



RESEARCH ARTICLE

MICROBIOLOGY

DECOLOURISATION OF RAYON EFFLUENTS AND BLEACHING OF KRAFT PULP BY *STEREUM OSTREA***G.RAMA KRISHNA, S.V.S.S.S.L.HIMA BINDU N AND *M. A. SINGARA CHARYA**

Department of Microbiology Kakatiya University Warangal – 506 009

**M. A. SINGARA CHARYA**

Department of Microbiology Kakatiya University Warangal – 506 009

*Corresponding author

ABSTRACT

A potential lignolytic fungus, *Stereum ostrea* was analysed for the decolourisation of industrial effluents discharged from rayon pulp mills. The colour removal was from 5580.85 to 2690.05 CU during 40 days of incubation time. The laccase activity during the decolorization process was increased up to 20th day (862.77 U/L) and after that it was very low. Chemical bleaching was effective and showed 92-94% of brightness with viscosity 14 cp, while biobleaching of pulp recorded 78% of brightness with 4 cp viscosity during 40 days of solid state fermentation. The rayon sheet, after biobleaching was prepared and 76% brightness and 8 cp viscosity was noticed, which was accepted with moderate response in industries.



KEY WORDS

Decolorization, *Stereum ostrea*, kraft pulp, rayon effluents, biobleaching.

INTRODUCTION

Billions of gallons of toxic and intensely colored waste effluents are released into the rivers and streams annually by the pulp and paper industries¹. The primary contributor to the colour and toxicity of these aquatic systems is the pulp bleach plant effluents (BPE), which contains largely high molecular weight, modified and chlorinated lignin and its degradation products^{2, 3, 4, 5}. These toxic effluents are generated during paper and rayon manufacturing process and also in the bleaching of pulp with conventional chemical methods in different stages. The bleaching agents used in pulp and paper industries are chlorine, alkali, hypochlorite and hydrogen peroxide. The use of chlorine based chemicals in the bleaching process generate chloro-lignin or chloro phenol compounds which are completely resistant to microbial attack and remain as recalcitrants^{6, 7}. Therefore, environmental concerns have led us to seek alternative ways to eliminate or at least reduce, the use of chlorine-based chemicals in bleaching of pulp. The current biotechnological approach is concerned to tackle this problem. The two phased solutions are being developed; one is biobleaching of the pulp^{8, 9} which can be used as alternate method for chemical bleaching to avoid the generation of toxic effluents and the second is biotreatment of bleach plant effluents¹⁰ for their decolourisation and dechlorination.

White rot basidiomycete fungi are the only known organisms which are capable of degrading lignin by releasing lignolytic enzymes such as laccase, lignin peroxidase (LiP) and manganese peroxidase (MnP)^{11, 12, 13, 14}. Lignolytic fungi, *Phanerochaete chrysosporium* and *Trametes versicolor* can efficiently decolourise and dechlorinate BPE^{15, 16, 17, 18, 19, 20}. Because of the importance of lignin as a renewable source for the production

of paper, feeds, chemicals and fuels, there has been an increasing research emphasis on the fungal degradation of lignin. Delignification of lignocellulosic materials by white rot fungi is of great interest and has been investigated to improve the digestibility of wood or straw for animal feed^{21, 22, 23, 24, 25}. Lignin biodegradation by *Phanerochaete chrysosporium* and *Trametes versicolor* has been intensively studied to delignify and brighten unbleached kraft pulp (UKP)^{26, 27, 28}. Better lignin-degrading systems were identified in different fungi for use in various biotechnological applications^{29, 30, 31, 32}.

In view of the above findings, a potential lignolytic fungus, *Stereum ostrea* was analysed for the decolourisation and biobleaching of bleach plant effluents and unbleached kraft pulp of rayon industries. The medium used in this experiment was optimized for maximum laccase production, comparing to other white rot fungi, *Phanerochaete chrysosporium*, *Coriolus versicolor* or *Phlebia radiata*.

MATERIAL AND METHODS

Collection of basidiomycete:

The fruit bodies of *Stereum ostrea* were collected in rainy season from the surroundings of Warangal. The fruit bodies were preserved by sealing in polythene bags and noted in our departmental fungal herbarium.

Identification of white rot fungi:

Fungus was identified based on the fruit body characters and spore print^{33, 34, 35, 36}.

