

**ANTICANCER ACTIVITY OF MILKY MUSHROOM (*CALOCYBE INDICA* VAR.APK2)
AGAINST A549 HUMAN LUNG CANCER CELL LINE STUDIES****A.PARVEEN NISHA*¹ AND R.KUMUTHAKALAVALLI¹**¹ Department of Biology, Gandhigram Rural Institute-Deemed University, Gandhigram 624 302.**ABSTRACT**

Methanolic extract of fruiting bodies of *Calocybe indica* Var.APK2 was evaluated for their anticancer activity against lung carcinoma (A549) cell line. The preliminary phytochemical characterization of the extract using FT-IR recorded the presence of different bioactive compounds, namely alcohols, phenols, aromatics, alkanes, unsaturated esters, carboxylic acid and alkyl halides in their major peaks. The antioxidant potential of *Calocybe indica* in terms of DPPH radical scavenging assay was evolved as 60% and hydroxyl radical scavenging activity as 66 % using ascorbic acid as standard. The IC₅₀ value of the sample to A549 lung cancer cell line was expressed as 4.9 µg/ml. Apoptosis profile of *C.indica* was indicated as 80.13% and maximum rate of inhibition (54%) was observed in S phase. These results indicate that *C. indica* possesses bioactive metabolites and phytochemicals capable of scavenging free radicals and inhibit the growth of A549 lung cancer cell line and suggest their potential for medicinal purpose.

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INTRODUCTION

Cancer remains to be one of the leading causes of death around the world. Many factors may increase the risk of cancer such as inadequate diet, excessive light intensities and toxins¹. The major characteristic is the lack of control of the cell proliferation, differentiation, and death including invading organs and tissues. There are many difficulties in the treatment of cancer such as drug resistance, toxicity, and low specificity. Most of the cancer treatments rely heavily on chemotherapy, however chemotherapy has limitations. Chemotherapeutic drugs lack of selectivity and they can also kill normal cells. Hence, there is an urgent need for new drugs that are highly effective and possess low toxicity. In this regard, several studies have reported the cytotoxic activity of organic extracts of spores, vegetative bodies² and fruiting bodies³ from several species of macro fungi against cancer cells. Lung cancer, also known as carcinoma of the lung or pulmonary carcinoma, is a malignant lung tumor characterized by uncontrolled cell growth in tissues of the lung. If left untreated, this growth can spread beyond the lung by process of metastasis into nearby tissue or other parts of the body. Lung cancer accounts for about 27% of all cancer deaths and is by far the leading cause of cancer death among both men and women. New concepts have appeared in this trend, such as nutraceuticals nutritional therapy, phytonutrients and phytotherapy. Functional foods and phytomedicines play positive role in maintaining well-being, enhancing health and modulating immune function to prevent specific disease. Macro fungi have long been used as valuable food source and as traditional medicines around the world, especially in the Orient⁴. Mushrooms are highly nutritive, low calorie food with good quality protein, vitamin, and minerals⁵. Macro fungi are known to produce large and diverse variety of secondary metabolites⁶. Several pharmacologically active compounds have been identified and isolated from Basidiomycota with a wide spectrum of biological activities and health promoting properties including immuno modulatory, antioxidant, antiviral, cholesterol lowering, anticancer and anti-inflammation activities⁷. *Calocybe indica*, commonly known as summer white mushroom or milky mushroom is one of the most economically important edible mushrooms that grow predominantly in hot humid climates in Tamilnadu, India. It is considered as valuable health food with high content of polysaccharides, vitamin B-complex, minerals and amino acids. Moreover, this mushroom has been demonstrated to possess various valuable biological properties including antimicrobial, anti-inflammatory, anti-diabetic, anti-tumor as well as antioxidant activities⁸. The present study has been aimed to determine *in vitro* anticancer activity of *Calocybe indica* Var APK2 against A549 lung cancer cell line.

MATERIALS AND METHODS

(i) Collection of Macro fungus

Cultivated fruiting bodies of *Calocybe indica* var. APK2 were obtained from the RPJ Mushroom farm, Usilanakottai, Ramanathapuram, Tamilnadu, India. The

samples were authenticated by Dr.A.S.Krishnamoorthy, Professor, Department of Plant Pathology, Tamilnadu Agricultural University, Coimbatore. All the samples were lyophilized, reduced to a fine dried powder (20 mesh) and mixed to obtain homogenous samples and stored in a desiccator protected from light, until further analysis.

