



## MANAGEMENT OF GENETIC ACTIVITY THROUGH HOMOEOPATHY

**DR. H. U. GANGAR**

Ex-Head, Engineering & Workshop,  
Central Institute for Research on Cotton Technology  
(Indian Council of Agricultural Research), Mumbai, India

*Corresponding author*      hugangar@hotmail.com

### ABSTRACT

Preliminary study on effects of homoeopathic drugs on plants was undertaken at Central Institute for Research on Cotton Technology, Mumbai. It was found that homoeopathic drugs influence genetic activity in the agricultural plant bodies in a big way. Since, there is neither brain nor any central nervous system in plants; known principles/theories of homeopathy cannot be applied here. Under this background, research study on the current subject is undertaken. It is found that medicated water (carrying homeopathic drug) contains intermolecular electrical charges. Present paper explains how these electrical charges perform the act of transferring the energy, (received from original drug), into the concerned gene within the DNA of that cell to trigger the respective gene into the act of activation, acceleration, retardation or turning off its relevant activities. This phenomenon is explained with the help of experiments on cotton, wheat, gram, mungbean etc.

### KEYWORDS

Homoeopathy, Resonance, Potency, Electromagnetic waves, DNA, Gene

### INTRODUCTORY THEORY

#### *Resonance phenomenon*

Each object, which also includes an electron, a molecule as well as a gene, has its own natural frequency of vibrations/oscillations called resonant frequency<sup>1</sup>. When bondage of an object with its surrounding is diluted and it is repeatedly pushed, it absorbs energy from each successive push and starts oscillating at its resonant frequency (Figure 1).

Resonance has many physical applications. Children use it to increase the size of the movement on a swing, by giving a push at the same point during each swing. Concept of resonance is used in musical instruments also. In case of stringed instruments, cross-sectional diameter and length of string decide the resonant frequency of vibrations which in-turn decides the pitch of musical sound. Amplitude of vibrations decides the volume of sound.

Homoeopathic drugs are prepared by successively diluting and simultaneously, vigorously shaking the drug in the medium of rectified spirit. When molecules of the drug are subjected to successive dilutions and vigorous shaking, the bonding between molecules themselves and also within a molecule and its

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electrons weaken. On account of loose bonding and vigorous shaking, molecules and also electrons within them start oscillating at their own individual resonant frequencies.

### Forced resonance of a swing

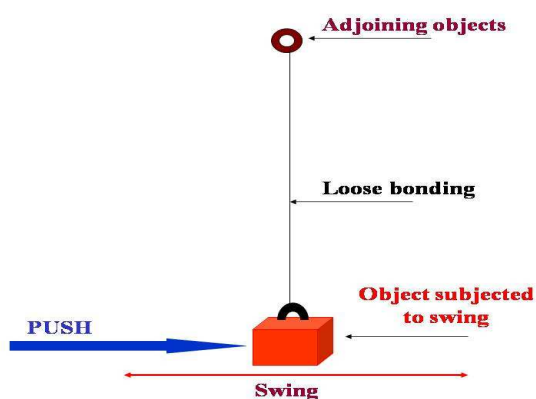


Figure 1

*With each push, size of swing increases*

### Modulation

However, the resonant frequency of a molecule is in the range of few MHz (million oscillations per second) while that of an electron is in the range of few GHz (billion oscillations per second)<sup>2</sup>. This makes oscillations of electrons to act as carrier on which oscillations of molecules get superimposed (modulation) (Figure 2).

### Modulations of oscillations of electron within a molecule

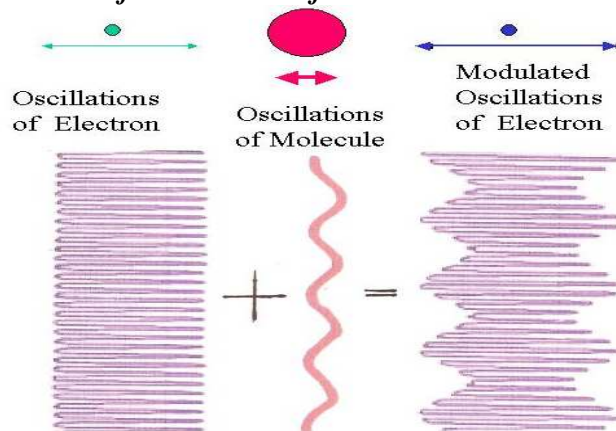


Figure 2

*Oscillations of electron get modulated by oscillations of molecule of a drug*

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### *Transfer of energy*

Weakening of bonds between outermost electrons and nucleus of molecule occurs during further dilutions and shaking. Beyond 12<sup>th</sup> potency, molecules of original drug disappear. However, electrons continue oscillating and carry with them oscillations of molecules of the original drug. More and more energy is pumped into oscillations of electrons during further dilutions and shaking and this energy remains stored in them (Figure 3)

