



POTENTIAL OF *NOSTOC MUSCORUM* FOR THE DECOLORIZATION OF TEXTILE DYE RGB-RED

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ABSTRACT

Nostoc muscorum, a dominant kind of cyanobacterial bloom was found as a viable biomass for decolorization of synthetic RGB-Red dye. Batch studies revealed the capacity of algal species in dye degradation which was dependent on initial pH (2-12), initial dye concentration (10 mg L^{-1} - 50 mg L^{-1}), temperature (20°C , 30°C , 40°C) and contact time (5 min-180 min). Maximum growth of algae was observed at 50 mg L^{-1} dye concentration. Optimal pH for maximum decolorization was determined at pH 3. The data obtained fitted well with Langmuir adsorption isotherm. This study confirmed the potential of blue green algae *Nostoc muscorum* to degrade textile dye, suggesting the possibility of its application in developing an oxidation pond system for textile effluent treatment at pilot scale study.

KEYWORDS: *Nostoc muscorum*, RGB-Red dye, optimization, Langmuir adsorption, Bioremediation



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INTRODUCTION

Water contamination with synthetic dyes is a severe environmental issue and represents a threat to human well being. Worldwide over 10,000 different dyes and pigments are used in dyeing and printing industries. The total world colorant production is estimated to be 8,00,000 tons per year and at least 10% of the used dyestuffs enter the environment through wastes¹. Dye is used to impart color to textiles, paper, leather, and other materials in such a manner that the coloring is not readily altered by washing, heat, light etc. Synthetic dyes are used especially in the textile and dyeing industries because of their ease and their cost effectiveness in synthesis, firmness, high stability to light, temperature, detergent, microbial attack and variety in color compared with the natural dyes². This has resulted in the discharge of highly colored effluents that affect water transparency and gas solubility in water bodies³. In the process of dyeing, about 15–20% of the dyes used for dyeing does not bind to the fibers and are lost in the effluent⁴. The textile industry produces large volumes of colored wastewater due to the use of synthetic dyes in dyeing products and low levels of dye fiber fixation⁵. Synthetic dyes have complex molecular structure that contain variety of functional groups which renders them stable against sunlight, oxidizing agents or microbial attack and also are responsible for the toxicity to some aquatic animals⁶. Azo dyes are the main constituents of such pollution because of their wide applicability and usage and therefore these are present in large quantity in industrial effluents⁷. More than 2000 azo dyes are produced annually constituting 50% of total dyes⁸. Reductive cleavage of azo dyes, which comprises about 70% of all dyes used, also results in the production of amines that are mutagenic to human and are also retained in the anaerobic compartment of the lower intestine by intestinal micro flora after ingestion of azo dyes⁹. Therefore, industrial effluents containing dyes must be treated prior to their discharge into the environment. Many physicochemical methods are used for the treatment of these dyes but these methods are limited by many factors like post treatment process and cost¹⁰. Also these processes require labors and have disposal problems^{11, 12}. Hence, in recent year's interest has been

focused on different biological treatment systems which are new, economical and involves the use of easily available micro-organisms like algae, bacteria, yeast and fungi from the removal of dyes from wastewater. These biological treatments are gaining interests due to their efficiency, low cost and environment friendly nature^{13, 14, 15, 16, 17, 18}. Algae have been shown to be capable of removing color from various dyes through mechanisms such as biosorption, bioconversion and biocoagulation. Algae is considered as promising biosorbents¹⁹ due to its high sorption capacity and available in unlimited amount²⁰ both in fresh water and salt water. For instance Vishal shah et al.²¹ showed that *Phormidium valderianum*, a marine cyanobacteria can remove up to 90% of textile dyes Acid red 119 and Direct black 115 from solutions in the basic pH range. A marine cyanobacteria *Oscillatoria formosa* NTDM02 was used to decolourize Amido black dye²². Also, free and immobilized cyanobacteria *Oscillatoria brevis* and *Westiellopsis prolifica* were used to remove color and other nutrients from dye industry effluents¹⁰. Some species of *Oscillatoria*, *C. pyrenoidosa* and *C. vulgaris* have ability to degrade azo dyes and decolorize dye wastewater^{23, 24, 25, 26}. *Nostoc muscorum* is a nitrogen fixing, photoautotrophic cyanobacteria. RGB-Red dyes contain a monoazo chromophore which gives color to the dye. This azo group is degraded by *Nostoc muscorum* and also the nitrogen present in the azo group is utilized by the algae for its growth. The aim of the present work is to establish the optimum experimental conditions for the algal growth and the maximum decolorization of RGB-Red dye from aqueous medium.