(ii) Preparation of Methanolic extract of *C.indica*

Powdered fruiting body (100g) of *C. indica* was extracted with 500 ml of methanol by using soxhlet apparatus for 6h at 50-60°C and the extract was concentrated under vacuum by using rotary evaporator and stored under 4°C⁹. The crude compound was dissolved in 4% of DMSO (Dimethyl sulfoxide) and stored at -20° C for further studies.

(iii) Preliminary phytochemical screening

The powdered fruiting bodies were mixed with potassium bromide (pellets) and subjected at spectra frequency region from 4000-400cm⁻¹ through FT-IR studies¹⁰.

(iv) Evaluation of antioxidant activity

Antioxidants, namely the free radical scavengers are highly reactive chemicals that have the potential to harm cells. Free radicals can be hazardous to the body and damage all major components of cells, including DNA, proteins and cell membranes¹¹. The Antioxidant potential of *C.indica* was evaluated by DPPH assay and hydroxyl radical scavenging assay.

(v) DPPH radical scavenging assay

Free radical scavenging activity of methanolic extracted *C. indica* was performed using DPPH assay. 0.1mM solution of DPPH was prepared in 100% methanol, and 1 ml of this solution was added to 4 ml of sample in 40% methanol at various concentrations (10-160 µg/ml). This mixture was shaken vigorously and incubated for 30 min at 37°C in the dark. The reduction of the DPPH radical was measured by continuous monitoring of the decrease of absorption at 517 nm using ELISA reader. The L-ascorbic acid was used as standards¹².

DPPH Scavenging effect (%) = [(A0-A1)/A0] x 100
where A0 was the absorbance of the control reaction and A1 was the absorbance in the presence of the standard sample or extract.

(vi) Hydroxyl radical scavenging assay

This assay was carried out according to the method Halliwell¹³. To the reaction mixture containing deoxyribose (3 mM, 0.2 ml), ferric chloride (0.1 mM, 0.2 ml), EDTA (0.1 mM, 0.2 ml), ascorbic acid (0.1 mM, 0.2 ml) and hydrogen peroxide (2 mM, 0.2 ml) in phosphate buffer (pH, 7.4, 20 mM), 0.2 ml of various concentrations of extracts or standards in DMSO were added to give a total volume of 1.2 ml. The solutions were then incubated for 30 min at 37°C. After incubation, ice-cold tri-chloroacetic acid (0.2 ml, 15 % w/v) and thiobarbituric acid (0.2 ml, 1 % w/v), in 0.25 N HCl were added. The reaction mixture was kept in a boiling water bath for 30 min, cooled and the absorbance was measured at 532 nm.

(vii) Cell culture

Cytotoxic activity of *C.indica* on lung cancer cell line

The A549 human lung adenocarcinoma epithelial cell lines were purchased from National Centre for Cell Science, Pune, India. These A549 cells were maintained in DMEM (Dulbecco's Modified Eagle Medium) supplemented with 10% heat-inactivated Fetal Bovine Serum 100 IU/ml penicillin and 100 mg/ml streptomycin at 37 °C in a humidified atmosphere of 95% air and 5% CO₂

(viii) Morphological observation

Morphological changes of cells in both of the treated group and control group were analyzed at 8 h; 24 h under the OLYMPUS 1×71 Inverted Fluorescence microscope with 10× magnification¹⁴.

(ix) Cytotoxic activity of *C. indica* on lung cancer cell line**MTT Assay**

The cytotoxic effect of methanolic extraction of *C. indica* against A549 lung cancer cell line was assayed by MTT assay. A549 cells were grown in 96 well micro titer plate (5×10³ cells well) for 24h after seeding. The plates were incubated with methanolic extract with different concentration for 24h and 48 h (check to put h) respectively. The medium was refreshed and 20µl of MTT (5mg/ml) was added. The plates were incubated for three h in dark. The formazan crystals developed were solubilized with 100µl of DMSO and the plates were kept in dark for another 5-10 min. The colour developed was measured in an ELISA reader at 570 nm. The IC₅₀ concentration was determined as the drug concentration that is required to reduce the absorbance to half that of the control¹⁵.

