## **International Journal of Pharma and Bio Sciences**

## ISOLATION AND ANTIBIOGRAM PATTERN OF *E. COLI* ISOLATES HAVING HEAVY METALS TOLERANCE

## VIRENDER SINGH\*, P.K. CHAUHAN, SEEMA , ANKUR TYAGI, KESHAV THAKUR, ANU KUMAR AND VIVEK KUMAR.

<sup>\*</sup>Dept. of Microbiology, H.N.B. Garhwal University, Srinagar (U.K.) Dept. of Microbiology, Himachal Institute of Life Sciences, Paonta Sahib (H.P.) India.

\*Corresponding Author Virender\_micro83@yahoo.co.in.

## ABSTRACT

In the present study total eight heavy metal resistant *E. coli* were isolated from sewage of industrial effluents from waste water treatment plant of Paonta Sahib H.P. India, against chromium, copper, nickel, cadmium. All the isolates exhibited high resistance to heavy metals with minimum inhibitory concentration (MIC) for heavy metals ranging from 50  $\mu$ g/ml to 450  $\mu$ g/ml. All isolates showed multiple tolerances to heavy metal and were multi antibiotic resistant. Heavy Metal Tolerance Test indicated maximum microbial tolerance of *E. coli* (Ec-7) to Chromium (400  $\mu$ g/ml) and lowest to Copper (60  $\mu$ g/ml).

## **KEY WORDS**

Antibiotics resistance, multiple tolerance, Heavy metal resistant bacteria (HMRB).

## INTRODUCTION

The pollution of the environment with toxic heavy metals is spreading throughout the world along with industrial progress. Copper, chromium, cadmium and nickel are known to be the most commonly heavy metals used and the more widespread contaminants of the environment (Patterson, 1977; Aksu, 1998; Doenmez and Aksu, 1999). Traces of these heavy metals are necessary as Co-factors of enzymatic reactions, but high levels of them may cause extreme toxicity to living organisms due to inhibition of metabolic reactions. The microorganisms respond to these heavy metals by several processes; including transport across the cell membrane, biosorption to the cell walls and entrapment in extracellular capsules. precipitation, complexation and oxidationreduction reactions (Rai et al., 1981; Macaskie and Dean, 1989; Huang et al., 1990; Avery and Tobin, 1993; Brady et al., 1994; Veglio et al., **1997**). The bioremediation of heavy metals using microorganisms has received a great deal of attention in recent years, not only as a scientific novelty but also for its potential application in industry. Metal accumulative bioprocess generally falls into one of two categories, bisorptive (passive) uptake by nonliving, non growing biomass or biomass products and bioaccumulation by living cells (Macaski and Dean, 1989; Aksu and Kutsal, 1990; Huang et al., 1990; Volesky et al., 1992; Avery and Tobin, 1993; Brady and Duncan, 1994; Aksu, 1998; Doenmez and Aksu, 1999; 2001). Industrial wastes containing toxic metals were characterized by their differences rather than their similarities. These toxic metals can arise from a wide variety of industrial processes. The quality and the quantity of the wastes containing toxic heavy metals are dependent upon their industrial sources. Intrinsic bacteria, which are capable of metal accumulation, existing in soil on or near the site of contamination have adapting mechanisms to the contaminant. Naturally occurring bacteria that are capable of metal accumulation have been extensively studied since it is difficult to imagine that a single bacterium could be capable to remove all heavy metals from its polluted site (Clausen, 2000). the metal. which Apparently, has been introduced into the bacterial suspension by vigorous mixing, forms complexes with various ligands available (constituents which will complex heavy metal ions) (Hussein et al., 1998; 2001). Consequently, the largest amount of metals will be found as hydroxide or as a stable metal-ligand complex. Under a specific stress conditions, a relatively constant amount of metal reacts to stable and inactive complexes with active cellular components (Hussein, 1999). However, it is very important before the optimization of the bacterial growth process is to study at which pH value will be found as metal ions to study the real interaction between the free metal ions and the bacterial strain. This study aimed to isolate and characterization of *E. coli* sp resistant to heavy metals contaminants from industrial effluents waste water treatment plant of Paonta Sahib H.P.

### **MATERIALS AND METHODS**

### Sample collection

The sampling area was the sewage of industrial effluents waste water treatment plant of Paonta Sahib H.P. Samples was collected in sterile plastic bottles. A total of twenty samples were taken for the study.

