



SCREENING OF MEDICINAL AND AROMATIC PLANTS OF HIMACHAL PRADESH FOR ANTIPROLIFERATIVE ACTIVITIES AGAINST BUDDING YEAST *SACCHAROMYCES CEREVISIAE*

ANIL KUMAR, KAMAL DEV AND ANURADHA SOURIRAJAN*

*Faculty of Applied Science and Biotechnology, Shoolini University,
Solan, Himachal Pradesh, India.*

ABSTRACT

Medicinal plants contain a lot of bioactive compounds having various activities against human diseases. Based on traditional knowledge 62 plants (including 46 medicinal, 2 aromatic and 14 non medicinal plants) belonging to 38 families were collected from different regions of Himachal Pradesh. The plants from Himalayan region were tested for their anti-proliferative activities by using *Saccharomyces cerevisiae* as a drug discovery tool. The ethanolic extracts and essential oils of different plant parts such as flowers, leaves, fruit and rhizome were tested for anti-proliferative activity against budding yeast using agar well diffusion assays. Amongst the tested plants, *Curcuma longa* (rhizome) (Zingiberaceae) showed highest anti-proliferative effect with a zone of clearance of 26 mm and found fungicidal for yeast growth. *Chrysanthemum indicum* (leaves) (Asteraceae) exhibited anti-proliferative activity, with zone of clearance of 19.5 mm and fungistatic effect. In comparison, *Aegle marmelose* (leaves) (Rutaceae) showed least anti-proliferative effect, with zone of clearance of 18.5 mm and found fungicidal. The essential oils of *Cymbopogon citratus* and *Rosmariuns officinalis* also exhibited strong anti-proliferative effects against yeast growth. The phytochemicals of these plants responsible for anti-proliferative effect in yeast can be purified and explored for anticancer potential in humans.

KEYWORDS: yeast, anti-proliferative, anti-cancer, cell cycle, medicinal plants, phytochemicals, Himachal Pradesh, *Saccharomyces cerevisiae*

*Corresponding author



ANURADHA SOURIRAJAN

Faculty of Applied Science and Biotechnology, Shoolini University,
Solan, Himachal Pradesh, India.

EMAIL: ASOURIRAJAN@GMAIL.COM

INTRODUCTION

Plants serve as a valuable resource for the isolation of novel bioactive compounds to combat various human diseases¹. There are several plants which have been used since ancient times as natural product for treating diseases by village folk². The phytoconstituents isolated from plants have chemical substances which bind to effective target sites in human body³. The most important bioactive compounds from plants are alkaloids, glycosides, terpenoids, steroids, flavanoids⁴, tannins, saponins, sugars, cumarins⁵, proteins, and phenols⁶. Medicinal plants have been used for treating communicable as well as non communicable diseases. Human communicable diseases caused by bacteria, fungi and viruses are the leading cause of deaths in developing countries⁷. Several phyto-compounds and essential oils have been reported from medicinal and aromatic plants for their anti-microbial action against bacterial and fungal pathogens⁸. However, they have not been explored much for treating non communicable diseases like cancer. Of the 250,000 -500,000 species found on earth, ~ 3000 plant species have reportedly been used in treatment of cancer⁹. Since 1990, the National Cancer Institute (NCI), USA has screened more than 60,000 compounds against different human cancer cell lines¹⁰. Almost 75% of the anti-cancer drugs prescribed are derived from medicinal plants¹¹. The most common and extensively used plant derived anti-cancer drugs include vinblastine, vincristine, taxanes, and camptothecins¹². Most of the plant derived bioactive compounds used in anticancer therapy are discovered through large scale screening programs, which are time-consuming and less economical¹³. The present study was conducted to explore the medicinal and aromatic plants of Himachal Pradesh (H.P.) for anti-proliferative activity by using budding yeast *Saccharomyces cerevisiae* as a drug discovery tool. The hill state of India, Himachal Pradesh is very rich in plant diversity, including medicinal and aromatic plants. Medicinal plants are used by village folk to cure many diseases¹⁴. However, there are no reports regarding exploration of medicinal plants of H.P. for anti-proliferative activities. Budding