### *Absorption and transfer of successive higher energy*

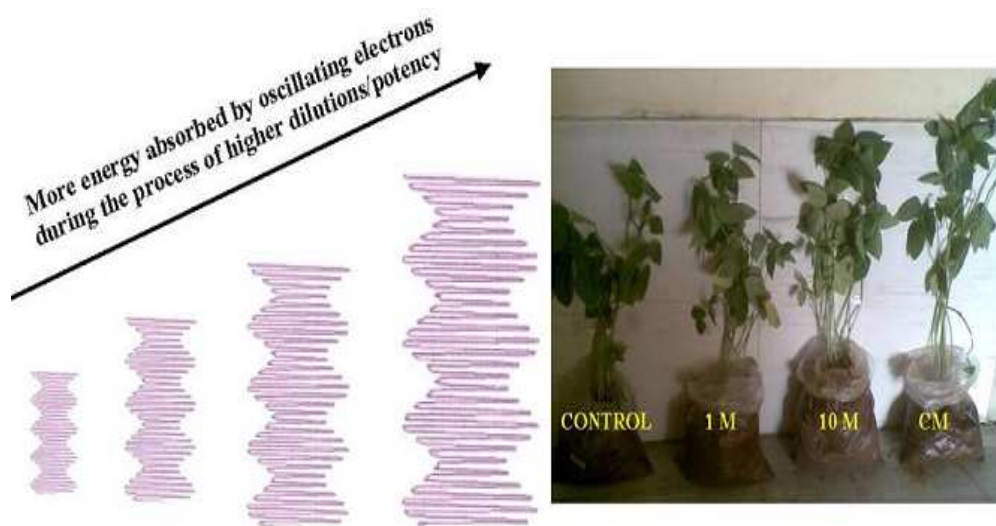
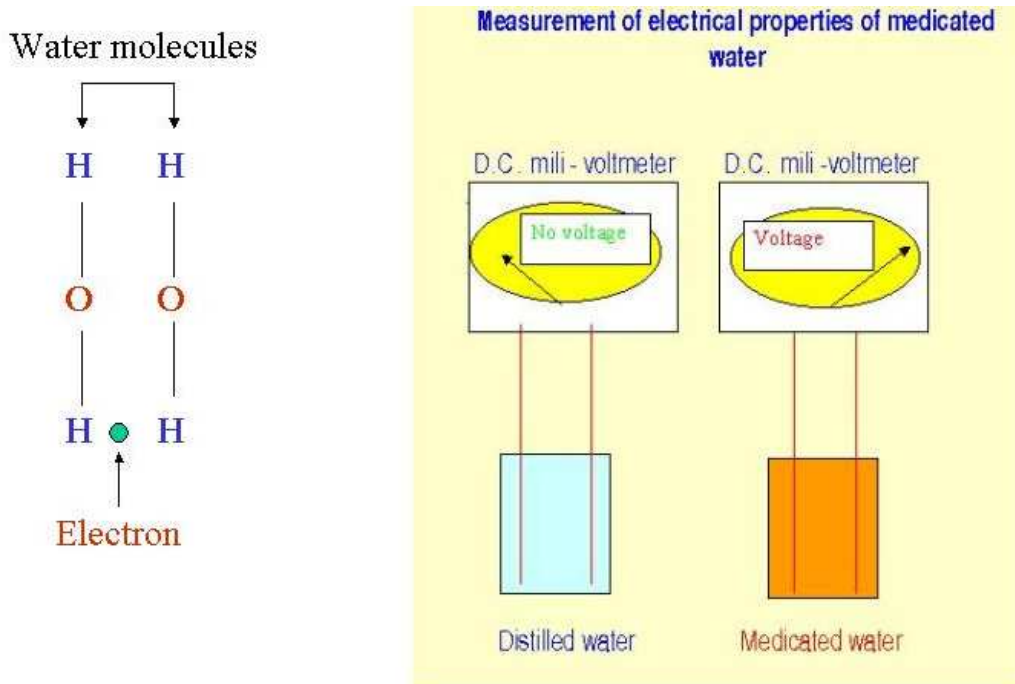


Figure 3 Oscillations of electrons absorb additional energy during the process of higher dilutions. Transfer of this energy results in enhanced genetic activity as seen in successive higher growth of medicated chauli plants with corresponding increase in dilutions of drug.

When few drops of drug are added into electrically neutral water, it develops internal electrical charges<sup>3</sup> as indicated by presence of D.C. voltage which does not get discharged on grounding the electrodes (Figure-4). It indicates that oscillating electrons have got attached to water molecules. Water molecules act as conductor for oscillating electrons when they penetrate with them into plant/animal cell by “Osmosis”.