MATERIALS AND METHODS

Algal biomass and growth conditions

The algal strain *Nostoc muscorum* was cultured in 500 mL CHU-10 media²⁷ contained in cotton stopped 1000 mL Erlenmeyer flasks at temperature 28⁰ C, providing 24 hrs fluorescent illumination (40watt, white tube light). The basal culture medium and

glassware were sterilized at 1.0546 kg/cm² pressure and 121^o C for 15 min before inoculation. The algal cells at late exponential growth phase (after 14 days) were harvested by centrifugation at 5,000 rpm for 15 min. In order to remove any algal exudates present in the initial alga pellet, the collected cells were washed two times with sterile deionized water through centrifugation and finally resuspended in CHU-10 media to obtain a concentrated algal suspension. The cell growth was monitored at 665 nm by UV-Visible spectrophotometer (Shimadzu UV-1650).

Dye and Chemicals

All the chemicals used were of highest purity available and of analytical grade. The dye used in this study was RGB-Red dye collected from the textile dyeing industry located at Faridabad, India. The chemical structure and general data of the dye is given in Table 1. A stock solution (200 mg L⁻¹) of RGB-Red dye was prepared in distilled water. The dye concentration in supernatant solution was determined by measuring the absorbance with UV-Visible Spectrophotometer (Shimadzu UV-1650) at maximum absorption wavelength (λ_{max} = 520 nm).

Table 1
Commercial name and the functional group of the dye used in the study

| Dye | Commercial name | Type | Chromophore | Chemical structure | Functional group |
|---------|-----------------|----------|-------------|--------------------|--------------------------------------|
| RGB-Red | RGB-Red | Reactive | Monoazo | Vinyl sulfone | Monochloro triazine bifunctional dye |

Decolorization Assay

The dye concentration in supernatant solution was determined at characteristic wavelength (λ_{max} =520) before and after treatment by double beam UV-VIS spectrophotometer (Shimadzu UV-1650). The efficiency of dye removal was expressed as the percentage ratio of decolorized dye concentration to that of initial one.

$$\text{Color removal (\%)} = C_i - C / C_i \times 100$$

where, C_i is the initial concentration of dye and C is the concentration of dye at time t .

The amount of dye bound to the algal biomass was determined using the following formulae

$$q_{eq} = (C_o - C_{eq}) \times V / W \times 1000$$

where, q_{eq} is the amount of dye adsorbed (mg g⁻¹), C_o is the initial concentration of dye (mg L⁻¹), C_{eq} is the equilibrium concentration of dye (mg L⁻¹), V is the volume of the solution (mL) and W is the dry weight of the cells (g).

Optimization of parameters

To evaluate the decolorization efficiency of algal biomass, batch experiments were performed at various initial dye concentration (10 mg L⁻¹-50 mg L⁻¹), temperature (20 °C, 30 °C, and 40 °C), contact time (5 min-180 min) and pH values (2-11). The pH was adjusted using 0.1N NaOH and 0.1N HCl solutions. The experiments were conducted in 250 mL

Erlenmeyer flasks containing 50 mL media, inoculated with homogenized algal inoculums having 0.4 O.D. at 665 nm and required amount of RGB-Red dye.

Statistical Analysis

All the experiments were performed in triplicates and the values are represented as mean \pm SD. Repeated measures one way ANNOVA was used to determine the statistical significance during time course studies with RGB-Red dye at $p < 0.05$ using SPSS 20. The adsorption isotherms were tested using regression analysis. The performance of *Nostoc muscorum* for dye removal was also statistically evaluated.

RESULTS AND DISCUSSION

UV-Visible spectra

Fig. 1 shows a UV-Visible spectrum of RGB-Red dye solution before and after treatment with algae. The spectrum of the dye in the visible region displays main peak at 520 nm. The decrease of the absorbance peak in this figure indicated that there was almost complete removal of dye. This decolorization of dye may be due to the adsorption as well as bioaccumulation of it by the algal cells²⁸.