yeast is being used as a screening tool for the identification of various bioactive molecules in a variety of biological processes¹⁵. Its genome is well sequenced and the existing knowledge of the fundamental processes in yeast is useful for drug discovery. Many cellular processes, including the cell cycle machinery are highly conserved between yeast and humans¹⁶. The budding yeast offers an excellent biological system for such large-scale screening owing to its amenability for rapid growth screen and rich repository of knowledge about cell cycle regulation. Thus, the anti-proliferative compounds identified from yeast are directly relevant for anti-cancer drug discovery in humans, owing to the conservation of cell cycle machinery between yeast and humans, the regulation of which is perturbed in cancer.

MATERIALS AND METHODS

Yeast strain and growth conditions

Wild-type *S. cerevisiae* strain of W303 background (*MATa ura3 can1*) was used for the study. The yeast strain was cultured in YPD media and grown at 30°C.

Collection of plant material

The medicinal plants and their parts (leaves, root, bark, rhizome and flowers) were collected from the following districts of Himachal Pradesh during the season of availability: Solan (altitude 1350 m, temperature 20-30°C), Hamirpur (altitude 738 m, temperature 30-35°C), Una (altitude 369 m, temperature 33-38°C), Mandi (altitude 1044m, temperature 23-29°C). These plants were identified from the herbarium of Dr Y.S. Parmar University of horticulture and Forestry, Nauni, Himachal Pradesh.

Preparation of plant extracts

The plant material used for the study was washed with tap water to remove dirt and soil particles, surface sterilized with 1% H₂O₂ for 10 minutes, and rinsed with sterile distilled water. The sterilized plant material was air dried (1 week) or dried in oven at 40°C for 4-5 days.

Dried plant material was crushed in mortar and pestle to form a fine powder. The fine powder (4 g) was extracted in soxhlet apparatus using ethanol as solvent¹⁷. The extract was dried under vacuum in a rota-evaporator at 40°C and stored at -20°C until further use. The extracts were resuspended in ethanol to a concentration of 0.4 mg/μl for performing anti-proliferative assays.

Anti-proliferative assay for *S. cerevisiae*

The anti-proliferative effect of plant extracts on *S. cerevisiae* growth was studied by agar well diffusion assay^{18 19}. An overnight grown culture of yeast cells (~3× 10⁸ cells) was mixed with molten YPD soft agar (1% agar) and overlaid on precast YPD hard agar (2% agar). The wells were punched with a cork borer (10 mm diameter) and plant extract (24 mg) was loaded into the wells. The assay plates were incubated for 24 h at 30°C. The zone of clearance (mm) was calculated as follows: diameter of zone of clearance of yeast growth measured in two directions and the average of the two values was taken. The standard error was calculated from the results of two independent experiments. To assay the fungicidal or fungistatic nature of anti-proliferative effect of the plant extracts, the cells were carefully scraped from the zone of clearance of yeast growth around the wells and streaked on YPD agar plates and assayed for growth after 48 h of incubation at 30°C.

Phytochemical analysis of plant extracts

The plant extracts exhibiting anti-proliferative effect on yeast growth were tested for the presence of alkaloid, glycosides, terpenoids, steroids, flavanoids, tannins, saponins, reducing sugar, coumarins, proteins and phenols. Phytochemical tests were performed as described^{20 21}.

RESULTS

The present study was conducted for determining the antiproliferative activity of the medicinal and aromatic plants of Himachal Pradesh, using the budding yeast *S. cerevisiae* as a screening system. The selection of traditional medicinal plants for this study was