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*Medicated distilled water develops D. C. Voltage*



**Figure 4**

*When few drops of homoeopathic drug are added in distilled water, it develops inter-molecular electrical charge*

Oscillating electrons generate oscillating electromagnetic waves<sup>4</sup> inside the cell. These waves carry with them, the resonance frequency (in the range of MHz) of original drug. Each gene present in DNA within the cell also has its own natural frequency of resonance in the range of MHz.

When resonance frequency of electromagnetic wave is nearly same as that of any gene in the cell, it triggers the relevant gene into oscillations. Activated gene emits electromagnetic waves<sup>5</sup>. These waves trigger corresponding genetic process.

If these waves transmitted by electrons are in same direction/phase as those of corresponding gene, it results in activation/acceleration of process. Similarly, if waves are in opposite direction/phase it results in retardation/deactivation of process.

This way energy from oscillating electrons is transferred to relevant gene to trigger it to activate/accelerate/retard/deactivate the corresponding process. Experiments conducted under the present study illustrate this ability of homoeopathic drugs to trigger the genes into the act for activation, acceleration, retardation and deactivation of corresponding processes in the body.



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### MATERIAL, METHODS AND RESULTS

Ten samples under control conditions and ten samples under medication were used for the present study in each experiment (on plant body). Pots and plastic bags were used for sowing seeds. Same variety of seeds was used for experiments under control and medication conditions.

In order to decide optimum dose, samples of medicated water containing medicine (Abrotanum CM+Barayta Carb CM) in different concentration (1%, 2%, 3% and 4%) were prepared. 0.1 ml, 0.2 ml, 0.3 ml and 0.4 ml of a drug was added in 10 ml of pure water to prepare samples of 1%, 2%, 3% and 4% concentration respectively. Rate of germination of cottonseeds submerged (for 4 days) in distilled water and that in medicated water was evaluated. It is seen from Table 1 that medicated water with 3% concentration produced maximum rate of germination. Hence for all the experiments medicated water containing 3% drug has been used.

**Table 1**  
*Effect of drug concentration on rate of germination*

Sr. No.	Average lengths of the sprouts in cm.				
	Control (water without drug)	Medicated water containing 1% drug	Medicated water containing 2% drug	Medicated water containing 3% drug	Medicated water containing 4% drug
1	6.4	14.0	14.8	17.5	12.4

Table 1 Medicated water containing 3% drug (Abrotanum CM+Barayta Carb CM) produced maximum effect on rate of germination

For evaluating effect of varying the potency on rate of genetic processes, one set of seeds of chauli were soaked for 4 hours in distilled water (for control condition) and another set of seeds were soaked for 4 hours in medicated water containing 1M, 10M and CM potency of drug (Natum Mur+Acid Picric) before sowing. It was observed that rate of germination and growth was highest in case of plants medicated with CM potency (Figure 3). Hence for all the experiments on plant bodies, drugs of CM potency have been used.

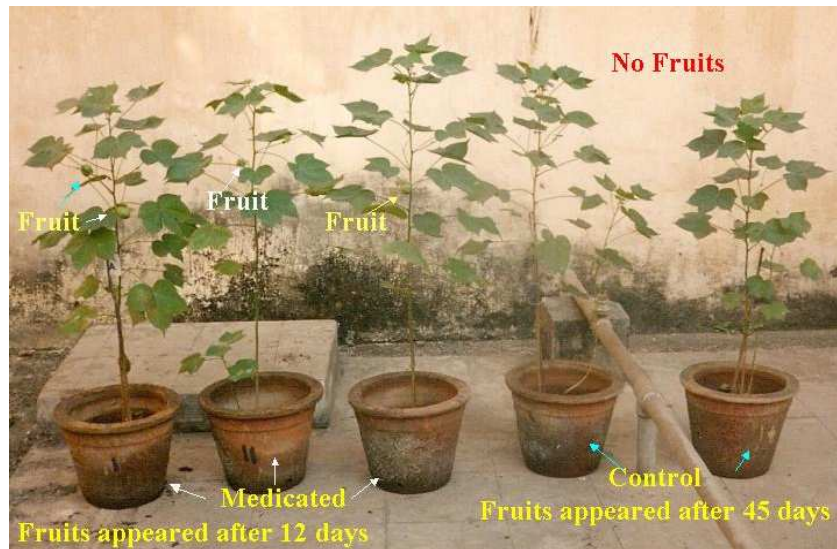
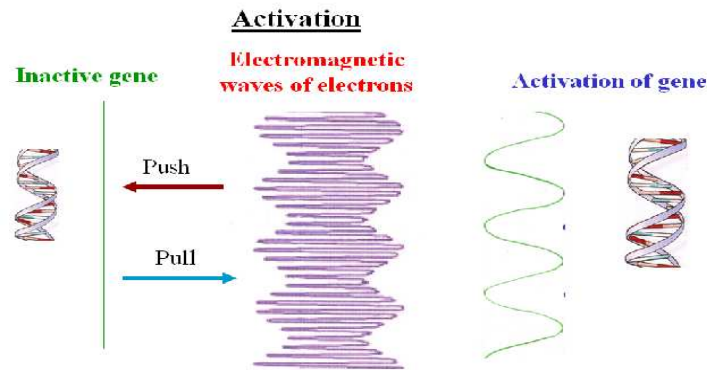
#### **Activation**