## Isolation and identification of heavy metal resistant bacteria

For the selective isolation of heavy metals resistant bacteria, heavy metals incorporated media were used. The Nutrient agar incorporated with heavy metals like Cr6+, Ni2+, Cd2+ and Cu2+ and were prepared. The concentration of each heavy metal was maintained at 50  $\mu$ g/ml of the medium. The wastewater sample directly streaked on (N. A.) media and incubated at 37°C for 24 h. After the incubation period the plates were observed for any kind of growth on the media. The isolated and distinct colonies of *E. coli sp* on the N.A. media were sub cultured and obtain in the form of pure culture and identified

on the basis of their morphology and biochemical characters.

### Determination of Minimum Inhibitory Concentration (MIC)

MIC of the heavy metal resistant *E. coli* sp grown on heavy metals incorporated media, against respective heavy metal was determined by gradually increasing the concentration of the heavy metal, 10  $\mu$ g/ml each time on N.A. plate until the strains failed to give colonies on the plate. The starting concentration used was 50 $\mu$ g/ml. The culture growing on the last concentration was transferred to the higher concentration by streaking on the plate. MIC was noted when the isolates failed to grow on plates.

# Determination of antibiotic sensitivity and resistance pattern

Antibiotic sensitivity and resistance of the isolated heavy metal resistant isolates were assayed according to the Kirby-Bauer disc diffusion method given by Bauer et al. (1996).

After incubation, the organisms were classified as sensitive or resistant to an antibiotic according to the diameter of inhibition zone given in standard antibiotic disc chart.

## **RESULTS AND CONCLUSION:**

## Isolation and identification of heavy metals resistant bacteria

Eight heavy metal resistant *E. coli* sp were isolated from sewage of industrial effluents from waste water treatment plant of Paonta Sahib H.P. India, against chromium, copper, nickel, cadmium. All the isolates exhibited high resistance to heavy metals with minimum inhibitory concentration (MIC) for heavy metals ranging from  $50\mu$ g/ml to  $450\mu$ g/ml. All isolates showed multiple tolerances to heavy metal and were multi antibiotic resistant. Heavy Metal Tolerance Test indicated highest tolerance to Chromium ( $400\mu$ g/ml) by Ec -7 isolates and lowest to Copper by Ec -8.

### Table 1.

BACTERIA	<b>RESISTENCE TO</b>	MIC
Ec -1	Ni <sup>+2</sup>	180 µg / ml
	Cd <sup>2+</sup>	120 µg / ml
	Cu <sup>+2</sup>	180 µg / ml
	Cr <sup>+6</sup>	130 µg / ml
Ec -2	Ni <sup>+2</sup>	125 µg / ml
	Cd <sup>2+</sup>	120 µg / ml
	Cu <sup>+2</sup>	180 µg / ml
	Cr <sup>+6</sup>	120 µg / ml
Ec -3	Ni <sup>+2</sup>	110 µg/ml
	Cd <sup>2+</sup>	80 µg / ml
	Cu <sup>+2</sup>	220 µg/ml
	Cr <sup>+6</sup>	140 µg/ml
Ec -4	Ni <sup>+2</sup>	140 µg / ml
	Cd <sup>2+</sup>	90 µg/ml
	Cu <sup>+2</sup>	180 µg / ml

Resistance of bacteria to other heavy metals. (Ec= E. coli isolates)

	Cr <sup>+6</sup>	110 µg / ml
Ec -5	Ni <sup>+2</sup>	80 µg/ml
	Cd <sup>2+</sup>	150 µg / ml
	Cu <sup>+2</sup>	280 µg / ml
	Cr <sup>+6</sup>	120 µg / ml
Ec -6	Ni <sup>+2</sup>	200 µg / ml
	Cd <sup>2+</sup>	120 µg / ml
	Cu <sup>+2</sup>	300 µg / ml
	Cr <sup>+6</sup>	70g µg / ml
Ec -7	Ni <sup>+2</sup>	100 µg / ml
	Cd <sup>2+</sup>	120 µg / ml
	Cu <sup>+2</sup>	180 µg / ml
	Cr <sup>+6</sup>	400 µg/ml
Ec -8	Ni <sup>+2</sup>	250 µg / ml
	Cd <sup>2+</sup>	210 µg / ml
	Cu <sup>+2</sup>	60 µg / ml
	Cr <sup>+6</sup>	60 µg/ml

### Table 2.

Antibiotic sensitivity and resistant activity of heavy metal resistant E. coli sp