based on their reported use in cancer treatment with scientific literature or folklore medicine used by the local population of Himachal Pradesh. A total number of 62 plants (including 46 medicinal, 2 aromatic and 14 non-medicinal plants) belonging to 38 different families were collected from different regions of Himachal Pradesh. Different plant material such as leaves, fruit, flower, bark, bulb, rhizome, seeds and stem were collected and their ethanolic extracts were tested for their effect on the growth of *S. cerevisiae* by using well-diffusion assay. Interestingly, several of these plant extracts inhibited the growth of yeast cells, thereby producing zone of clearance around the respective well, which was measured for determining the extent of anti-proliferative activity (Fig.1A). The effect was specific to the tested plant extracts, since solvent alone (control) failed to produce any zone of clearance (Fig.1A). A representative result of anti-proliferative assays are shown in Fig 1A. Amongst the plants tested, extracts from 13 different families of plants were found to exhibit anti-proliferative activity (Table 1). The ethanolic extracts of 17 plants and essential oil of 2 aromatic plants exhibited anti-proliferative properties on the growth of *S. cerevisiae*, while the extracts of remaining 43 plants failed to exhibit any detectable anti-yeast activity (Table 1). On the basis of the anti-proliferative activities quantified by the magnitude of the zone of clearance of yeast growth, the plants were categorized into three classes; 1) Class I- The plants whose extracts showed zone of clearance in the range of 18-26 mm; 2) Class II- Plants extract showing zone of clearance below 18 mm; 3) Class III- Plants whose extracts had no detectable antiyeast activity. The class I plants with strong anti-proliferative activity include *Curcuma longa* (rhizome), *Chrysanthemum indicum* (leaves), *Aegle marmelose* (leaves, fruit) and *Cymbopogon citratus* (oil), with zone of clearance of 26±0.2, 19.5±0.2, 18.5±0.2 and 19±0.2 mm respectively (Table 1). The class 2 included 15 plants whose extracts showed zone of clearance in the range of 11 to 15 mm. The extracts of *A. marmelos* and *C. longa* showed a fungicidal effect on yeast growth. On the other hand, *C. indicum* extracts exhibited the fungistatic effect (Fig. 1B; Table 2). The essential

oil of *R. officinalis* and *C. citratus* oil exhibited a dose-dependent killing effect on yeast cells (Fig. 1B; Table 2). The plants extracts showing strong anti-proliferative activity was subjected to phytochemical screening to analyse the constituents present. As shown in Table 2, terpenoids, steroids and flavanoids were present in all the plants tested. The presence of tannins was confirmed in all the plant extracts, except *R. officinalis* essential oil. The alkaloids and saponins were present in *C. citratus* oil, and absent in rest of the plant extracts (Table 2). All the plants were positive for the presence of glycosides, except *C. longa* extract (Table 2). Reducing sugars were present only in *C. longa* extracts, and oils from *C. citratus* and *R. officinalis* (Table 2). The phytochemical analysis revealed the presence of coumarins in *C. citratus* oil (Table 2). Proteins were absent in all the plant extracts. Phenolics were present in all plant extracts, except *C. citratus* oil (Table 2). Based on these results, the phytochemical compound/s responsible for anti-proliferative activity against yeast cells can be purified by bio-assay guided fractionation.

DISCUSSION

Medicinal and aromatic plants are the largest source of bioactive compounds on Earth that have been used by humans for treating various diseases since ancient times. Plant based products have also played an important role in cancer treatment. Vincristine, vinblastine, and taxol are some examples of plant based drugs being used for cancer therapy. The natural products are easily available, cost effective, and less toxic as compared to the allopathic drugs available for cancer treatment. As a result, extensive screens for plant based medicines have gained impetus world-wide²². Himalayan region is very rich in the medicinally important plants. Different plant material of 35 plant species was reported for medicinal value for treating various diseases by folklore of tribal communities of chhota Bhangal, Western Himalayas²³. The hill state of Himachal Pradesh encompasses a diverse range of medicinal flora, whose potential is not fully explored. *S. cerevisiae* has been used as a system for drug