For conducting this test, untreated medium staple length hybrid cotton seeds were sown in pots. It was observed that after 110 days all the plants were fully grown, but flowering process was not started on any of the plants<sup>6</sup>. These plants were divided into two sets. Plants from first group were not given any

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treatment. Plants from second set were given a dose of Phosphorus CM by injecting 0.05 ml of medicated water into its stem. Within one week flowers appeared on treated plants. In the second week, fruits were seen on these plants (Figure-5). In case of untreated (control) plants, flowers appeared 45 days later. After 60 days from administration of first dose, second dose of same drug was given to plants under medication. This time also the reproduction process was repeated, making it possible to get second crop. Thus the relevant drug has triggered the gene to activate the reproduction process much earlier than its scheduled time. This also resulted in increase of yield and improvement in quality of cotton<sup>3</sup>.

*Triggering the gene into activation*



**Figure 5**

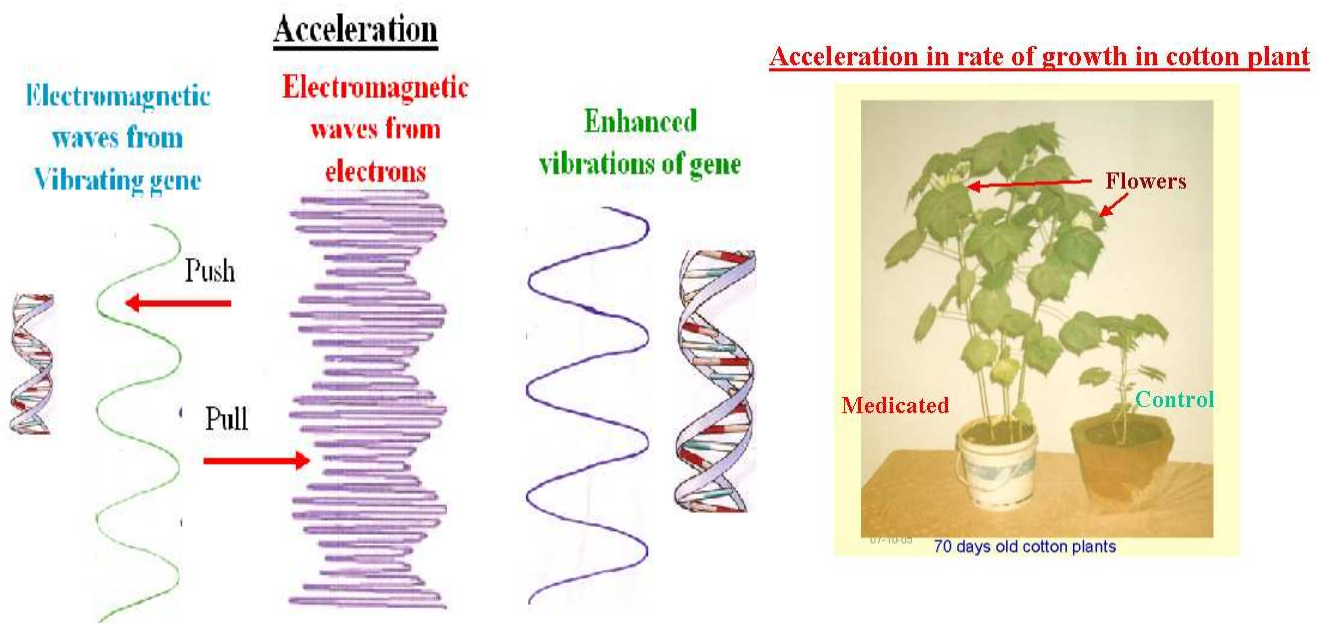
*Reproduction process in cotton plants started much earlier by activation of relevant gene by homoeopathic drug*