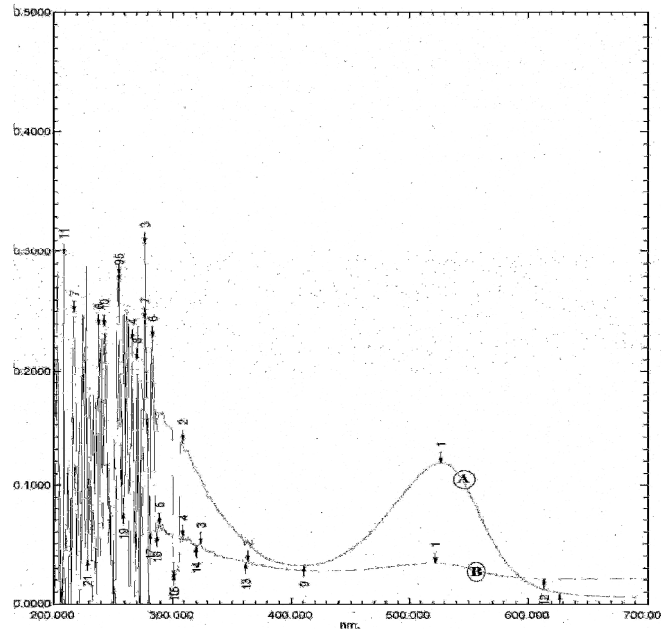


Figure 1
UV-Visible spectrum of RGB-Red dye. A depicts dye solution before treatment and B depicts dye solution treated with algae

Effect of Dye Concentration on growth of algae

The experiment was carried out to evaluate the growth of *Nostoc muscorum* in different concentration of RGB-Red dye (10 mg L⁻¹ to

90 mg L⁻¹). Fig. 2 depicts that the maximum growth of cells was found at 50 ppm on 6th day which is 1.03 times more than that of control.

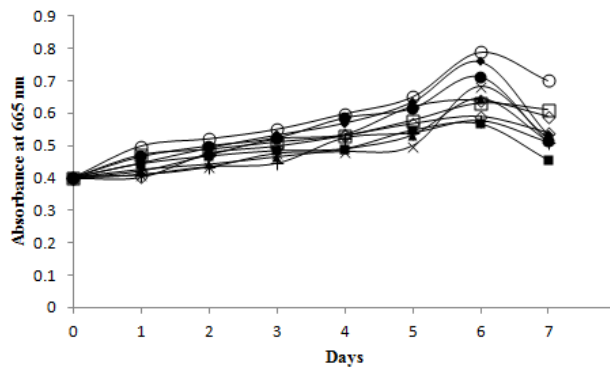


Figure 2
Effect of RGB-Red concentrations on growth. [Algae] = 0.4 OD
 (♦ control, ■ 10 mg L⁻¹, ▲ 20 mg L⁻¹, × 30 mg L⁻¹, ● 40 mg L⁻¹, ○ 50 mg L⁻¹,
 △ 60 mg L⁻¹, □ 70 mg L⁻¹, ◇ 80 mg L⁻¹, + 90 mg L⁻¹)

Effect of initial pH on decolorization

Removal of dye by the blue-green algae is also dependent on the pH of the dye solution^{29, 30}. Fig. 3 shows the ability of *Nostoc muscorum* in removal of RGB-Red dye at different pH ranging from 2-11. Algal cells were found dead at pH 2. Highest percentage of dye removal (95%) was attained at pH 3 with an initial dye concentration of 40 mg L⁻¹ after 3 h. It showed that an increase in pH

from 4 to 11, leads to decrease in decolorization rate. Maximum percent removal of dye at lower pH was because of the electrostatic interaction between negatively charged dye and positively charged protonated functional groups (e.g. carboxyl and amino groups) present on algal cell wall. During the study it was also found that the pH of the solutions below 7 and above 9 was

adjusted in the range of pH 7.5-8.5 after addition of algal inoculums within few hours. This increase and decrease in the pH was

may be due to production of metabolites and enzymes by the algae in the medium during the degradation process of dye²².