discovery²⁴, but rarely for screening anti-proliferative compounds²⁵. There are only few reports on the use of budding yeast for anti-proliferative activities²⁶. Our study represents the first report on screening of medicinal and aromatic plants of Himachal Pradesh, India for anti-proliferative activities against *S. cerevisiae*. *C. indicum* (leaves), *C. longa* (rhizome), *A. marmelos* (leaves, fruit) and *C. citratus* (oil) showed strongest anti-proliferative effect against yeast growth. There are no reports of any of these plants for anti-proliferative effect on budding yeast, *S. cerevisiae*. However, their antimicrobial effects have been well studied in the related pathogenic yeast, *C. albicans*²⁷. Hence, the results of the present study are discussed in comparison to these studies. In our study, ethanolic and acetone extracts of leaves of *A. marmelos* exhibited strong anti-proliferative effect, with fungicidal effect on yeast growth, indicating that its phytochemicals kill the yeast cells. *Candida albicans* was found to be the most susceptible fungus to methanolic fruit extract of *A. marmelos* amongst the tested fungal strains²⁸. The petroleum ether extract from the leaves of *A. marmelos* exhibited the highest antifungal efficacy against *C. albicans* and other tested fungal species²⁹. In our study, ethanolic extracts of *C. indicum* leaves, but not its essential oil, exhibited strong anti-proliferative effect with fungistatic effect on yeast growth (Table 1; data not shown). Pradhan *et al.* (2011) reported that essential oils from *C. indicum* showed maximum antifungal activity at 1% v/v against *Candida albicans* (zone of inhibition of 18.0 mm)³⁰. In the present study, ethanolic extracts of *C. longa* exhibited strong fungicidal anti-proliferative effect on yeast growth. Curcuminoids from ethyl acetate extract of *C. longa* rhizomes were found to be antifungal against *Candida albicans*, *C. kruseii* and *C. parapsilosis*^{31 32}. Curcumin, a compound isolated from *C. longa*, caused induction of apoptotic cell death and arrested human bladder cancer cells in G2/M phase³³. However, in the present study, purified curcumin failed to have significant anti-proliferative effect on yeast cells. *C. longa* is documented for anti-inflammatory³⁴, anticancer, antioxidant and hepato-protective effects³⁵. In our study, essential oil of *C. citratus* leaves exhibited strong

anti-proliferative activity, with fungicidal effect on yeast growth. Citral, 3,7-dimethyl-2,6-octadienal, a key component of the essential oil of *C. citratus* was observed to induce apoptosis in acute promyelocytic leukemia cell line (NB4)³⁶. In the present study, essential oil of *R. officinalis* leaves showed moderate anti-proliferative effects with fungicidal effect on yeast growth. Antimicrobial

activity of the essential oil, and methanolic extract of *R. officinalis* against *S. cerevisiae* and *Candida krusei*³⁷ was reported by Tavassoli (2011). The anti-proliferative effect of the polyphenols, carnesol and carnosic acid present in *R. officinalis* was reported against colorectal cancer cells³⁸.

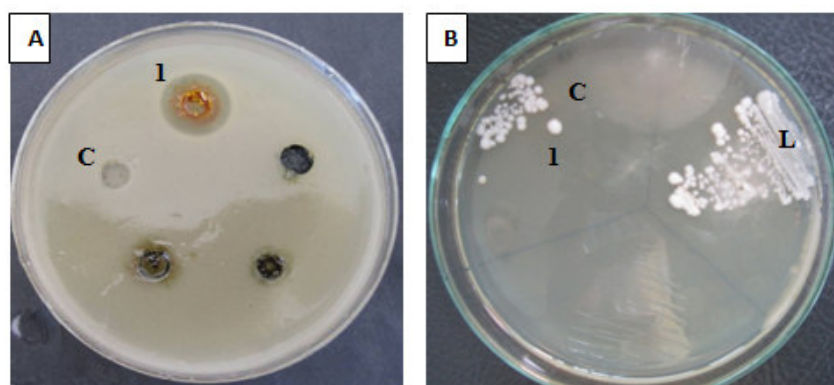


Figure 1

Antiproliferative assay of ethanolic plants extracts against *S. cerevisiae*. A. A representative result of agar well diffusion assay to test the anti-proliferative effects of ethanolic extract of *Curcuma longa*. C: solvent control (ethanol); 1-ethanolic extract of *Curcuma longa*. The lawn of yeast growth and the zone of clearance of yeast growth caused by the ethanolic extract are visible around the well labeled 1. B. Nature of anti-proliferative effect. The yeast cells from the surrounding the well and the zone of clearance (1) shown in panel A were scraped and streaked onto YPD agar media. L: cells scraped from lawn of yeast cells.