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### ***Acceleration***

For accelerating rate of germination of cottonseed<sup>3</sup> medicated water containing Abrotanum CM+Barayta Carb CM was used. Similarly for germination of wheat, water containing Platina CM+Abrotanum CM was used. Germination tests on cottonseed was carried out by submerging seeds in 5 ml of distilled water and medicated water for control condition and that under medication respectively. For evaluating acceleration in germination under soil, cotton seeds and wheat seeds were soaked momentarily in distilled water and medicated water for control and medication test respectively before sowing. For accelerating rate of growth of cotton plant, Iodum CM was used to medicate water. Distilled water was used to water cotton plant under control condition. Same quantity of medicated water at same interval of time was used to water plants under medication at the same time. In this case, of experiment on growth of cotton plant, the entire life cycle was reduced to 100 days in place of 200 days<sup>6</sup> for control plants. The acceleration in rate of genetic processes is clearly seen in Figure-6 (a), (b) and (c).



**Figure 6**

*(a) The higher rate of genetic activity is clearly seen in increased growth of cotton plant under medication*

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**Acceleration in rate of germination in wheat**

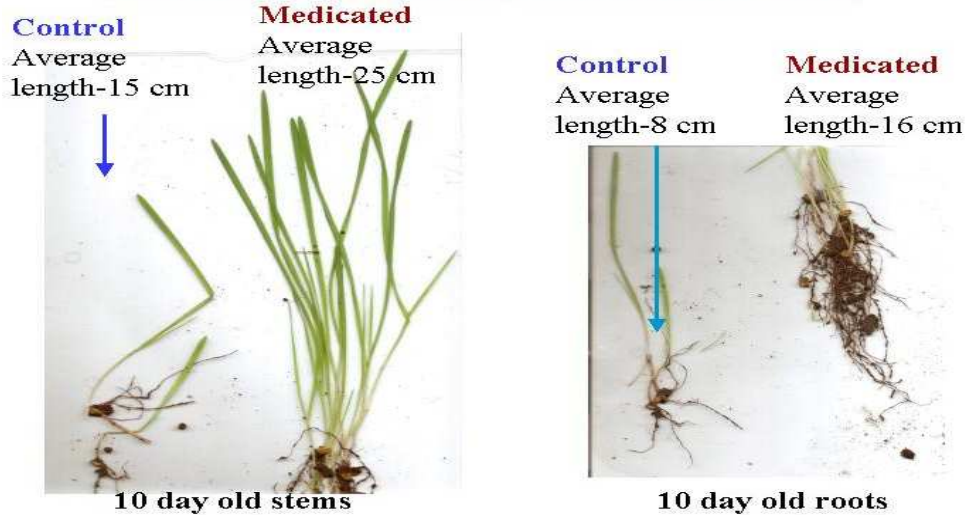


Figure 6

(b) Higher rate of germination, in wheat, is clearly seen in increase in length of stem and roots due to medication

**Acceleration in rate of germination in cotton**

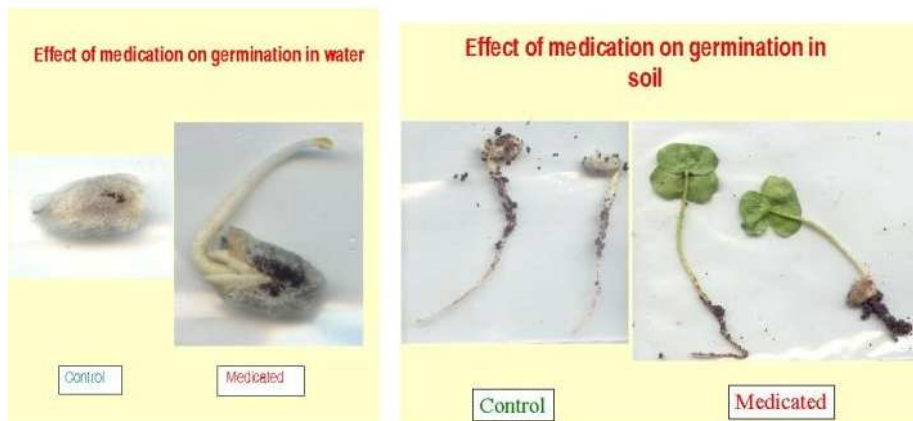


Figure 6

(c) (Left) Cotton seeds submerged under medicated water for 24 hours germinated faster (Right) Seedlings of cotton seed sown in soil for 3 days are displayed. Sprouts under control conditions did not appear on surface of soil. Medicated sprouts appeared on surface of soil within two days. Roots of medicated seeds were well spread in the soil and could not be taken out.