(x) Cell cycle analysis

A549 lung cancer cells were seeded in 6 or 12 well culture plates. After cell adherence, cells were treated with mushroom extract (500µg/ml) for 24 h and, the cells were harvested using trypsin (0.05%)-EDTA (0.54 mM) solution. Later, the cells were washed using media thoroughly and centrifuged at 1200 rpm for 5 min, the pellets were collected and 300 µl of PBS was added. These cells were added with 700 µl ice cold ethanol drop by drop and the cells were fixed with ethanol and stored overnight at 4°C. For flow cytometry analysis, the cells were washed with PBS. Triton X 556 µl containing RNase 20 µl were added with the cells and

incubated for an hour. After incubation, propidium iodide 24 µl was added and incubated in dark for 45 min and the sample was analyzed using flow cytometer¹⁶.

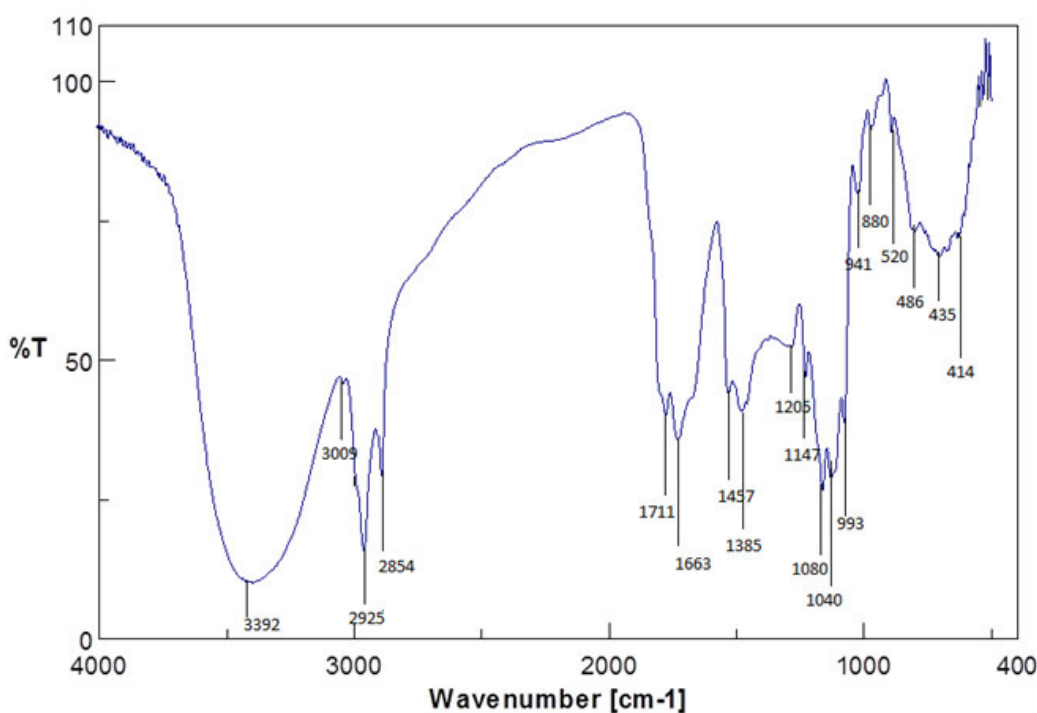
RESULT AND DISCUSSION

Mushrooms are highly nutritive, low calorie food with good quality of protein and vitamins. Macro fungi are known to produce a large and diverse variety of secondary metabolites¹⁷. These secondary metabolites have health promoting properties such as antioxidant, antimicrobial, anticancer, cholesterol lowering and immunostimulatory effects¹⁸. Radiation therapy is commonly used in the treatment for cancer, in order to damage the DNA of the tumour cell to inhibit proliferation and induced apoptosis. The main side effect of using radiation therapy is DNA damage to the surrounding healthy tissues. Hence protection of non – cancerous tissue against radiation is vital for reducing the side effect. The present study was undertaken to elucidate the antioxidant effect and *in vitro* cytotoxic potential of edible mushroom *Calocybe indica* varAPK2. Therefore organic extractions, such as methanol, ethyl acetate, chloroform and aqueous extraction were taken. In FT-IR profile of all the solvent extracts, the methanolic extraction eluted the potential phytochemicals such as alcohols, phenols, alkanes, aromatics, unsaturated esters, carbonyl groups, esters, ethers, primary amines and alkyl halides (Table 1). The methanolic extraction of fruiting bodies of *Ganoderma lucidum* produced two fold increased cytotoxic activity in mouse melanoma cell line¹⁹. Similarly, phytochemical screening of methanolic extraction of the fruiting bodies of *Cantharellus cibarius* showed phenols, flavonoids, saponins and anthroquinones²⁰. The phenols and flavonoids have reported to provide the protection against oxidative stress induced diseases and responsible for the antioxidant activity of the mushroom²¹. Alkaloids are suggested to stimulate the anesthetic effect in ophthalmology²². Flavonoids and phenols have been reported to provide protection against oxidative stress induced diseases and majorly responsible for the antioxidant activity of mushrooms²³. The synergistic effect of the phytochemicals and nutraceuticals present in mushroom has been implicated their antimicrobial and antioxidant activity²⁴.