Culturing of fungal strain:

A small piece of fruiting body was dipped in 0.01% mercuric chloride to remove the surface contamination and washed several times with distilled water to remove the traces of HgCl₂



and transferred aseptically onto 3% malt agar slants. Slants were incubated for 5 to 7 days and initial growth of mycelium was observed on 5th day.

Maintenance of fungi:

The mycelium collected from the growing edge was transferred onto new malt agar slant and incubated further 5 to 7 days and slants were stored at 4°C until use. The culture was sub cultured at an interval of 30 days before being utilized as an inoculum.

Decolorisation studies:

The liquid and solid-state fermentation systems were followed in the studies on decolourisation of effluents and bleaching of pulp.

Liquid fermentation system:

Primary out let effluents released from the pulp bleaching plant of A.P. Rayon factory was used for the liquid cultures. This effluent released from the different bleaching stages contains pulp in diluted concentration and is used for secondary recovery of the pulp. Malt medium containing 33.3% of this effluent (control medium) supplemented with Cu⁺⁺ (2 mgL⁻¹) was used as liquid culture. Each 250 mL of flask containing 100 mL medium was inoculated with three pieces of 2 mm² fresh fungal mycelium from ten days precultured malt agar plates. An uninoculated medium served as blank. Decolourisation was observed up to 40 days. Culture harvests of different days were filtered and filtrates were centrifuged for 15 min. at 3000 rpm for 30 min. and reduction in colour was read at 465 nm with reference blank. Colour units were calculated³⁷ by using conversion factor 4,015 standard platinum cobalt colour units per A₄₆₅ of 1.0.

Solid state fermentation system:

Biobleaching of the pulp in the solid-state fermentation system was followed³⁸. Fifteen mL of M2 medium supplemented with Cu⁺⁺ (2 mgL⁻¹) was added to each 250 mL conical flask containing 50 g of unbleached cooked pulp (brightness 45%, viscosity). Flasks were sterilized by autoclaving at 10 lbs pressure for 30 min. Flasks were inoculated with 2 mm² of fresh fungal mycelium from 10 days precultured malt agar plates. An uninoculated flask containing pulp was served as blank. Rate of decolourization was measured in terms of increase in brightness. After incubation with fungi, pulp samples were washed with water and pulp sheets were prepared with a porcelain Buchner funnel (diameter, 11 mm) and air dried. Brightness was determined with Aimil glass reflectance meter (Cat.No.070, S.No.95358). Viscosity of the pulp was measured by the method developed by department of quality control, A.P. Rayon factory, using capillary viscometer. 0.255 g of pulp was dissolved in 25 mL of cuprammonium solution and 2-3 drops of pyrogalla (1:3 diluted) was added and stirred for 15 min. on magnetic stirrer. This solution was used to measure the flow time of the solution. The flow time was converted into centipoises (cp) using standard chart.

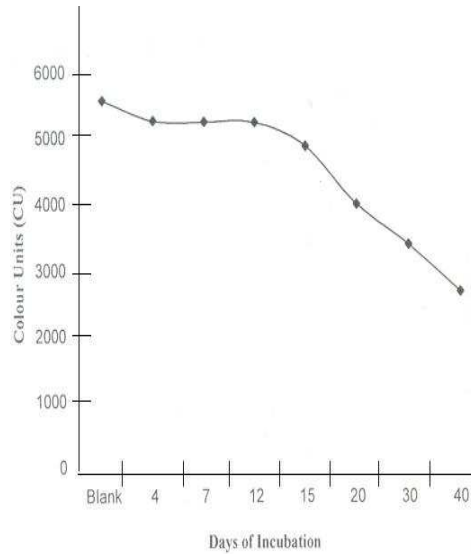
RESULT AND DISCUSSION

Decolourisation of effluent was studied in liquid and solid cultures up to 40 days. In liquid cultures of *Stereum ostrea* the colour reduced from 5580.85 to 2690.05 CU (Fig.1). Decolourisation was 5.6% in 4th day cultures and, the reduction in colour was very less up to 12th day. On 40th day of incubation the percentage of decolourisation was maximum (50%).