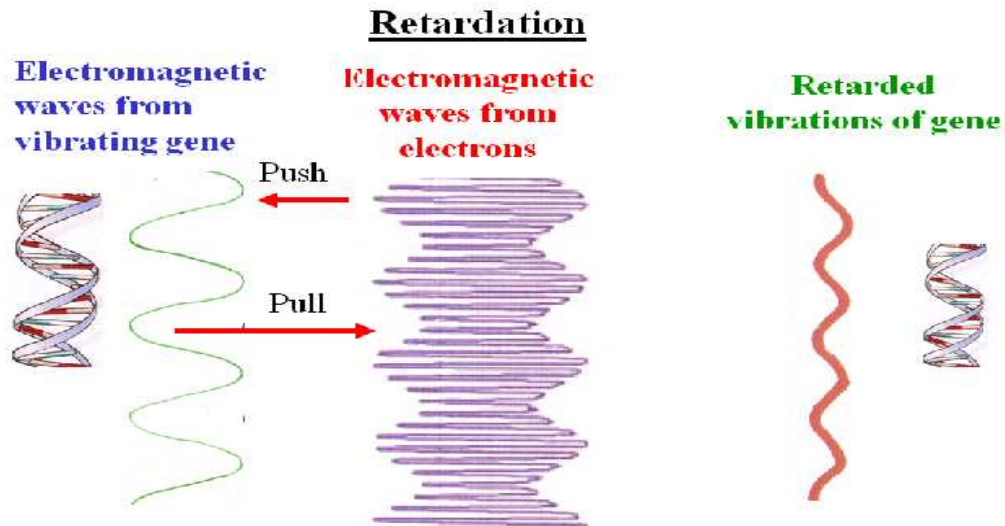
**Retardation**

For retarding the rate of germination in mung Acid Picric CM, and for that in gram Abrotanum CM+Acid Picric CM were used to medicate the water. Seeds of both these species were soaked

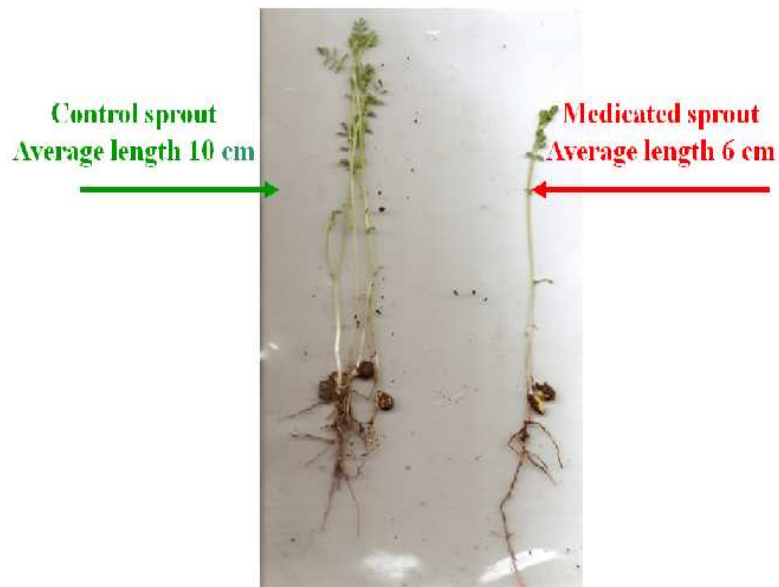


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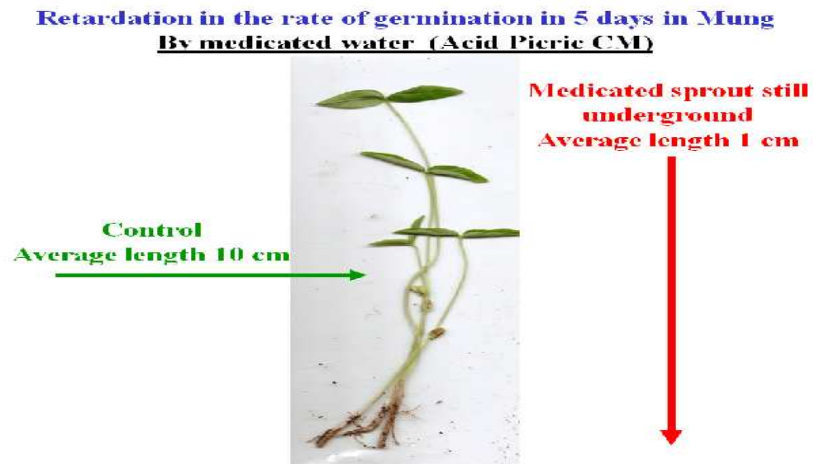
momentarily in distilled water and medicated water respectively before sowing. Retardation in genetic process is clearly visible in Figure-7.