Table 1
FT-IR frequency and functional group of *Calocybe indica*

S. No	Peak Value	Functional Groups
1.	3500-3200	Alcohol and phenol
2.	3100-3000	Aromatics
3.	3000-2850	Alkanes
4.	3000-2850	Alkanes
5.	1730-1715	α and β unsaturated esters
6.	1760-1665	Carbonyl(general)
7.	1500-1400	Aromatics
8.	1400- 1300	Aromatics
9.	1320-1000	Alcohols, carboxylic acid esters and ethers
10.	1320-1000	Alcohols, carboxylic acid, esters and ethers
11.	1320-1000	Alcohols, carboxylic acid, esters and ethers
12.	1320-1000	Alcohols, carboxylic acid, esters and ethers
13.	950-910	Carboxylic acid
14.	900-665	Primary and Secondary amines
15.	690-515	Alkyl halides

Figure 1
FT-IR analysis of *Calocybe indica*



(1) Antioxidant Assays

Oxidative stress is an important parameter that increases cancer risk. Free radicals are known to induce oxidative damage in biomolecules and play an important role in aging, cardiovascular disease and unbalanced immune system²⁵. Antioxidant activity is manifested in wide variety of actions such as inhibition of oxidation enzymes, chelating of transition metals and enzyme detoxification of ROS²⁶. The scavenging activity of the methanolic extraction of *C.indica* was compared with L-Ascorbic acid as standard. The DPPH radical scavenging activity was increased at sample concentration 50 to 250 μ g/ml. The dose dependent of DPPH radical scavenging activity of methanolic extract of *C.indica* recorded the higher scavenging activity (Figure 2). The EC₅₀ value of methanolic extract of the

C.indica was expressed as 89 μ g/ml. The hydroxyl radical is one of the representative reactive oxygen species generated in the body by the common biological reaction of Iron (II) based Fenton reaction. The hydroxyl radicals are considered to be major harmful Reactive Oxygen Species (ROS) that may cause the oxidative damage of the biomolecules²⁷. The EC₅₀ value of *C.indica* was expressed as 68 μ g/ml (Figure 3).The radical scavenging capacity may be attributed to phenolic compounds in methanolic extract with the ability to accept electrons, which can combine with free radical competitively to decrease hydroxyl radical. Methanolic extraction of *Phillusrimosus* showed significant hydroxyl radical scavenging activity and the EC₅₀ value ranging from 25.3 μ g/ml²⁸.