Fig. 1

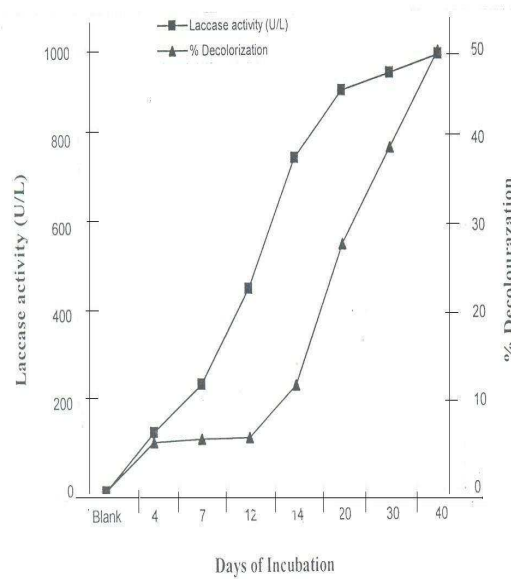
Decolorisation of bleachery effluents by *Stereum ostrea* in different days of incubation



In the comparison between laccase production and decolourisation, it was noticed that the laccase activity in culture filtrates was gradually increased up to 20th day of incubation (862.77 U/L) and the raise in laccase activity was very low between 20th and 40th day of incubation. The increase in laccase activity within 20 days of incubation was 86.94% and between 20th to 40th day it was 13% (Fig. 2).

Fig. 2

Correlation between laccase activity and decolorisation by *Stereum ostrea* in different days of incubation





The relationships between brightness of the pulp and viscosity changes during chemical bleaching were studied and recorded (Table 1a). By chemical bleaching the percent of brightness was gradually increased (45-94%) and at the same time there was a reduction in the viscosity. The viscosity should be optimum

in the pulp to prepare and strengthen the quality of rayon sheets. In the final step of chemical treatment, the brightness of the rayon sheet was 92-94% and viscosity was 14 cp, which was ideal for industrial rayon sheet synthesis.

Table 1**a) Brightness and viscosity changes during chemical bleaching of kraft pulp**

Bleaching stage	Brightness (%)	Viscosity (in centipoises)
Unbleached pulp	45	35
Chlorination stage (Cl ₂)	55	28
Oxygenated H ₂ O ₂ stage (EOP)	68	21
Hypo I stage	85	18
ClO ₂	89	16
Extracted peroxidase stage (EP)	90	15
Rayon sheet	92-94	14

The correlation between brightness of the pulp and its viscosity status during solid state fermentation (SSF) with *S.ostrea* was recorded (Table 1b). The brightness of the pulp was improved from 45-78%, while viscosity loss was 35-4.4 cp. Though the manufacture of

the rayon sheet was possible with this viscosity, the sheet strength and quality were not up to the mark. This is due to utilization of cellulose by the growing organism which converts the cellulose to sugars for its easy carbon assimilation.

Table 1**b) Brightness and viscosity changes during biobleaching by *Stereum ostrea* during solid state fermentation of kraft pulp**

Days of incubation	Brightness (%)	Viscosity (in centipoises)
Blank	45	35
4	49	23.4
7	54	11.8
12	71	9.2
15	73	8.8
20	76	8
25	78	6.2
30	78	5.5
40	78	4.4



The decolourization of pulp by *S.ostrea* during 40 days of SSF was presented in plate 1A. The activity of the organism to utilize the lignin and initiation of brightness was started after 7 days of incubation period and gradually it reached to 78% by 40 days. A comparison in the rayon sheet prepared with unbleached kraft pulp and biobleached pulp after 20days of incubation was presented in plate 1B. In this biobleached rayon sheet the brightness percent was 76 and viscosity was 8 cp. This was considered as moderately acceptable

process of pulp for the rayon preparation. Here, the critical point is improvement of brightness of the pulp without causing any damage to the cellulose,

Though the process was viewed as cost-effective (by comparison with chemicals), renewable and eco-friendly, care must be taken to maintain the viscosity of the pulp for quality rayon sheet. For this purpose the *S.ostrea* was subjected to mutations by ultra violet radiations for the development of cellulose minus strains (data not presented).