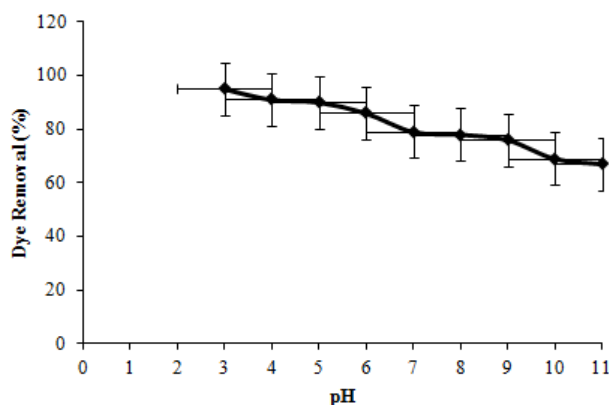


Figure 3
Effect of pH on the decolorization of RGB-Red
[Algae]=0.4 OD, [RGB-Red] = 40 mg L⁻¹

Effect of contact time and initial dye concentration on decolorization

The effect of contact time on decolorization was carried out to evaluate how quickly the sorption equilibrium was achieved between the algae and the solution they were immersed in. The equilibrium time required to attain maximum sorption (Fig. 4) under static condition was found 30 min due to the rapid adsorption of dye molecule on the surface of algae followed by slower rate of dye decolorization which may be because of further diffusion of the dye molecules from

aqueous phase to the solid phase of the algal cells³¹. Results illustrate that the amount of dye bound to the algal biomass (q_{eq}) increased with increasing dye concentration and within 30 min equilibrium was achieved. The q_{eq} value increased from 5.97 mg g⁻¹ to 31.53 mg g⁻¹ of biomass with increase in initial concentration of dye from 10 mg L⁻¹ to 50 mg L⁻¹. It was also proved by using one way repeated measure analysis of variance that there was a statistical difference ($p > 0.05$) in the uptake of dye by algae at different dye concentration with respect to time.

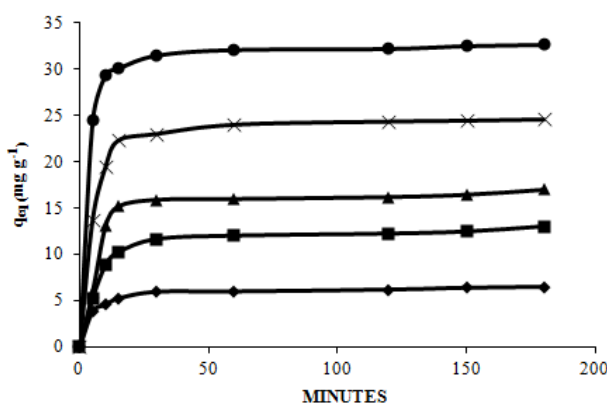


Figure 4
Effect of contact time and initial dye concentration on decolorization of RGB-Red. [Algae]=0.4 OD, pH= 3.0 (♦ 10 mg L⁻¹, ■ 20 mg L⁻¹, ▲ 30 mg L⁻¹, × 40 mg L⁻¹, ● 50 mg L⁻¹)

Effect of temperature on decolorization

The effect of temperature on the decolorization capacity of *Nostoc muscorum*

was studied at 20 °C, 30 °C and 40 °C. It can be seen in Fig. 5 that the decolorization

efficiency increased from 67% to 83% with an increase in temperature from 20 °C to 30 °C while from at 40 °C there was 40% decline in decolorization capacity after three hours of

exposure. This decrease in the color removal efficiency can be attributed to the loss of cell viability at higher temperature^{32, 33}.

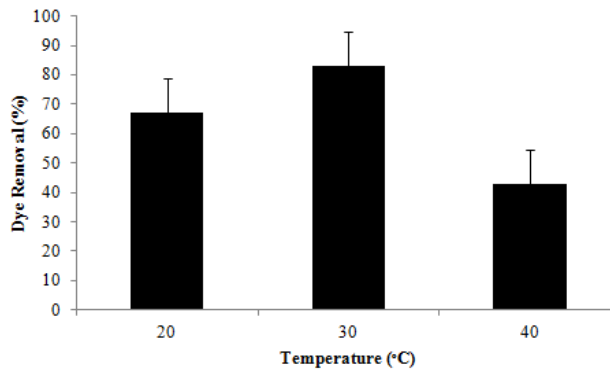


Figure 5

Effect of temperature on decolorization on RGB-Red. [Algae]=0.4 OD, pH=3.0.