Figure 2
DPPH scavenging activity

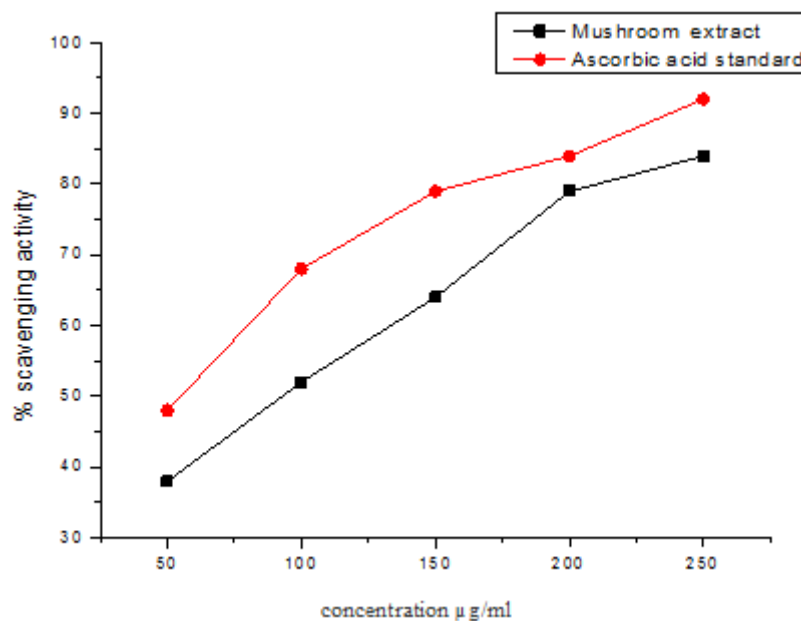
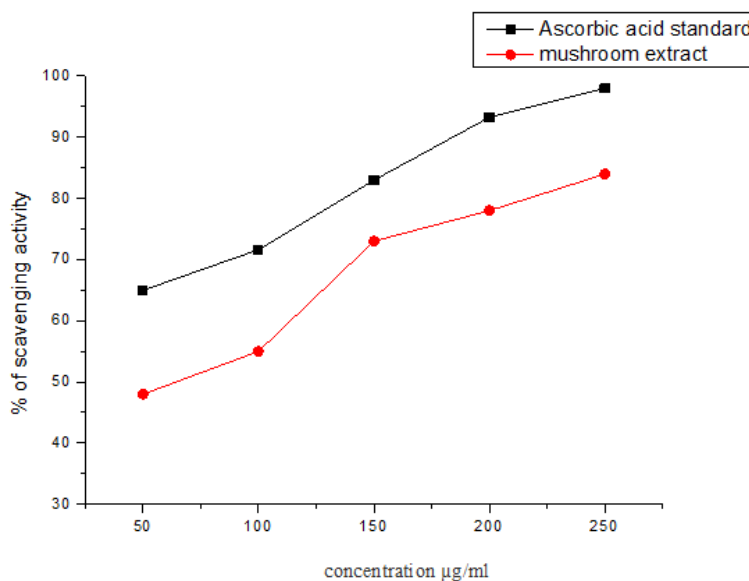


Table: 3
Hydroxyl radical scavenging assay

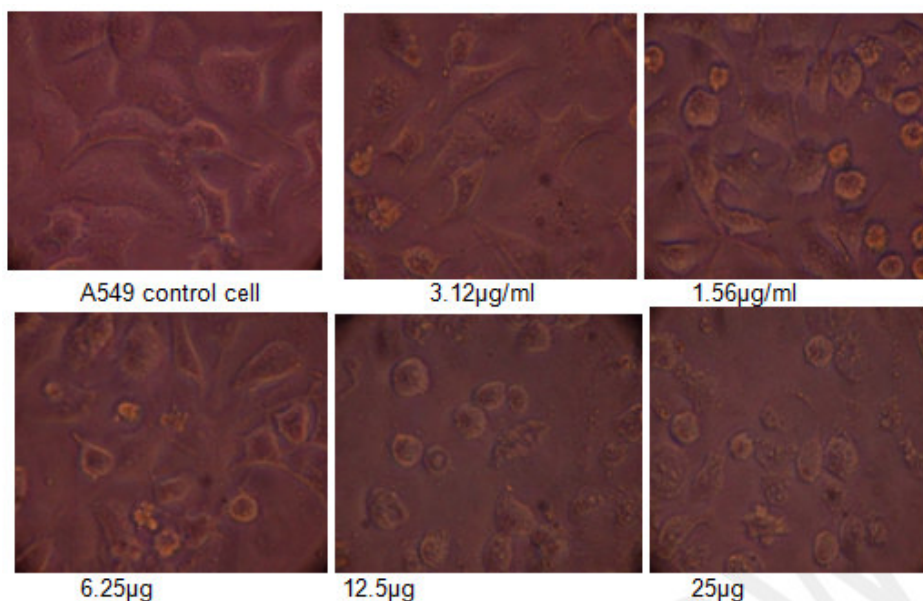


(2) Cytotoxic activity

Antiproliferative activity of A549 lung cancer cells against *Calocybe indica* was assessed using MTT assay as it depends mainly on the mitochondrion dehydrogenase which is the important parameter to assess the viability of the cells²⁹. The mushroom extract expressed the potent cytotoxic activity against human