Plate 1A

Decolourisatation of kraft pulp by Stereum ostrea in different days of incubation (50 g of pulp + 15 mL of M2 medium)

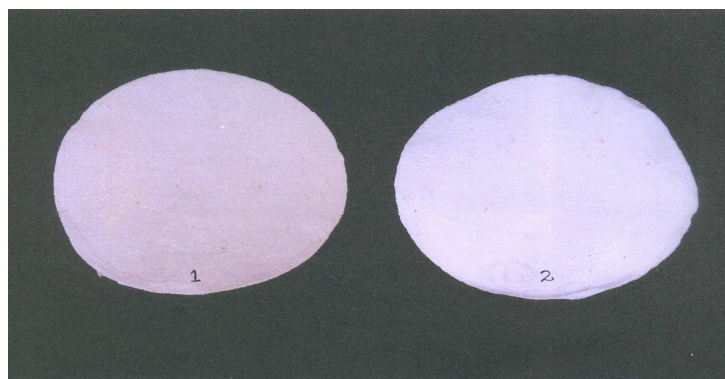


A

0: Blank; 1: Four days; 2: Seven days; 3: Twelve days;
4: Fifteen days; 5: Twenty days; 6: Twenty five days;
7: Thirty days; 8: Forty days.

Plate 1B

Comparison between unbleached (1) and bleached (2) pulp sheet with Stereum ostrea after twenty days of incubation



B



A number of previous studies^{20, 39, 40}, have shown that *Phanerochaete chrysosporium* as well as some other white rot fungi rapidly decolourise BPE. BPE from first alkali extraction stage after chlorination was decomposed to low molecular weight colourless products¹⁶. *Phanerochaete chrysosporium* was immobilized^{2,17} on partially submerged rotating disks to decolourise industrial BPE. The role of lignin peroxidase and manganese peroxidase activities were not reported in any of these previous studies on decolourisation or degradation of lignin based effluents. The present study is aimed at the delignification and decolorisation of rayon effluent and unbleached kraft pulp by ligninolytic enzymes produced by *Stereum ostrea*. The culture conditions were optimized for the maximum production of laccase. The results indicated that the *Stereum ostrea* can decolorize the diluted effluents up to 50% in 40 days of incubation. But the laccase activity in this culture was increased up to 20 days of incubation but remained almost constant after 20th day. This suggests that the decolourisation and laccase production are not happening simultaneously. These results are comparable with the reports available on decolourisation of kraft bleach plant effluent^{41, 42, 43, 44}.

Effluent cultures were used with final concentration of 3000 CU and achieved 85% decolourisation with lignin peroxidase and manganese peroxidase produced by *Phanerochaete chrysosporium*³⁷. Ninety percent was achieved decolorisation in CEH effluent with *Stagnospora gigaspora*, a coelomycetous fungi¹⁰. *Phanerochaete chrysosporium* was used for biobleaching in liquid-state fermentation system and reported that kappa number was decreased by 60% after a subsequent alkali treatment⁴⁵. *Trametes versicolor* in liquid-state fermentation system achieved 15 point increase in brightness⁴⁶. The same level of cumulative manganese peroxidase production by *Phanerochaete chrysosporium* and *Trametes versicolor* in the solid-state and liquid-state

fermentation systems, but observed negligible increase in brightness in the liquid-state fermentation system⁴⁰.

Most studies on the biobleaching of UKP by *Phanerochaete chrysosporium* and *Trametes versicolor* were carried out in the liquid-state fermentation systems with 15% pulp consistencies^{39,45}. On the other hand biobleaching of the unbleached kraft pulp in the solid-state fermentation afforded a marked brightness increase^{40,46}. The sequential treatment of softwood kraft pulps with laccase resulted in lower kappa number and higher brightness⁴⁷. Therefore, the present study aimed at biobleaching of unbleached kraft pulp by *Stereum ostrea* in the solid state fermentation system with little supplementation of nitrogen limited medium with Cu⁺⁺ ions (2 mgL⁻¹). It was recorded that maximum increase in brightness was in the solid-state conditions. These results supported the concept of effective lignin-degradation under solid-state fermentation by white-rot fungi^{48, 49}. The results further clarified that the biobleaching process can significantly increase the brightness of the pulp and reduce the use of chlorine based chemicals and pollution load of waste liquor. The white rot fungus, IZU-154, degraded wood lignin more extensively and selectively than *Phanerochaete chrysosporium* and *Trametes versicolor*⁵⁰. In addition, the use of chlorine based chemicals were reduced in bleaching of UKP in the biobleaching process, which combined a IZU-154 treatment and chemical bleaching^{51, 52, 53}.