Effect of repeated uses

Repeated batch operations were performed to examine the reusability of *Nostoc muscorum* in the removal of RGB-Red dye. After the four repeated use of *Nostoc muscorum*, its decolorization efficiency was found the same

as it showed on the first day (Fig. 6). This may be due to an adaptation effect, since the cells were repeatedly exposed to the dye. The results obtained were similar to that observed in *Cosmarium* species²⁸.

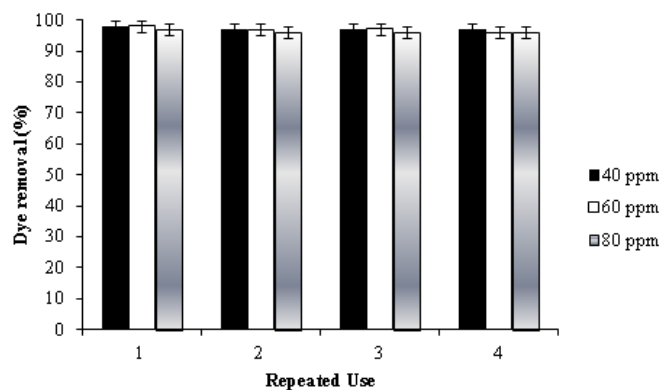


Figure 6

Effect of repeated use of algal biomass on decolorization

Langmuir Adsorption isotherm

In the present study, the experimental data of RGB-Red dye and algae equilibrium isotherm was established using Langmuir isotherm. Langmuir sorption model projects the maximum uptake values, which could not be attained in the experiments. Langmuir isotherm is expressed by the following equation³⁴:

$$1/q_{eq} = (1/bq_m)(1/C_{eq}) + 1/q_m$$

where, C_{eq} is the equilibrium concentration of dye solution and q_{eq} is the adsorbed dye on the algal cells. q_m and k_a are constants corresponding to the adsorption capacity and rate of adsorption, respectively. Values of q_m and K_a for the adsorption of dye onto the algal cells are determined from the slope and intercept plot of $1/q_{eq}$ vs $1/C_{eq}$.

Table 2
Langmuir isotherm constant for the decolorization of RGB-Red dye by *Nostoc muscorum*

| Algae | q_{\max} (mg g ⁻¹) | K_a | R^2 |
|------------------------|----------------------------------|-------|-------|
| <i>Nostoc muscorum</i> | 454.54 | .007 | .9959 |

The adsorption isotherm of the RGB-Red was found to be linear over the entire concentration range and the correlation coefficient (R^2) was less than one and greater than zero illustrating that the algae is supportive for the adsorption of RGB-Red dye.

Statistical assessment of the performance of *Nostoc muscorum* for RGB-Red dye removal

Since *Nostoc muscorum* were cultured in different batches at different time, it is necessary to have a statistical assessment of the performance of the *Nostoc muscorum* in terms of precision between batch to batch, repeated use of algae and harvested algal cells for different batches (Table 3). A set of five different batches (A-1, A-2, B-3, B-4, Z-5) were studied to obtain the maximum dye

removal capacity for each batch after 5 days of incubation of 50 mg L⁻¹ dye concentration with the algae. In precision between repeated use of algae, the dye removal capacity of *Nostoc muscorum* were found to be 470.66 mg g⁻¹ and 402.16 mg g⁻¹ with maximum relative standard deviation (R.S.D) of 6.21% confirming the reproducibility in repeated uses. However, the precision significantly varied with the third batch of algae which was harvested for different batches at different time as was evident from the high value of R.S.D (20.40%). While comparing the precision between the batch to batch variability, repeated use of algae and harvested algal cells for different batches, the chance of imprecision is possible as the cells were cultured at different time intervals.

Table 3
Precision of experimentally determined dye removal capacity of *Nostoc muscorum*