**Retardation in the rate of germination in 5 days in gram  
By medicated water (Acid Picric CM+Abrotanum CM)**



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**Figure 7**

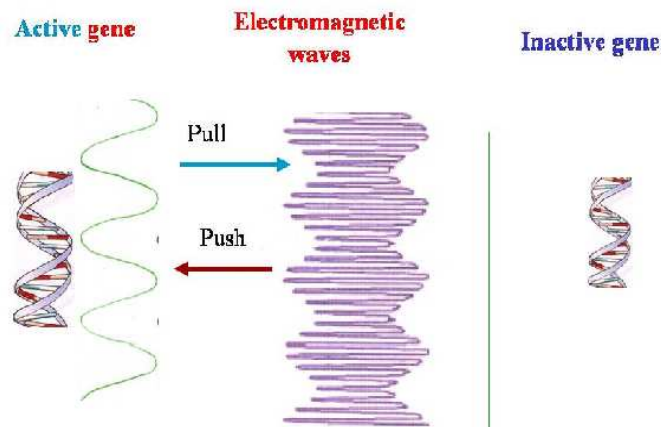
*Retardation in germination process due to medication is seen in case of gram and mungbeans. However, in case of mungbean rate of retardation is more.*

### **Deactivation**

For completely deactivating the germination of chauli, medicated water containing Natrum Mur CM+Acid Picric CM was used. Seeds of chauli were soaked momentarily in distilled water and medicated water before sowing for test under control and medication conditions respectively. There was absolutely no germination in respect of medicated seeds as seen in Figure-8.

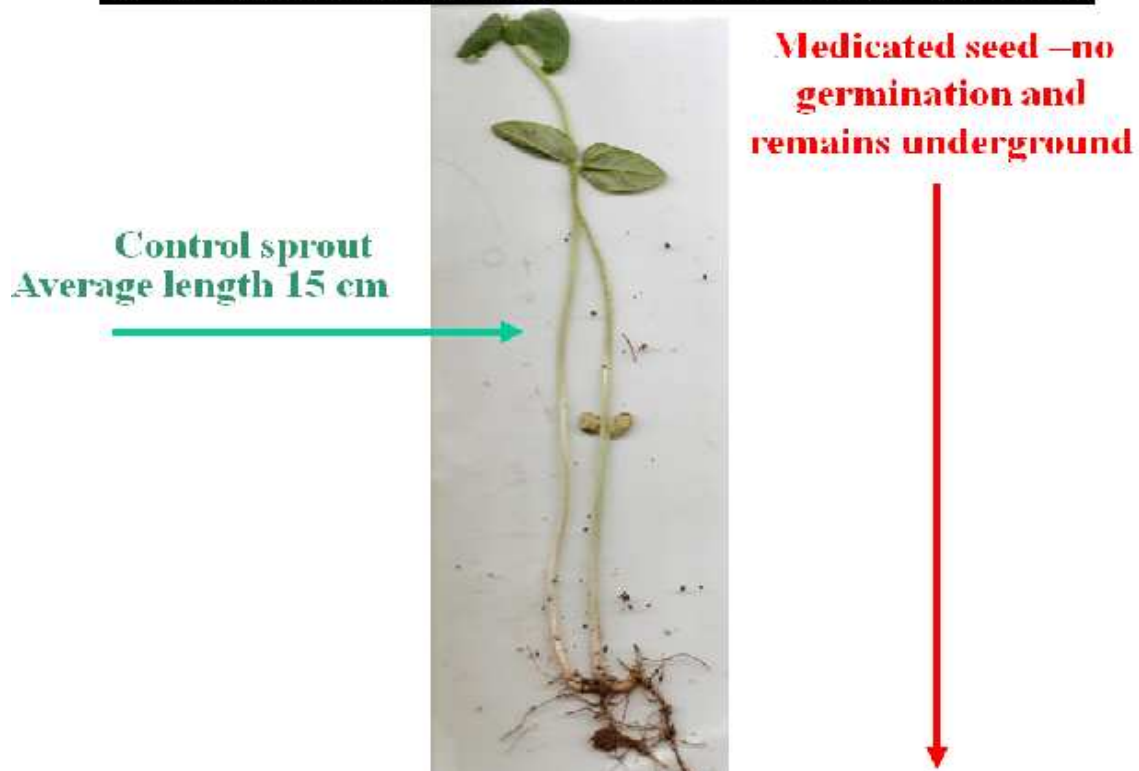
The experiment on chauli was repeated by soaking the seeds for 4 hours, continuously in distilled water and medicated water containing Natrum Mur+Acid Picric in 1M, 10M and CM potencies before sowing, the result was totally different. In this case right from germination stage till the adult stage (45 days), medicated plants grew faster. The rate of growth increased with potency (Figure 3).