A solid state bioreactor design for application in industrial process was attempted⁵⁴. The viscosity of the pulp is the crucial factor in the preparation of rayon sheets, which dropped substantially during solid state fermentation by *Stereum ostrea*. This may be due to the presence of cellulose enzyme which are acting on the cellulose fibrils of the pulp and reducing the viscosity. The enzyme production, brightness and viscosity losses are not in a line during the delignification of kraft pulp by *Stereum ostrea* in different incubation periods. The correlation



between degradation of lignin with effluent decolorization by white rot fungi was studied, where they used the black liquor and pulp effluents to observe the colour removal⁵⁵. The decolorization of triaryl methane, indigo, azo and anthraquinone dyes by *Trametes hirsute* was recorded⁵⁶. The waste water of textile industry has been decolorized efficiently by white rot fungus, *Irpex lacteus* without adding any chemicals and recorded⁵⁷ the decolorization of dye effluents by shaking (59%) and stationary cultures (93%).

The pulp bleaching in chlorine free sequence using laccase mediator system of *Pycnoporus cinnabarinus*, *Trametes versicolor* and *Pleurotus eryngii* and recorded⁵⁸ best preservation of cellulose and the largest removal of residual lignin content. The degradation and decolorization of synthetic effluents consisting of mixture of ten dyes by using *Trametes villosa* and *Pycnoporus sanguineus* was found the potentiality of these fungi for environmental decontamination⁵⁹. The treatment of the effluents from kraft bleach plant with *Pleurotus sajor-caju* and *P.ostreatus* and recorded the decolorization by 57 and 76

percent respectively after 14 days of incubation⁶⁰. The extracellular enzyme activities and ability to degrade the synthetic dyes of a white rot fungus, *Grammothelium subargentea* was reported that this organism is a useful tool in the pulp industry and bioremediation⁶¹. The biological decolorization of Cibacron black W-NN using five commercial *Pleurotus* species was reported among which *P. sajor-caju* fully decolorized the dye⁶². The industrial and biotechnological applications of lignolytic enzymes of the basidiomycete members were studied to replace the conventional chemical processes of several industries⁶³. The colour reduction of paper industry effluents by white rot fungus, *Sporotrichum pulverulentum* was showed that aeration and sterilization played an important role in the colour removal⁶⁴.

ACKNOWLEDGEMENTS

Authors are grateful to the Head, Department of Microbiology, Kakatiya University for providing necessary facilities.

REFERENCES

1. Springer A, Industrial environmental control, pulp and paper industry, Jon Wiley and Sons, Inc., New York, (1985).
2. Campbell A G, Jr, A bench scale evaluation of a process for decolorization of bleach plant effluent using the white rot fungus *Phanerochaete chrysosporium*. Ph. D. dissertation, North Carolina State University, Raleigh, (1983).
3. Paice M G and Jurasek L, Peroxidase-catalyzed color removal from bleach plant effluent, *Biotechnol Bioeng*, 26:477-480, (1984).
4. Huynh VB, Chang H M, Joyce T W and Kirk T K, Dechlorination of chloro-organics by a white rot fungus, *Tappi J*, 68:98-102, (1985).
5. Boominathan K and Reddy C A, Lignin degradation by fungi: biotechnological applications. In: D.K. Arora, K.G. Mukerji and R.P. Elander (ed.), *Handbook of applied mycology*, Vol.4, Biotechnology, Marcel Dekker, Inc., New York, (1991).
6. Ander P, Eriksson K E, Kolar M C and Kringstad K P, Studies on the mutagenic properties of bleaching effluents, *Sven Papperstidn*, 80:454-459, (1977).
7. Rappe C, Swanson S, Glass B, Kringstad K P, Sousa F D, Johansson L and Abe Z, On the formation of PCDDs and PCDFs in the bleaching of pulp, *Pulp Pap Can*, 90:T273-T278, (1989).
8. Leisola M S A, Brown C, Lausila M, Ulmer D and Fiecher A, Polysaccharide synthesis by *Phanerochaete chrysosporium* during degradation of kraft lignin, *Appl Environ Microbiol*, 15: 180-184, (1982).
9. Kirkpatrick N, Reid I, Ziomek E, Ho C and Paice M G, Relationship between fungal biomass production and the brightening of