Table 1

Antiproliferative activities of plant extracts against *S. cerevisiae*. The zone of clearance of yeast growth (mm) produced by the indicated plant extract are listed.

S.No.	Family of the plant used	Name of the medicinal plant / part used	Zone of clearance of yeast growth (mm) ± SE
1.	Asteraceae	<i>Ageratum conyzoides</i> / L, F	13.5±0.5
		<i>Chrysanthemum indicum</i> / L	19.5±0.5
2.	Apocynaceae	<i>Catharanthus roseus</i> / L	Nd
		<i>Nerium indicum</i> / L, F	Nd
		<i>Carissa spinarum</i> / L	Nd
3.	Lauraceae	<i>Cinnamomum camphora</i> / L	14±0.2
4.	Zingiberaceae	<i>Curcuma longa</i> / R	26±0.2
		<i>Zingiber officinale</i> / R	Nd
5.	Ginkgoaceae	<i>Ginkgo biloba</i> / L	14±0.2
6.	Lamiaceae	<i>Salvia officinalis</i> / Fl	Nd
		<i>Lavendula angustifolia</i> / L	12±0.2
		<i>Mentha citrata</i> / L	Nd
		<i>Vitex nigundo</i> / L	12±0.2
		<i>Rosmarinus officinalis</i> / L*	12±0.2
		<i>Caryopteris divaricata</i> / Fl	12±0.2
		<i>Thymus serpyllum</i> / L	Nd
<i>Colebrookea oppositifolia</i> / L, F	12±0.2		

7.	Menispermaceae	<i>Tinospora cordifolia</i> / L, Sm	Nd
8.	Solanaceae	<i>Withania somnifera</i> / L, F	Nd
		<i>Datura metel</i> / L, F	Nd
		<i>Datura stramonium</i> / L, F	Nd
9.	Acanthaceae	<i>Vasica adathoda</i> / L, F	Nd
10.	Rhamnaceae	<i>Zizyphus mauritiana</i> / L	Nd
11.	Convolvulaceae	<i>Ipomoea carnea</i> / L	12±0.2
		<i>Cuscuta reflexa</i> / Sm	Nd
12.	Rutaceae	<i>Murraya koenigii</i> / L, F	Nd
		<i>Aegle marmelos</i> / L, F	18.5±0.2
		<i>Zanthoxylum armatum</i> / L	Nd
13.	Meliaceae	<i>Azadirachta indica</i> / L	Nd
14.	Santalaceae	<i>Santalum album</i> / L	14±0.2
		<i>Osyris arborea</i> / L	Nd
15.	Malvaceae	<i>Bombax ceiba</i> / Fl	Nd
16.	Phyllanthaceae	<i>Emblica officinalis</i> / L, F	Nd
17.	Fabaceae	<i>Acacia catechu</i> / L, B	Nd
		<i>Cassia fistula</i> / L, F	Nd
		<i>Bauhinia variegata</i> / L, F	Nd
		<i>Indigofera tinctoria</i> / Fl	12±0.2
		<i>Abrus precatorius</i> / S	12±0.2
18.	Musaceae	<i>Musa paradisiaca</i> / L	Nd
19.	Poaceae	<i>Cymbopogon citratus</i> *	19±0.2
20.	Lythraceae	<i>Lawsonia alba</i> / L	Nd
		<i>Punica granatum</i> / L, F	Nd
21.	Brassicaceae	<i>Raphanus sativus</i> / L	Nd
22.	Amaryllidaceae	<i>Allium sativum</i> / Bb	12±0.2
23.	Ranunculaceae	<i>Nigella sativa</i> / S	Nd
24.	Hypericaceae	<i>Hypericum perforatum</i> / Fl	Nd
25.	Urticaceae	<i>Urtica dioica</i> / L	Nd
26.	Cannabaceae	<i>Cannabis sativa</i> / L	Nd
27.	Anacardiaceae	<i>Rhus cotinus</i> / L	14±0.2
28.	Salicaceae	<i>Salix herbacea</i> / L	14±0.2
		<i>Populus Canadensis</i> / L	Nd
29.	Berberidaceae	<i>Podophyllum hexandrum</i> / S	Nd
		<i>Berberis aristata</i> / L	Nd
30.	Betulaceae	<i>Betula utilis</i> / L	Nd
31.	Taxaceae	<i>Taxus bacata</i> / L	Nd
32.	Combretaceae	<i>Terminalia chebula</i> / F	Nd
33.	Araceae	<i>Colocasia esculenta</i> / Sm	Nd
34.	Polygonaceae	<i>Rumex acetosa</i> / L	Nd
35.	Theaceae	<i>Camellia sinensis</i> / L	Nd
36.	Colchicaceae	<i>Gloriosa superba</i> / F	Nd
37.	Asparagaceae	<i>Asparagus racemosus</i> / L	Nd
38.	Euphorbiaceae	<i>Ricinus communis</i> / L	Nd