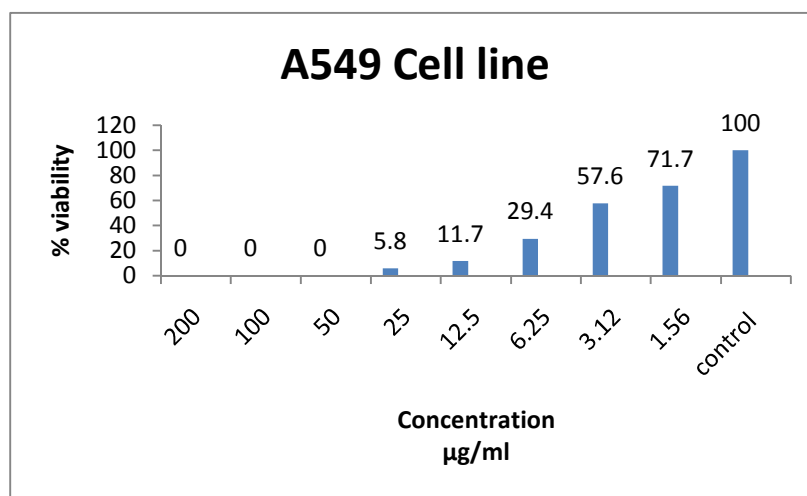
lung cancer cell line (Table 2). Morphological observation of A549 cell was determined by the changes induced by the *C.indica*. Changes such as shrinking of the cells, membrane, ballooning, were observed in predicting the apoptotic mechanism for cell death (Figure 4).

Figure 4
Morphological changes of A549 control and *C.indica* treated cells



The fruiting bodies of *C. indica* expressed maximum cytotoxic activity of 71.7% at 1.56µg/ml concentration and IC50 value of 4.9µg/ml. The methanolic extraction of fruiting bodies of *Pleurotus ostratus* expressed maximum cytotoxic activity of 87% and the IC50 value was 39µg/ml³⁰. Moreover the cytotoxic activity of *Cordyceps taii* (Caterpillar mushroom) against A549 lung and human gastric cancer cells in dose dependent manner from 1.9 to 250µg/ml³¹.

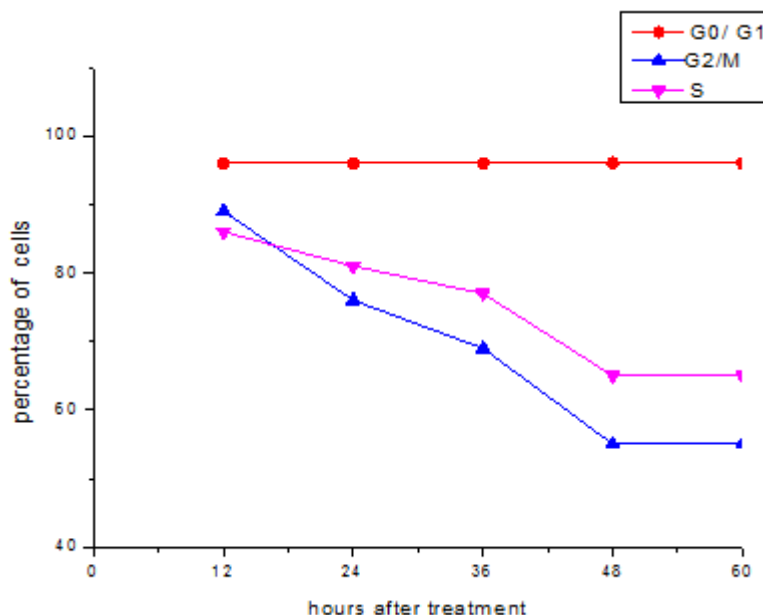
Table 2
Cytotoxic activity of *C.indica* (MTT assay)



Cancer cells have a deregulated cell cycle leading to uncontrolled cell proliferation. In mammalian cells, the cell cycle generally occur in regular fashion from G1 to S and M phase in the presence of Cyclin dependent protein kinase (Cdks) and accelerated by the dependent kinase inhibitors P16, P21 and P27, retinoblastoma, tumor suppressor protein, D type cyclins such as D1, D2 and D3 are involved in regulation of transition from suppressor protein. Anticancer agents can arrest the cell cycle at G0/G1, S or G2/M phase reduce the rate of cell proliferation³². To investigate the

basis antiproliferative properties of methanol extract of *Calocybe indica*, recorded accumulation of cells in G0-G1 phase. In G2/M phase (56%), the number of cells decreased gradually from 12-48h. Similar trend has been noticed in S phase also (68%) (Figure 5). Antiproliferative properties have been reported in alcoholic extract of *Ganoderma lucidum* which induces cell cycle arrest at G1 phase in MCF7 cells³³.