- hardwood kraft pulp by *Coriolus versicolor*, Appl Environ Microbiol, 55:1147-1152, (1989).
10. Bergbauer M, Eggert C and Kraepelin G, Biotechnology in pulp and paper manufacture, In: T.K. Kirk, and H.M. Chang, (ed), Butterworth – Heinemann, Boston, 1990, pp.263-269.
 11. Tein M and Kirk T K, Lignin-degrading enzyme from the hymenomycete *Phanerochaete chrysosporium* burds, Science, 221:661-663, (1983).
 12. Gold M H, Kuwahara M, Chily A A and Glenn J K, Purification and characterization of extracellular H₂O₂ -requiring diaryl propane oxygenase from the white rot basidiomycete, *Phanerochaete chrysosporium*, Arch Biochem Biophys, 234:353-362, (1984).
 13. Reinhammar B, Laccase. In: R. Lontie (ed.), Copper proteins and copper enzymes, Vol.3, CRC Press, Inc., Boca Raton, Fla,1984,pp.1-35.
 14. Dobson P J, Evans C S, Harvey P J and Palmer J M, Production and properties of an extracellular peroxidase from *Coriolus versicolor* which catalyzes cleavage in a lignin model compound, FEMS Microbiol Lett, 42:17-22, (1987).
 15. Lundquist K, Kirk T K and Connors W J, Fungal degradation of kraft lignin and lignin sulfonates prepared from synthetic ¹⁴C-lignins, Arch Microbiol, 112:291-296, (1977).
 16. Sundman G, Kirk T K and Chang H M, Fungal decolorization of kraft bleach plant effluent, Tappi J, 64:145-148, (1981).
 17. Eaton D C, Chang H M, Joyce T W, Jefferies T W and Kirk T K, Method obtains fungal reduction of the color of extraction stage kraft bleach effluents, Tappi J, 65:89-92, (1983).
 18. Livernoche D, Jurasek L, Desrochers M, Dorica J and Veliky I A, Removal of color from kraft mill waste waters with cultures of white rot fungi and with immobilized mycelium of *Coriolus versicolor*, Biotechnol Bioeng, 25:2055-2065, (1983).
 19. Yin C F, Joyce T W, Chang H M, Kinetics of bleach plant effluent decolorization by *Phanerochaete chrysosporium*, J Biotechnol, 10:67-76, (1989).
 20. Archibald M, Paice G and Jurasek L, Decolorization of kraft bleachery effluent chromophores by *Coriolus (Trametes) versicolor*, Enzyme Microb Technol, 12:846-853, (1990).
 21. Agosin E, Monties B and Odier E, Structural changes in wheat straw components during decay by lignin – degrading white rot fungi in relation to improvement of digestibility for ruminants, J Sci Food Agric, 36:925-935, (1985).
 22. Zadrazil E, Screening of fungi for lignin decomposition and conversion of straw into feed, Ann Rev Bot, 59:433-452, (1985).
 23. Valmaseda M, Almendros G and Martinez A T, Chemical transformation of wheat straw constituents after solid state fermentation with selected lignocellulose degrading fungi, Biomass Bioenergy, 1: 261-266, (1991).
 24. Jung H J G, Valdez F R, Abad A R, Blanchette R A and Hatfield R D, Effect of white rot basidiomycetes on chemical composition and in vitro digestibility of oat straw and alfalfa stems, J Anim Sci, 70:1928-1935, (1992).
 25. Vares T, Kalsi M and Hatakka A, Lignin peroxidases, Manganese peroxidases, and other ligninolytic enzymes produced by *Phlebia radiata* during solid state fermentation of wheat straw, Appl Environ Microbiol, 61:3515-3520, (1995).
 26. Eriksson K E and Lindholm U, Lignets mikrobiela nedbrytning, Sven Papperstidn, 74:701-706, (1971).
 27. Perez J and Jeffreids T W, Roles of manganese and organic acid chelators in regulating lignin degradation and biosynthesis of peroxidases by *Phanerochaete chrysosporium*, Appl Environ Microbiol, 58:2402-2409, (1992).
 28. D' souza T M, Merritt C S and Reddy C A, Lignin – modifying enzymes of the white rot basidiomycete *Ganoderma lucidum* Appl Environ Microbiol, 65:5307-5313, (1999).
 29. Orth A B, Royse D J and Tien M, Ubiquity of lignin-degrading peroxidases among various wood degrading fungi, Appl Environ Microbiol, 59:4017-4023, (1993).