Plant parts: L- leaves, F- fruit, Fl-Flower, B- bark, Bb-Bulb, R-Rhizome, S-Seeds, Sm-Stem, Nd: no zone of clearance detected; The zone of clearance values are an average from two independent experiments. SE denotes Standard error. * In case of *R. officinalis* and *C. citratus*, essential oil fractions were used in the assay.'

Table 2

Evaluation of the nature of anti-proliferative effect of extracts and essential oils of medicinal plants and their phytochemical screening. Fungicidal indicates no growth of yeast cells, while fungistatic indicates growth of yeast cells when streaked on YPD.

S.no.	Plant name	Part used	Nature of anti-proliferative effect	Test for phytochemicals*										
				1	2	3	4	5	6	7	8	9	10	11
1.	<i>C. indicum</i>	Leaves, flower	S	-	+	+	+	+	+	-	-	-	-	+
2.	<i>C. longa</i>	Rhizome	C	-	-	+	+	+	+	-	+	-	-	+
3.	<i>A. marmelos</i>	Leaves, fruit	C	-	+	+	+	+	+	-	-	-	-	+
4.	<i>C. citratus</i>	Oil	C	+	+	+	+	+	+	+	+	+	-	-
5.	<i>R. officinalis</i>	Oil	C	-	+	+	+	+	-	-	+	-	-	+

*C-fungicidal, S-fungistatic, *1-Alkaloid, 2-Glycosides, 3-Terpenoids, 4-Steroids, 5-Flavanoids, 6-Tannins, 7-Saponins, 8-Reducing Sugars, 9-Coumarins, 10-Proteins, 11-Phenols*; - : No reaction in the test; + : Positive reaction in the test for the indicated phytochemical.

CONCLUSION

The study was conducted to screen for anti-proliferative activities of different medicinal and aromatic plants of Himachal Pradesh. The plants showing potent anti-proliferative activity can be used for the isolation of active phyto-compounds. These compounds can further be tested against human cancer cell lines for anticancer therapy, based on the fact that cell

cycle genes between *S. cerevisiae* and humans are conserved. Further, our study validates the use of *S. cerevisiae* as an effective and economical tool for screening anti-proliferative activities from medicinal and aromatic plants towards anti-cancer drug discovery.

