivars. In the present study, the decrease in lipid peroxidase activity reported after 72 hrs similar, report was also noticed by Liang *et al.*,²⁰ in salt treated barley plant. The intensity of stress factor has a

direct correlation with lipid peroxidation development. The balance between lipid peroxidation and antioxidant system is serve as pre-requisite for proper functioning of cells.

CONCLUSION

Thus, it has been concluded that, salinity induced product of lipid peroxidation may act as signal molecule that might help to reduce adverse effects of salinity on further development of soybean plants. Present study will help to study correlation between antioxidant enzymes and lipid peroxidase under salinity stress.

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REFERENCES

1. FAO, Global network on integrated soil management for sustainable use of salt-affected soils. (2000). Available in <http://www.fao.org/ag/AGL/agll/spush/intro.htm>.
2. Bhatnagar PS, and Joshi OP, Current status of soybean in production and utilization in India. Proceedings of VII World Soybean Research Conference - IV International Soybean Processing and Utilization Conference – III Brazil, 2004, pp. 27–37.
3. Patil DB, Jadhav SH, and Bhamburdekar SB, Interactive effect of Boron and NaCl on germination performance in soybean. International J. of Applied Biology and Pharmaceutical Technology, 3: 366-368, (2012).

4. Bailly C, Active oxygen species and antioxidants in seed biology. *Seed Sci. Res*, 14: 93–107, (2004).
5. Bailly C, El-Maarouf-Bouteau H, Corbineau F. From intercellular signaling networks to cell death: the dual role of reactive oxygen species in seed physiology. *CR Biology*, 331(10): 806-814, (2008).
6. Breusegem FV, Vranova E, Dat JF, and Inze D, The role of active oxygen species in plant signal transduction, *Plant Science*, 161:405-414, (2001).
7. Scandalios JG, Oxygen stress and superoxide dismutase, *Plant Physiology*, 101: 712-726, (1993).
8. Quiles MJ., Lopez NI, Photoinhibition of photosystems I and II induced by exposure to high light intensity during oat plant grown effects on the chloroplastic NADH dehydrogenase complex, *Plant Science*, 163: 1037-1046, (2004).
9. Gutteridge JM C, Wilkins S, Copper dependent hydroxyl radical damage to ascorbic acid. *Fed. Eur. Soc. Letters*, 137: 327-329, (1995).
10. Isamah GK, Asagba SO, and Thomas AE, Lipid peroxidation o-diphenolase, superoxide dismutase, and catalase profile along the three physiological regions of *Dioscorea rotundata* pir cv Omi. *Food Chem*, 69: 1-4, (2000).
11. Shimizu N, Hosogi N, Hyon GS, Jiang K, Park P, Reactive oxygen species (ROS) generation and ROS induced lipid peroxidation are associated with plant membrane modifications in host cells in response to AK-toxin from *Alternaria Japanese Pear* pathotype. *J. Gen Plant Pathol*, 72: 6-15, (2006).
12. Cakmak I, and Horst JH, Effect of aluminium on lipid peroxidation, superoxide dismutase, catalase and peroxidase activities in root tips of soybean (*Glycine max*). *Physiologia Plantarum*, 83: 463-468, (1991).
13. Anita T, and Usha R, Effect of salinity stress on physiological, biological and antioxidant defense systems of high yielding cultivars of soybean. *International J. of Pharma and Biosciences*, 3(4): 851-864, (2012).
14. Neto Azevedo de Andre Doas, Jose Tarquinio Prisco, Joaquim Eneas-Filho, Carlos Eduardo Braga de Abreu And Eneas Gomes- Filho, Effect of salt stress on Antioxidative enzymes and lipid peroxidation in leaves and roots of salt-tolerant and salt-sensitive maize genotypes. *Environmental and Experimental Botany*, 56: 87-94, (2006).
15. Esfandiari E, Fariborz Shekari, Farid Shekari, Manouchehr Esfandiari, The effect of salt stress on antioxidant enzymes activity and lipid peroxidation on the wheat seedling, *Not. Bot. Hort. Agrobot. Cluj*, 35(1): 1842-4309, (2007).
16. Tijen D, and Ismail T, Comparative lipid peroxidation, antioxidant defense systems and praline content in roots of two rice cultivars differing in salt tolerance. *Environ Exp Bot*, 53: 247-257, (2005).
17. Wang XS, and Han JG, Effects of NaCl and silicon on ion distribution in the roots, shoots and leaves of two alfalfa cultivars with different salt tolerance. *Soil Sci Plant Nutr*, 53(3): 278-285, (2007).
18. Diego AM, Marco AO, Carlos AM, Jose C, Photosynthesis and activity of superoxide dismutase peroxidase and glutathione reductase in cotton under salt stress. *Environ Exp Bot*, 49: 69-76, (2003).
19. Sreenivasulu N, Ramanjulu V, Ramachandru K, Prakash S, Shekar-Shetty H, Savithri S, Sudhakar C, Total peroxidase activity and peroxidase isoforms as modified by salt stress in two cultivars of fox tail millet with differential salt tolerance. *Plant Sci*, 141: 1-9, (1999).
20. Liang, Yongchao Qin Chen, Qian Liu, Wenhua Zhang and Ruixing Ding, Exogenous silicon (Si) increases antioxidant enzyme activity and reduces lipid peroxidation in roots of salt-stressed barley (*H. Vulgare* L.). *Journal of plant physiology*, 160: 1157-1164, (2003).