30. Hatakka A, Lignin-modifying enzymes from selected white rot fungi: production and role in lignin degradation, *FEMS Microbiol Res*, 13:125-135, (1994).
31. Perez F, Martinez M J and Martinez A T, Screening of 68 species of basidiomycetes involved in lignin degradation, *Mycol Res*, 99:37-42, (1995).
32. D' souza T M, Boominathan K and Reddy C A, Isolation of laccase gene specific sequences from white rot and brown rot fungi by PCR, *Appl Environ Microbiol*, 62:3739-3744, (1996).
33. Alexopolous C J, *Introductory Mycology*, IInd edition, John Wiley and Sons, New York, (1962).
34. McNabb R F R and Talbot P H B, *Holobasidiomycetidae-Exobasidiales, Brachybasidiales, Dacrymycetales, Tulasnellales*. In: *The fungi*, Vol, IVB, Eds. G.C.Ainsworth, F.K.Sparrow and A.S.Sussman, Academic Press, New York, 1973, pp.317-325.
35. Singer R, *The Agaricales in Modern Taxonomy*, 3rd edition, J. Cramer, Weinheim, (1975).
36. Mehrotra R S and Aneja K R, *An Introduction to Mycology*, New Age International, New Delhi, 1990.
37. Michel F C Jr, Dass S B, Grulke E A and Reddy C A, Role of manganese peroxidases and lignin peroxidases of *Phanerochaete chrysosporium* in the decolorization of kraft bleach plant effluent, *Appl Environ Microbiol*, 57: 2368-2375, (1991).
38. Nagarathnamma R and Bajpai P, Decolorization and detoxification of extraction stage effluent from chlorine bleaching of kraft pulp by *Rhizopus oryzae*, *Appl Environ Microbiol*, 65:1078-1082, (1999).
39. Kirk T K and Yang H H, Partial delignification of unbleached kraft pulp with ligninolytic fungi, *Biotechnol Lett*, 1:347-352, (1979).
40. Katagiri N, Tsutsumi Y and Nishida T, Correlation of brightening with cumulative enzyme activity related to lignin biodegradation during biobleaching of kraft pulp by white rot fungi in the solid-state fermentation system, *Appl Environ Microbiol*, 61:617-622, (1995).
41. Jager A, Croan S and Kirk T K, Production of ligninases and degradation of lignin in agitated submerged cultures of *Phanerochaete chrysosporium*, *Appl Environ Microbiol*, 50:1274-1278, (1985).
42. Kelley R L and Reddy C A, Identification of glucose oxidase activity as the primary source of hydrogen peroxide production in ligninolytic cultures of *Phanerochaete chrysosporium*, *Arch Microbiol*, 144:248-253, (1986).
43. Bonnarme P and Jeffries T W, Mn (II) regulation of lignin peroxidases and manganese-dependent peroxidases from lignin-degrading white rot fungi, *Appl Environ Microbiol*, 56:210-217, (1990).
44. Lamar R T, Larsen M J and Kirk T K, Sensitivity to and degradation of pentachlorophenol by *Phanerochaete* sp., *Appl Environ Microbiol*, 56:3519-3526, (1990).
45. Kirk T K, Schultz E, Connors W J, Lorenz L F and Zeikus J G, Influence of culture parameters on lignin metabolism by *Phanerochaete chrysosporium*. *Arch Microbiol*, 117:277-285, (1978).
46. Paice M G and Jurasek L, Ho C, Bourbonnais R and Archilbald F, Direct biological bleaching of hardwood kraft pulp with the fungus *Coriolus versicolor*, *Tappi J*, 72:217-221, (1990).
47. Hunt K, Lee C L, Bourbonnais R and Paice M G, Pulp bleaching with dimethyldioxirane and lignin oxidizing enzymes, *J Pulp Pap Sci*, 24:55-59, (1998).
48. Keyser P, Kirk T K and Zeikus J G, Ligninolytic enzyme system of *Phanerochaete chrysosporium*: synthesized in the absence of lignin response to nitrogen starvation, *J Bacteriol*, 135:790-797, (1978).
49. Reid I D, Solid state fermentation for biological delignification, *Enz Microb Technol*, 11:786-803, (1989).
50. Nishida T, Kashino Y, Mimura A and Takahara Y, Lignin biodegradation by wood rotting fungi. I. Screening of lignin