| Name | <i>Nostoc muscorum</i> | <i>Nostoc muscorum</i> | <i>Nostoc muscorum</i> | <i>Nostoc muscorum</i> | <i>Nostoc muscorum</i> |
|--|------------------------|------------------------|------------------------|------------------------|------------------------|
| | mg g ⁻¹ | mg g ⁻¹ | mg g ⁻¹ | mg g ⁻¹ | mg g ⁻¹ |
| Batch to batch variability | | | | | |
| A-1 | 447 | A-2 | 490 | B-3 | 390 |
| | 450 | | 495 | | 393 |
| | 453 | | 489 | | 387 |
| Mean | 450 | | 491.33 | | 390 |
| | | | | | 414.33 |
| S.D | 3 | | 3.21 | | 3 |
| | | | | | 4.04 |
| R.S.D (%) | 0.67% | | 0.65% | | 0.77% |
| | | | | | 0.98% |
| | | | | | 1.34% |
| Precision between repeated use of algae | | | | | |
| A-1 | 450 | B-3 | 390 | | |
| A-2 | 491.33 | B-4 | 414.33 | | |
| Mean | 470.66 | | 402.16 | | |
| S.D | 29.22 | | 17.20 | | |
| R.S.D (%) | 6.21% | | 4.28% | | |
| Precision between the harvested algal cells for different batches | | | | | |
| A | 470.66 | | | | |
| B | 402.16 | | | | |
| Z | 310.33 | | | | |
| Mean | 394.38 | | | | |
| S.D | 80.44 | | | | |
| R.S.D (%) | 20.40% | | | | |

CONCLUSION

In conclusion, the present study showed that the blue green alga *Nostoc muscorum* was able to grow in RGB-Red dye, although its growth was less in control. The amount of dye bound to the algal biomass increased with the dye concentration. Maximum adsorption capacity was found to be 31.53 mg g⁻¹ algae at an initial dye concentration of 50 mg L⁻¹ in 30 min. About 95% dye was removed at pH 3 after 3 h. Therefore it may be suggested that *Nostoc muscorum* is an efficient source for

removal of synthetic dyes from effluents and could be adopted as a cost effective approach for decolorization of dyes by developing oxidative pond at pilot scale.

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REFERENCES

1. Krishnaveni M & Kowsalya R. Characterization and Decolorization of Dye and Textile effluent by Laccase from *Pleurotus Florida*. *Int. J. Pharm. Bio Sci.* 2: 117-123, (2011).
2. Couto SR. Dye removal by immobilised fungi. *Biotech. Adv.* 27: 227-235, (2009).
3. Banat IM, Nigam P, Singh D, Marchant R. Microbial decolorization of textile dye containing effluents: a review. *Bioresour. Technol.* 58: 217-227, (1996).
4. Husain Q. Peroxidase mediated decolorization and remediation of wastewater containing industrial dyes: a review. *Environ. Sci. Biotechnol.* 9(2): 117-140, (2010).
5. Hornik M, Sunovska A, Partelova D, Pipiska M, Augustin J. Continuous sorption of synthetic dyes on dried biomass of microalga *C. Vulgaris*. *Chemical Papers.* 67(3): 254-264, (2013).
6. Aksuz. Application of biosorption for the removal of organic pollutant: A review. *Process. Biochem.* 40: 997-1026, (2005).
7. Lamia AM. Toxicity Assessment of Textile dyes via oxidative stress hypothesis for Iraqi textile workers. *Int. J. Pharm. Bio Sci.* 4(4): (B) 577 – 587, (2013).
8. Kamble K & More M. Bacterial decolorization of yellow dye obtained from textile industry effluents. *Int. J. Pharm. Bio Sci.* 4(4): (B) 763 – 769, (2013).
9. Hanan HO. Algal decolorization and degradation of Monoazo and Diazo Dyes. *Pak. J. Biol. Sci.* 11(10): 1310-1316, (2008).
10. Subramaniyam V& Chockaiya M. Treatment of Dye Industry effluent using free and immobilized Cyanobacteria. *J. Bioremed. Biodeg.* 3:10, (2012).
11. Chen GQ, Zeng GM, Tang L, Du CY, Jiang XY, Huang GH, Liu HL, Shen GL. Cadmium removal from simulated wastewater to biomass byproduct of *Lentinus edodes*. *Bioresour Technol.* 99: 7034-7040, (2008).
12. Tang L, Zeng GM, Shen GL, Li YP, Zhang Y, Huang DL. Rapid detection of picloram in agricultural field samples using a disposable immunomembranebased electrochemical sensor. *Environ. Sci. Technol.* 42: 1207-1212, (2008).
13. Quader AKM A. Treatment of textile waste water with chlorine: an effective method. *Chem. Eng. Res. Bull.* 14(1): 59-63, (2010).
14. Sing LL, Chu WL, Phang SM. Use of *Chlorella vulgaris* for bioremediation of textile wastewater. *Bioresour Technol.* 101(19): 7314–7322, (2010).
15. Chu WL, See YC, Phang SM. Use of immobilised *Chlorella vulgaris* for the removal of colour from textile dyes. *J. Appl. Phycol.* 21: 641–648, (2009).
16. Gomathi LD, Kumar SG, Reddy KM. Photo fenton like process Fe⁺³/ (NH₄)₂S₂O₈/ UV for the degradation of Di azo dye congo red using low iron concentration. *Cent. Eur. J. Chem.* 7 (3): 468-477, (2009).