Figure 5
Cell cycle arrest (Flow cytometer assay of *C.indica*)



CONCLUSION

The milky mushroom *Calocybe indica* could be used as the promising functional food and phyto-medicine as it recorded to have the cytotoxic effect against A549 lung cancer cell line through *invitro* studies. However further studies should be extended to identify the molecular targets, optimum dosage, efficacy, safety etc for the invention of pharmaceutical drugs.

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REFERENCES

- Jia-Chuan Lei, Cheng-Xiong Yang, Yuan Yang, Wu Zhang, Jian-Qing Yu. Antioxidant and antitumour activities of extracts from *Patrinia villosa* and its active constituents. *Pharmaceutical Biol.* 2009; 47(5): 384–389.
- Choi YS, Kim HK, Lee BJ, Kim YG. Characteristics and breeding of a new variety *Cordyceps militaris*. *J. Mushroom Science and Prod.* 2009; 7: 182–186.
- Takaku T, Kimura Y, Okuda H. Isolation of an antitumor compound from *Agaricus blazei* and its mechanism of action. *J Nutrition.* 2001 ;131: 1409–1413.
- Wasser SP. Medicinal mushrooms as a source of anti-tumour and immunostimulating polysaccharides. *Appl. Microbiol. Biotech.* 2002;60: 258 - 274.
- Khatun, S, Islam, A, Cakilcioglu U, Chatterjee, N. C. Research on mushroom as a potential source of Nutraceuticals A review on Indian perspective. *American Journal of Experimental Agriculture.* 2012; 2 (1): 47-73.
- Liu J K. Secondary metabolites from higher fungi in China and their biological activity. *Drug Discov. Ther.* 2007; 1(2): 94 – 103.
- Fabricao S. Silva, Matheus S. de Sá1, José Fernando O, Costa, Fernanda P. Pinto, *In vitro* pharmacological screening of macro fungi extracts from the Brazilian northeastern region. *Pharmaceutical Biol.* 2009; 47(5): 384–389
- Selvi S, Umadevi P, Suja S, Sridhar K, Chinnaswamy P. Inhibition of *invitro* lipid peroxidation (LPO) evoked by *Calocybe indica* (milky mushroom). *Ancient Science of Life.* 2006; Vol : XXVI : 41-45.
- Feyzaoke and Belma Aslima B. Protective effect of two edible mushrooms against oxidative cell damage and their phenolic composition. *Food chem.* 2011; (128): 613-616.

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CONFLICTS OF INTEREST

Disclosure the authors report no conflicts of interest in this work.