- degrading fungus, *Mokuzai Gakkaishi*, 34:530-536, (1988).
51. Fugita K, Kondo R, Sakai K, Kashino Y, Nishida T and Takahara Y, Biobleaching of soft wood kraft pulp with white rot fungus IZU-154, *Tappi J*, 74:124-127, (1991).
 52. Murata S, Kondo R, Sakai K, Kashino Y, Nishida T and Takahara Y, Chlorine-free bleaching process of kraft pulp with the fungus IZU-154, *Tappi J*, 75:91-94, (1992).
 53. Fugita K, Kondo R, Sakai K, Kashino Y, Nishida T and Takahara Y, Biobleaching of soft wood kraft pulp with white rot fungus IZU-154, *Tappi J*, 76:81-84, (1993).
 54. Rivela I, Rodriguez Counto S and Sanroman A, Extracellular ligninolytic enzyme production by *Phanerochaete chrysosporium* in a new solid-state bioreactor, *Biotech Lett*, 22:1443-1447, (2000).
 55. Ernie Martani and Sebastian Margino, Decolorization of pulp mill effluents by white rot fungi and its correlation to lignin degradation, *Biologi* 2: 265-276, (1998).
 56. Abadulla Elias, Tzanko Tzanov, Silgia Costa, Karl-Heinz Robra, Artur Cavaco-Paulo and Georg M Gu" Bitz, Decolorization and detoxification of textile dyes with a laccase from *Trametes hirsuta*, *Appl Environ Microbiol*, 66:3357-3362, (2000).
 57. Kwang-Soo Shin, The Role of Enzymes Produced by White-Rot Fungus *Irpex lacteus* in the Decolorization of the Textile Industry Effluent, *The J Microbiol*, 42:37-41, (2004).
 58. Susana Camarero, Olga Garcia, Teresa Vidal, José Colom, José C del Rio, Ana Gutiérrez, José M Gras, Rebeca Monje, Maria J. Martinez, Ángel T Martinez, Efficient bleaching of non-wood high-quality paper pulp using laccase-mediator system, *Enz Microb Technol*, 35: 13-120, (2004).
 59. Kátia M G Machado, Luciana C A Compart, Rúbio O Morais, Luiz H Rosa, Mércia H Santos, Biodegradation of reactive textile dyes by basidiomycetous fungi from Brazilian ecosystems, *Brazilian J Microbiol*, 37:481-487, (2006).
 60. Belem A, Panteleitchouk A V, Duarte A C, Rocha-Santos T A P and Freitas A C, Treatment of the effluent from a kraft bleach plant with white rot fungi *Pleurotus sajor caju* and *Pleurotus ostreatus*, *Global NEST J*, 10:426-431, (2008).
 61. Mario C N Saparrat, Paulina Mocchiutti, Constanza S Liggieri, Mo'nica B Aulicino, Ne'stor O Caffini, Pedro A Balatti, Mari'a Jesu's Marti'nez, Ligninolytic enzyme ability and potential biotechnology applications of the white-rot fungus *Grammothele subargentea* LPSC no. 436 strain, *Proc Biochem*, 43:368-375, (2008).
 62. Halil Bıyık, Fatih Kalyoncu, Erman Oryasin, Nuri Azbar, Erbil Kalmıs and Gamze Basbülbül, Evaluation of wild and commercial types of *Pleurotus* strains for their ability to decolorize cibacron black WNN textile dye, *Afr J Microbiol Res*, 3:325-329, (2009).
 63. Maciel M J, Silva A C and Ribeiro H C T, Industrial and biotechnological applications of lignolytic enzymes of the basidiomycetes: A review, *Electronic J Biotechnol*, 13: 1-12, (2010).
 64. Rajendra Bhai D Vasait, A biotechnological approach for the removal of color of effluent from a paper industry by using white rot fungi, *Int Res J*, 1:20-23, (2010).