17. Mauskar JM. Advanced Methods for the Treatment of Textile Industries Effluents. Dr. B. Sengupta, Member Secretary, Central Pollution Control Board, Delhi.p.2. (2007).
18. Aslam MM, Baig MA, Hassan I, Qazi IA, Malik M, Saeed H. Textile waste water characterization and reduction of its COD and BOD by oxidation. *Elec. J. Env., Agricult Food Chem.* 3(6): 804-811, (2004).
19. Veligo F, Beolchini F. Removal of metals by biosorption—A review. *Hydrometallurgy* 44: 301, (1997).
20. Klimmek S, Stan HJ, Wilke A, Bunke, G, Buchholz R. Comparative Analysis of the Biosorption of Cadmium, Lead, Nickel and Zinc by Algae. *Environ. Sci. Technol.* 35: 4283–4288, (2001).
21. Vishal S, Garg N, Datta M. An integrated process of textile dye removal and hydrogen evolution using cyanobacterium, *Phormidium valderianum*. *World J. Microbiol Biotechnol.* 17: 499-504, (2001).
22. Mubarak Ali D, Suresh A, Kumar PR, Gunasekaran M, Thajuddin N. Efficiency of textile dye decolorization by marine cyanobacteria *Oscillatoria formosa* NTDM02. *Afr. J. Basic. Appl. Sci.* 3(1): 09-13, (2011).
23. Mona S, Kaushik A, Kaushik CP. Biosorption of reactive dye by waste biomass of *Nostoc linckia*, *Ecol. Eng.* 37: 1589–1594, (2010).
24. Mustafa M, Sheekhs E, Gharieb MM, Abou-El-Souod GW. Biodegradation of dyes by some green algae and cyanobacteria. *Int. Biodeterior. Biodegrad.* 63(6): 699–704, (2009).
25. Ertugrul S, Bakur M, Donmez G. Treatment of dye-rich wastewater by an immobilized thermophilic cyanobacterial strain: *Phormidium* sp. *Ecol. Eng.* 32: 244–248, (2008).
26. Acumer E, Dilek FB. Treatment of tectilon yellow 2G by *Chlorella vulgaris*. *Proc. Biochem.* 39: 623–631, (2004).
27. Chu SP. The influence of the mineral composition of the medium on the growth of planktonic algae. Part I. Methods And culture media. *J.Ecol.* 30, 284-325, (1942).
28. Daneshwar N, Ayazloo M, Khataee AR, Pourhassan M. Biological decolorization of dye solution containing malachite green by microalgae *Cosmarium* species. *Bioresour. Technol.* 98(6): 1176-82, (2006).
29. Aksu Z & Tezar S. Biosorption of reactive dyes on the green alga *Chlorella Vulgaris*. *Process Biochem.* 40: 1347-1361, (2005).
30. Kumar KV, Sivanesan S, Ramamurthi V. Adsorption of malachite green onto *Pithora* sp., a fresh water algae: equilibrium and kinetic modeling. Department of chemical Engineering, A.C. Anna University, Chennai, India, (2005).
31. Abd-El-Kareem MS, Md Taha H. Decolorization of malachite green and methylene blue by two microalgal species. *Int. J. Chem. Enviro. Eng.* 3(5), (2012).
32. Chang JS, Chou YP, Chen SY. Decolorization of azo dyes with immobilized *Pseudomonas luteola*. *Process Biochem.* 36: 757-763, (2001).
33. Saratale RG, Saratale GD, Chang JS, Govindwar SP. Ecofriendly decolorization and degradation of Reactive Green 19A using *Micrococcus glutamicus* NCIM-2168. *Bioresour. Technol.* 110: 3897-3905, (2009).
34. Bulent A, Fatih T. Optimum isotherm parameters for reactive azo dye onto pistachio nut shells: comparison of linear and non linear methods. *Pol. J. Environ. Stud.* 22(4): 1007-1011, (2013).