10. JagMohan. Organic spectroscopy principles and applications. Narosa publishing House, Daryagani Delhi.2005.
11. Valko M, Leibfritz D ,Monocol J, Cronin M ,Mazur M ,Tulser J. Free radical and antioxidant in normal physiological function and human diseases. I J.Biochem Cell biol. 2007; (39):44-84.
12. Brand-Williams W,Cuvelier M E, Berset C. Use of free radical method to evaluate antioxidant activity. LebensmWiss Technology. 1995; 28:25-30.
13. HalliwellB.Antioxidant in human health and Disease. Annual review of nutrition. 1996;16: 33-50.
14. Ramesh, Santosh K, Pavunraj M, KarunakaranC ,Rajendran A .*In-vitro* antifungal and anticancer potential of *Xylariacurta* fruiting body fractions against human fungal pathogen and cancer cell lines. Current Research in Environmental & Applied Mycology.2015; 5 (1): 20–26.
15. Mosmann,T, Rapid colorimetric assay for cellular growth ,survival application to proliferation and cytotoxicity assays. J. Immunol. Methods.1983; (65): 55.
16. Hongbo H U, Nam-Shik A H, Xinlinyang,Yong-Soon lee, Kyung-Sun.*Ganoderma lucidum* extract induces cell cycle arrest and apoptosis in MCF7 human breast cancer cell. Int.J.Cancer. 2002; 102:250–253.
17. Liu J-K. Secondary metabolites from higher fungi in China and their biological activity. Drug Discov Ther.2007; 1(2): 94 – 103.
18. Anderson J B ,Stasovski E. Molecular phylogeny of Northern Hemisphere species of *Armillaria*.Mycologia. 1992; 84: 505- 516.
19. Tong CC, Choong YK, Umar NB, Noordin MM , Mohamed S. Cytotoxic activity induced by crude extracts of *Ganoderma lucidum* (W. Curt.: Fr.) P. Karst. on mouse myeloma cancer cell-line. World Journal of Microbiology & Biotechnology. 2009; 25(4):687-695.
20. Aina D A, Jonathan S G, Olawuyi O J, Ojelabi D O ,Durowoju. Antioxidant, antimicrobial and phytochemical properties of alcoholic extracts of *Cantharelluscibarius* – A Nigerian mushroom. Reactive Oxygen Species. *In The handbook of oxidative metabolism. Massachusetts.* 2012; (10): 1-4.
21. Barros L, Calhelha, R C, Vaz J A, Ferreira I, Baptista P and Estevinho L M Antimicrobial activity and bioactive compounds of Portuguese wild edible mushrooms. Eurp Food Res Technol .2007; 225:151-6.
22. Egwin, E C, Elem R C,Egwuche, R U.Proximate composition, phytochemical screening and antioxidant activity of ten selected wild edible Nigerian mushroom. American Journal of Food and Nutrition.2011; 1 (2): 89-94.
23. Egwin, E C, Elem R C ,Egwuche, R U. 2011. Proximate composition, phytochemical screening and antioxidant activity of ten selected wild edible Nigerian mushroom. American Journal of Food and Nutrition.2011; 1 (2): 89-94.
24. Barros L, Venturini B, BaptistaP,Freirec, Vilas and Ferreira. Antioxidant activity of *Agaricus*Sp Mushrooms by chemical,biochemical and electro chemical assays. Food chemistry. 2008; 111:61-6
25. Wang X and Quinn P. Vitamine E and its function in membrane. Prog Lipid Research. 1999;(38):306.
26. Halliwell B. Antioxidants in human health and disease. Annu. Rev. Nutr.1996;16: 33-50.
27. Pallavi Sharma, Ambuj Bhushan Jha, Rama Shanker Dubey, Mohammad Pessaraki.. Reactive Oxygen Species, Oxidative Damage, and Antioxidative Defense Mechanism in Plants under Stressful Conditions Journal of Botany. 2012; 26.
28. Ajith T A,Janarthan K K. Antioxidant and anti-inflammatory activity of methanol extract of *Phellinus rimosus*(Berk)Pilat. Indian journal of experimental biology. 2007;(39):1166-1169.
29. Aleksandra R. Novaković¹, Maja A,Karaman, Ivan L. Milovanović, Miona M.. Edible mycorrhizal species *Lactarius controversus* Pers. 1800 as a source of antioxidant and cytotoxic agents. Hemijskaindustrija Publication on Research gate .2015.
30. Rahman M, Faridur, Karim M, Rezaul, Islam.M, Farhadul,etal. Phytochemical and cytotoxic investigation on Oyster mushroom (*Pleurotus ostreatus*).International research journal of pharmacy 2010;1(1):342-345.
31. Ru-Ming Liu, Xiao-Jie Zhang, Gui-You Liang, Yong-Fu Yang, Jian-Jiang ZhongJian-Hui Xiao.Antitumor and antimetastatic activities of chloroform extract of medicinal mushroom *Cordyceps taii* in mouse models. BMC Complementary and Alternative Medicine.2015; 15:216.
32. Mei-Yin Chang, Den-En Shieh, Chung-Chi Chen, Ching-Sheng Yeh, Huei-Ping DongGuido ,Haenen R. Linalool Induces Cell Cycle Arrest and Apoptosis in Leukemia Cells and Cervical Cancer Cells through CDKIs.Int J Mol Sci. 2015; 16(12): 28169–28179.
33. Hongbo HU,Nam-Shik AHN,Xinlin YANG, Yong-Soon LEE, Kyung-Sun KANG. *Ganoderma lucidum*extractinducescellcyclearrest and apoptosis in MCF-7humanbreastcancer cell Int.J.Cancer.2002;102 :250–253.