### Deactivation



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**Retardation in the rate of germination in 5 days in chauli**  
**By medicated water (Natrum Mur CM+Acid Picric CM)**



**Figure 8**  
*Germination process was completely absent in case of medicated seeds of chauli.*



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**P. D. HINDUJA NATIONAL HOSPITAL & MEDICAL RESEARCH CENTRE**  
VEER SAVARKAR MARG, MAHIM, MUMBAI - 400 016, INDIA  
PHONE: 2644 1515, 2445 2222, 2444 9199 FAX: 2644 9151

**DEPARTMENT OF IMAGING**

NAME: [REDACTED] AGE: 16 YEARS SEX: FEMALE  
HM NO.: 658912 LOCATION: OPD ADM NO.: EXAM NO.: US 59930  
REFERRED BY: DR. NARVEKAR ORDER NO: 5425980 DATE: 02/08/2004

**\*\*EXAMINATION\*\*  
ULTRASOUND OF PELVIS**

**\*\* REPORT DETAILS \*\***

Realtime sonography of the pelvis was performed.

The uterus measures 6.8 x 3.2 x 2.2 cm in size and is normal in shape and echotexture. No intrauterine or parauterine mass lesion seen. Endometrial thickness is 3mm (normal).

The right ovary measures 3.8 x 3.5 x 2.5 cm and the left ovary measures 3.3 x 3.3 x 2.1 cm in size. Both ovaries are bulky with multiple small follicles arranged peripherally.

The urinary bladder distends well and is normal in shape and contour. The bladder wall is normal in thickness.

There is no evidence of free fluid in the pelvis.

**CONCLUSION:**  
1) FINDINGS SUGGESTIVE OF POLYCYSTIC OVARIAN DISEASE.  
2) NO OTHER SIGNIFICANT ABNORMALITY IS SEEN ON THIS STUDY.

*R. Sukthankar*  
DR. R SUKTHANKAR - M.D.  
CONSULTANT - RADIOLOGIST  
03/09/2004

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**P. D. HINDUJA NATIONAL HOSPITAL & MEDICAL RESEARCH CENTRE**  
VEER SAVARKAR MARG, MAHIM, MUMBAI - 400 016, INDIA  
PHONE: 2644 1515, 2445 2222, 2444 9199 FAX: 2644 9151

**DEPARTMENT OF IMAGING**

NAME: [REDACTED] AGE: 17 YEARS SEX: FEMALE  
HM NO.: 658912 LOCATION: OPD ADM NO.: EXAM NO.: US 70236  
REFERRED BY: DR. NARVEKAR ORDER NO: 7062850 DATE: 27/10/2005

**\*\*EXAMINATION\*\*  
ULTRASOUND OF PELVIS**

**\*\* REPORT DETAILS \*\***

Realtime sonography of the pelvis was performed.

The uterus measures 9.0 x 3.9 x 2.6 cms and is normal in shape and echotexture. No intrauterine or parauterine mass lesion seen. Endometrial thickness is 6 mm.

The right ovary measures 3.2 x 2.6 x 2.3 cms and is normal in shape and echotexture. The left ovary measures 3.3 x 3.2 x 1.7 cms in size and shows a dominant follicle measuring 2.4 x 1.3 cm.

The urinary bladder distends well and is normal in shape and contour. The bladder wall is normal in thickness.

Minimal free fluid in Pouch of Douglas.

**CONCLUSION:**  
- MINIMAL FREE FLUID IN POUCH OF DOUGLAS.  
- NO OTHER SIGNIFICANT ABNORMALITY IS SEEN ON THIS STUDY.

*N. Mehta*  
DR. NIRAD MEHTA - M.D.  
CONSULTANT - RADIOLOGIST  
28-10-2005

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Figure 9

Report after medication for six months indicate disappearance of cysts and normalization of ovary sizes

## CONCLUSION

Life cycle of agricultural plants is between 100 to 200 days in most of the cases. Hence effect of homoeopathic drugs on genetic activity in plant bodies can be evaluated quickly, clearly and safely as compared to that on human bodies whose life span is about 100 years. However, in case of human body also effect of drugs on stem cell gene can be concluded from a case of a 16-year girl whose menstruation was totally stopped due to multiple cysts on inflamed ovaries. She was treated with homeopathic drugs for six months. Her period started soon after the commencement of treatment and became regular and normal by the end of treatment. Relevant sonography reports (Figure-9) indicate presence and disappearance of cysts on ovaries and normalization of ovary sizes. In her case 8/10 drops of medicated water containing Oophorinum 30+Aurum Iodide 30 in 10/15 ml of plain water were administered twice a day.

The present study indicates that homoeopathic drugs are able to trigger genes to activate, accelerate, retard and deactivate its corresponding genetic process. This leads to endless possibilities in its obvious multiple uses in the lives of plants, animals and humans.



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