SR.N	BACTERIA	SENSTIVE TO	<b>RESITANT TO</b>
1	Ec -1	Meropenem, Bacitracin, Vancomycin, Chloramphenicol, Erythromycin, Tetracycline, Amikacin and kanamycin	Ampicillin,Cotrimoxazole, Ciprofloxacin,Tetracycline,
2	Ec -2	Ampicillin,Cotrimoxazole, Ciprofloxacin,Chloramphenicol, Erythromycin, Amikacin and kanamycin	Tetracycline, Meropenem,Bacitracin,Vanco mycin,
3	Ec -3	Ampicillin, Meropenem, Bacitracin, Vancomycin, Chloramphenicol, Erythromycin, Tetracycline and Amikacin	Cotrimoxazole, Ciprofloxacin, and kanamycin
4	Ec -4	Ampicillin, Cotrimoxazole, Ciprofloxacin, Meropenem, Bacitracin, Vancomycin and kanamycin	Chloramphenicol,Erythromycin ,Tetracycline, and Amikacin
5	Ec -5	Cotrimoxazole, Ciprofloxacin, Meropenem, Bacitracin, Vancomycin, Chloramphenicol and Erythromycin	Ampicillin, Tetracycline, Amikacin and kanamycin

6	Ec -6	Cotrimoxazole, Ciprofloxacin,Tetracycline, Meropenem, Bacitracin, Tetracycline, Amikacin and kanamycin	Ampicillin, Vancomycin, Chloramphenicol, Erythromycin,
7	Ec -7	Ampicillin,Cotrimoxazole, Ciprofloxacin, Vancomycin, Chloramphenicol, Erythromycin, Amikacin and kanamycin	Tetracycline, Meropenem, Bacitracin,
8	Ec -8	Ampicillin, kanamycin, Ciprofloxacin,Tetracycline, Meropenem,Bacitracin,Vancomyc in,Chloramphenicol,Erythromycin, Tetracycline,Amikacin.	Cotrimoxazole,

# Antibiotic sensitivity of heavy metals resistant isolates

All the isolates were resistant to antibiotics of which EC-8 was found to be single antibiotic resistant while the rest of the isolates were found to be multi-antibiotic resistant (Table 2). The sewage of industrial effluents waste water treatment plant of Paonta Sahib H.P. collects all the domestic as well as industrial wastewater from Paonta region. The wastewater coming from domestic and industrial sources is the environment appropriate where the microorganisms can develop resistance to heavy metals and antibiotics. The presence of small amount of antibiotics and heavy metals in the wastewater induce the emergence of Antibiotic resistance of heavy metal resistant isolates antibiotic resistant and heavy metal resistant microorganisms. Most of the isolates in the present study showed multiple tolerances to both heavy metals and antibiotics. The microbial resistance to heavy metal is attributed to a variety of detoxifying mechanism developed by resistant microorganisms such as complexation by xopolysaccharides, binding with bacterial cell envelopes, metal reduction, metal efflux etc.

These mechanisms are sometime encoded in plasmid genes facilitating the transfer of toxic metal resistance from one cell to another (Silver, 1996). Filali et al. (1999) studied wastewater bacteria isolates Psuedomonas aeroginosa, Klebsiella pneumoniae, Proteus mirabilis and Staphylococcus resistant to heavy metals and antibiotics. Similarly, Sharma et al. (2000) isolated highly cadmium resistant Klebsiella that was found to precipitate significant amount of CdS. The heavy metal resistant organism could be a potential agent for bioremediation of heavy metals pollution. Multiple tolerances occur only to toxic compounds that have similar mechanisms underlying their toxicity. Since heavy metals are all similar in their toxic mechanism, multiple tolerances are common phenomena among heavy metal resistant bacteria. In wastewater, there are some substances that have the potential to select for antibiotic resistance even though they are not antibiotics themselves. Heavy metals and biocides are two of them. The exposure to heavy metals or biocides results in the selection of bacterial strain also able to resist antibiotics. This happens because genes encoding heavy metals and biocides are located

together with antibiotic resistance genes or alternatively because bacteria can have unspecific mechanism of resistance common to different substances including heavy metals, biocides and antibiotics (Dalsgarrd and Guardbassi, 2002). It is therefore, likely that selective pressure by one such compound indirectly selects for the whole set of resistances.