REFERENCES

- Sakthivel K.M. and C. Guruvayoorappan, *Biophytumsensitivum*: Ancient medicine, modern targets. J Adv Pharm Technol Res, 3 (2): 83–91, (2012).
- Mazid M., Khanb T.A. and F. Mohammad, Medicinal Plants of Rural India: A Review of Use by Indian Folks. Indo Global Journal of Pharmaceutical Sciences, 2 (3): 286-304, (2012).
- Newman D.J., Cragg G.M. and K.M. Snader, The influence of natural products upon drug discovery. Nat Prod Rep.17 (3): 215-34, (2000).
- Hairborne J.B., The Flavonoids Advances in Research since 1986, Chapman & Hall: London, UK, 1–676, (1993).
- Musa M.A., Cooperwood J.S. and M.O. Khan, A review of coumarin derivatives in pharmacotherapy of breast cancer. Curr Med Chem, 15 (26): 2664-79, (2008).
- Surh Y.J., Cancer chemoprevention with dietary phytochemicals. Nat Rev Cancer, 3 (10): 768-80, (2003).
- Schlipkoter U. and A. Flahault, Communicable diseases: achievements and challenges for public health. Public Health Reviews, 32: 90-119, (2010).
- Hammer K.A., Carson C.F. and T.V. Riley, Antimicrobial activity of essential oils and other plant extracts. J. Appl. Microbiol, 86: 985–990, (1999).
- Cragg G.M. and D.J. Newman, Plants as a source of anti-cancer agents. J Ethnopharmacol, 100: 72-90, (2005).
- Weinstein J.N., Myers T.G., O'Connor P.M., Friend S.H., Fornace A.J., Kohn K.W., Fojo T., Bates S.E., Rubinstein L.V., Anderson N.L., Buolamwini J.K., Van Osdol W.W., Monks A.P., Scudiero D.A., Sausville E.A., Zaharevitz D.W., Bunow B., Viswanadhan V.N., Johnson G.S.,

- Wittes R.E. and K.D. Paull, An information-intensive approach to the molecular pharmacology of cancer. *Science*, 275: 343-349, (1997).
11. Craig W.J., Health-promoting properties of common herbs. *Am. J.Clin. Nutr*, 70: S491-S499, (1999).
 12. Bhanot A., Sharma R. and N. Malleshappa, Natural sources as potential anticancer agents: A review. *International Journal of Phytomedicine*, 3: 9-26, (2011).
 13. Pezzuto J.M., Plant-derived anticancer agents, *Biochemical pharmacology*, 53 (2): 121–133, (1997).
 14. Uniyal S.R., Singh K.N., Jamwal P. and B. Lal, Traditional use of medicinal plants among the tribal communities of ChhotaBhangal. *Western Himalaya. Journal of Ethnobiology and Ethnomedicine*, 2:14, (2006).
 15. Qaddouri B., Guaadaoui A., Bellirou A., Hamal A., Melhaoui A., Brown G.W. and M. Bellaoui, The Budding Yeast ‘*Saccharomyces cerevisiae*’ as a Drug Discovery Tool to Identify Plant-Derived Natural Products with Anti-Proliferative Properties. *Ecam*, 10: 1-5, (2009).
 16. Simon J.A. and A. Bedalov, Yeast as a model system for anticancer drug discovery. *Nature Reviews Cancer*, 4: 481-487, (2004).
 17. Hawthorne S.B., Grabanski C.B., Martin E. and D.J. Miller, Comparisons of Soxhlet extraction, pressurized liquid extraction, supercritical fluid extraction and subcritical water extraction for environmental solids: recovery, selectivity and effects on sample matrix. *Journal of Chromatography A*, 892 (1–2): 421–433, (2000).
 18. Perez C., Paul M. and P. Bazerque, Antibiotic assay by agar well diffusion method. *Acta Biol Med Exp*, 15: 113-115, (1990).
 19. Kim K.J., Sung W.S., Suh B.K., Moon S.K., Choi J.S., Kim J.G. and D.G. Lee, Antifungal activity and mode of action of silver nanoparticles on *Candida albicans*. *Biometals*, 22 (2): 235-242, (2009).
 20. Harborne J.B., *Phytochemicals Methods: a guide to modern techniques of plant analysis*, 1st ed. Chapman and Hall Ltd: London, UK, 49-188, (1973).
 21. Sofowara A., *Medicinal plants and traditional medicine in Africa II*, ed., Spectrum Books Ltd: Ibadan, Nigeria, 289, (1993).
 22. Solowey E., Lichtenstein M., Sallon S., Paavilainen H., Solowey E. and H.L. Galski, Evaluating Medicinal Plants for Anticancer Activity. *The Scientific World Journal*, 2014: Article ID 721402, 12 pages doi:10.1155/2014/721402, (2014).
 23. Uniyal S.R., Singh K.N., Jamwal P. and B. Lal, Traditional use of medicinal plants among the tribal communities of ChhotaBhangal. *Western Himalaya. Journal of Ethnobiology and Ethnomedicine*, 2:14, (2006).
 24. Barberis A.A., Gunde T., Berset C., Audetat S. and U.L. thi, Yeast as a screening tool. *Drug discovery today: technologies*, 2(2): 187-192, (2005).
 25. Gao G., Chen L. and C. Huang, Anti-cancer Drug Discovery: Update and Comparisons in *Yeast*, *Drosophila*, and *Zebrafish*. *Curr Mol Pharmacol*, 7(1): 44-51, (2014).
 26. Saboo S., Khadabadi S. and G. Tapadiya, In vitro Evaluation of Antimitotic, Antiproliferative, DNA fragmentation and Anticancer activity of Chloroform and Ethanol extracts of *Revia hypocrateriformi*. *Asian Pacific Journal of Tropical Disease*, 2012: S503-S5082, (2012).
 27. Sharanappa R. and G.M. Vidyasagar, Anti-Candida activity of medicinal plants. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(4): 9-16, (2013).
 28. Parihar N. and S. Kumar, Study of antifungal potential of *Aegle marmelos*: a medicinal plant. *International journal of plant, animal and environmental sciences*, 3(1): 126-129, (2013).
 29. Kothari S., Mishra V., Bharat S. and S.D. Tonpay, Antimicrobial activity and phytochemical screening of serial extracts from leaves of *Aegle marmelos* (Linn.). *Acta Poloniae Pharmaceutica n Drug Research*, 68(5): 687-692, (2011).

30. Pradhan C.K., katara A., Singh, Mishra G. and R.L. Khosa, Chemical characterization by GLC & GC-MS and antimicrobial activity of essential oil from leaves of *Chrysanthemum indicum* Linn. International Journal of Pharm Tech Research, 3(1): 175-179, (2011).
31. Roth G.N., ChandraA. and M.G. Nair, Bioactive Natural Products. *J. Nat. Prod.*, 61(4): 542–545, (1998).
32. Harit J., Barapatre A., Prajapati M., Aadil K.R. and S. Senapati, Antimicrobial activity of rhizome of selected *Curcuma* variety. *Int. J. LifeSc. Bt & Pharm. Res.*, 2(3): 1-9, (2013).
33. Karunagaran D., Rashmi R. and T.R.S. Kumar, Induction of Apoptosis by Curcumin and Its Implications for Cancer Therapy. *Current Cancer Drug Targets*, 5:117-129, (2005).
34. Jurenka J.S., Anti-inflammatory properties of curcumin, a major constituent of *Curcuma longa*: a review of preclinical and clinical research. *Altern Med Rev.*, 14(2): 141-53, (2009).
35. Labban L., Review article Medicinal and pharmacological properties of Turmeric (*Curcuma longa*): A review. *Int J Pharm Biomed Sci.*, 5(1): 17-23, (2014).
36. Hailong X., LiangW., SongQ., ChenX., ChenX. and J. Hong, The in vitro study of apoptosis in NB4 cell induced by citral Novel Bioactivities of *Curcuma longa* Constituents. *Cytotechnology*. 65(1): 49–57, (2013).
37. Tavassoli S., Mousavi S.M., Emam-Djomeh Z. and S.H. Razavi, Comparative Study of the Antimicrobial Activity of *Rosmarinus officinalis* L. Essential Oil and Methanolic Extract. *Middle-East Journal of Scientific Research*, 9(4): 467-471, (2011).
38. Visanji J.M., Thompson D.G. and P.J. Padfield, Induction of G2/M phase cell cycle arrest by carnosol and carnosic acid is associated with alteration of cyclin A and cyclin B1 levels. *Cancer Lett.*, 237(1): 130-136, (2006).