### REFERENCES

- Aksu, Z. (1998). Biosorption of heavy metals by microalgae in batch and continuous systems. In: Algae for waste water treatment. eds Y.-S. Wong and N. F. Y. Tam, 99.37-53. Springer, Germany.
- 2 Aksu, Z. and Kutsal, T. (1990). A comparative study for biosorption characteristics of heavy metal ions with C. vulgaris. Environ. Technol., 11: 979-987.
- 3 Avery S.V. and Tobin, J.M. (1993). Mechanism of adsorption of hard and soft metal ions to Saccaromyces cerevisiae and influence of hard and soft anions. Appl. Environ. Microbiol, 59: 2851-2856.
- 4 Brady, D. and Duncan, J.R. (1994). Chemical and enzymatic extraction of heavy metal binding polymers from isolated cell walls of Sccharomyces cerevisiae. Biotechnol. Bioeng., 44: 297-302.
- 5 Brady D., Stoll A.D., Starke L. and Duncan, J.R. (1994). Bioaccumulation of metal cations by Saccaromyces cerevisiae. Appl. Microbiol. Biotechnol., 41: 149-154.
- 6 Clausen, C.A. (2000). Isolating metal-tolerant bacteria capable of removing copper,

chromium, and arsenic from treated wood. Waste Manage Res., 18: 264-268.

- Doenmez, G. and Aksu, Z. (1999). The effect of copper(II) ions on the growth and bioaccumulation properties of some yeasts. Process Biochem., 35: 135-142.
- 8 Dalsgarrd, A. and L. Guardbassi (2002). Occurrence and fate of antibiotics resistant bacteria in sewage. Danish, EPA Environment Project No. 722.
- 9 Doenmez, G. and Aksu, Z. (2001). Bioaccumulation of copper(II) and Nickel (II) by the non adapted and adapted growing Candida sp. Wat. Res., 35: 1425-1434.
- 10 Filali, B.K., J. Taoufik, Y. Zeroual, F.A.Z. Dzairi, M. Talbi, and M. Blaghen (1999). Waste water bacterial isolates resistant to heavy metals and antibiotics.*Current Microbiology 41*: 151-156.
- 11 Huang C., Huang, C. and Morehart, A.L. (1990). The removal of copper from dilute aqueous solutions by Saccharomyces cerevisiae. Wat. Res., 24: 433-439.
- 12 Hussein, H. (1999). Influence of heavy metals on the biodegradation of hazardous wastewater. Ph.D thesis, University of Alexandria.
- 13 Hussein, H., Krull, R., Abou-ElEla, S.I. and Hempel, D.C. (1998). Influence of heavy metal ions on the microbial degradation of xenobiotic waste water compounds, AWT98-Advanced wastewater treatment, recycling and reuse, Milano, 14-16 September.
- 14 Hussein, H., Krull, R., Abou-ElEla, S.I. and Hempel, D.C. (2001). Interaction of the different heavy metal ions with immobilised

bacterial culture degrading xenobiotic waste water compounds. Second International Water Asociation World Water Conference (2<sup>nd</sup> IWA) Berlin, 15-19 October.

- 15 Koedam, N., Wittouck, E., Gaballa, A., Gillis, A., Hfte, M. and Cornelis, P. (1994): Detection differentiation of microbial siderophores by isoelectric focusing and chrome azurol S overlay. Biometals, 7: 287-Krieg, N.K. (Ed.) (1984). Bergey`s manual of systematic bacteriology. Baltimore, Hong Kong, London, and sydney, 1: 178-182.
- 16 Macaskie, L. and Dean, A.C.R. (1989). Microbial metabolism, desolubilisation and deposition of heavy metals: Metal uptake by immobilised cells and application to the detoxification of liquid wastes. Adv. Biotechnol. Proc., 12: 159-172.
- 17 Patterson, J.W. (1977). Wastewater treatment technology. Ann Arbor science Publishers,Ann Arbor, MI, USA.

- Rai, L.C., Gaur, J.P. and Kumar, H.D. (1981). Phycology and heavy metal pollution. Biol. Rev., 56: 99-151.
- 19 Silver, S. (1996). Bacterial resistances to toxic metal ions-a review. *Gene 179*: 9-19.
- Sharma, K.P., A. Frenkel, and L.D. Balkwill (2000). A new *Klebsiella planticola* strain (cd-1) grows anaerobically at high cadmium concentrations and precipitates cadmium sulphide. *Applied and Environmental Microbiology 66*: 3083-3087.
- 21 Veglio, F., Beolchini, F. and Gasbarro, A. (1997). Biosorption of toxic heavy metals: an equilibrium study using free cells of Arthrobacter sp. Process Biochem., 32: 99-105.
- 22 Volesky, B., May, H. and Holan, Z.R. (1992).Cadmium biosorption by Saccharomyces cerevisiae. Biotechnol. Bioeng., 41: 826-829.