



RESEARCH ARTICLE

ENTOMOLOGY

STUDIES ON THE COMPARATIVE FEED EFFICACY OF *BOMBYX MORI* (L.) (LEPIDOPTERA: BOMBYCIDAE) FED WITH SILVER NANOPARTICLES (AGNPS) AND *SPIRULINA* TREATED MR₂ MULBERRY LEAVES IN RELATION TO GROWTH AND DEVELOPMENT

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ABSTRACT

Silkworm *Bombyx mori* is an important economic insect and also a tool to convert leaf protein into silk. The industrial and commercial use of silk, the historical and economic importance of production and its application in all over the world finely contributed to the silkworm promotion as a powerful laboratory model for the basic research in biology. The mulberry leaf is the only important source of the silkworms. The present study has been undertaken to the effectiveness of foliar applications of silver nanoparticles (AgNps) (25%) and single cell protein (SCP) containing microalgae *Spirulina* (0.03%) treated MR₂ mulberry leaves fed with the popular Indian bivoltine hybrid (CSR2×CSR4) silkworm *B. mori* larvae (3rd, 4th and 5th instar) to study the larval growth and development (length, width and weight) and economic parameters like cocoon (length, width and weight) shell and pupal weight were significantly increased than control group. This study was indicated that the AgNps and *Spirulina* exhibit the presence of certain growth stimulant and can be used to increase the silk yield in commercial silkworm rearing.



KEYWORDS

Bombyx mori, Silver Nanoparticles (AgNps), *Spirulina*, Mulberry leaves.

INTRODUCTION

The silkworm rearing is a traditional industry in Asia and the life of many people is depended on it. Increase of larval growth and cocoon quality and quantity would result better economics for this industry and meet the production needs. Consequently, the enrichment of mulberry leaves by supplementary compounds with the aim of increasing the production of cocoon is a very important aspect. Many investigations have been done on this topic and various reports have been published [1-3]. Fortification of mulberry leaves with complementary compounds was found to increase the larval growth and post cocoon characters [1, 4].

Morus alba L. (mulberry), a woody plant belonging to the genus *Morus*, family Moraceae, is a widely distributed plant in China, whose leaves, root bark and branches have long been used in Chinese medicine to treat fever, protect the liver, improve eyesight, facilitate discharge of urine and lower blood pressure [5-6]. The mulberry leaves are rich in flavonoids, alkaloids and polysaccharides components which are known as the most potent major active compounds by chemical constituent investigations [7]. Among those, the flavonoids like rutin, quercetin, isoquercitrin and quercetin 3-(6-malonylglucoside) [8] are found in mulberry leaves. The health benefits of flavonoids are well known and are displayed as a remarkable range of biochemical and pharmacological properties that may significantly affect the function of various mammalian cells [9]. The anti-inflammatory, antioxidant, antithrombotic and anticarcinogenic effects of flavonoids are some of the properties that have been under

consideration in view of therapeutical purposes for several human diseases [10]. Flavonoids are thought to be one of the most critical constituents in mulberry that have therapeutic functions. But the anti-fatigue effects of flavonoids from mulberry leaves (FML) have also been reported [11].

Silkworm mortality is mainly caused by bacteria and other pathogenic microorganisms. AgNps have antimicrobial activity against several pathogenic microorganisms. Antibacterial properties of silver are documented since 1000 B.C., when silver vessels were used to preserve water. The first scientific papers describing the medical use of silver report the prevention of eye infection in neonates in 1881 and internal antisepsis in 1901. After this, silver nitrate and silver sulfadiazine have been widely used for the treatment of superficial and deep dermal burns of wounds and for the removal of warts [12]. The term “silver nanomaterials” refers to any silver-containing materials with enhanced activity due to their nanoscale features. In some cases, commercial products containing metallic AgNps in the range of 5–50 nm or ionic silver are given the name ‘nanosilver’ [13]. AgNps mode of action is presumed to be dependent on Ag^+ ions, which strongly inhibit bacterial growth through suppression of respiratory enzymes and electron transport components and through interference with DNA functions [14]. AgNps in a nanometric scale (less than 100 nm) has different catalytical properties compared with those attributed to the bulk form of the noble metal, like surface plasmon resonance, large effective scattering cross section of individual AgNps,



and strong toxicity to a wide range of microorganisms [15].

Mulberry leaf supplemented with *Spirulina* as a feed to *B. mori* L. (Lepidoptera: Bombycidae) orally found to be effective in enhancing the larval and cocoon characters. *Spirulina*, blue-green algae contains 18 amino acids viz., glutamine, glycine, histidine, lysine, methionine, creatine, cysteine, phenylalanine, serine, proline, tryptophan, asparagine, pyruvic acid and vital vitamins like biotin, tocopherol, thiamine, riboflavin, niacin, folic acid, pyridoxoic acid, beta-carotene and vitamin B₁₂ [16]. Various researches have been carried out on the diet supplementation of mulberry leaves fed to silkworms. These supplementations include vitamins such as ascorbic acid, thiamine, niacin, folic acid and multivitamins. Its nutrients are very easy to digest protein (biliprotein), carbohydrates (mucopolysaccharides, rhamnose and glycogen), 50 different minerals and trace minerals, beta-carotene, chlorophyll, GLA omega3 fatty acid, and many other nutrients [2]. The dietary supplements like protein, vitamins, lipids etc. evinced their specificity at specific dose for various metabolic activities of silkworm [17]. Amino acids such as aspartic acid and glutamic acid are considered to be essential for silkworm growth [18]. Nutritional study on silkworm is an essential prerequisite for its proper commercial exploitation. Nutrition of silkworm is sole factor which almost individually augment quality and quantity of silk [19].

MATERIALS METHODS

Silkworm rearing method

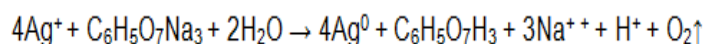
The eggs of popular Indian bivoltine hybrid (CSR2×CSR4) silkworm *B. mori* were collected from farmers training centre at Jayankondapattinam, Tamilnadu, India. The eggs were placed at ambient temperature of

25±2° C and relative humidity of 70 to 80% in an incubator for hatching. After hatching, larvae were isolated from stock culture. The larvae were divided into 3 experimental groups including control (distilled water treatment), each group consisting of 6 larvae. The larvae were reared in cardboard boxes measuring 22×15×5 cms covered with nylon net and placed in an iron stand with ant wells. The control and treated MR₂ mulberry leaves were fed to silkworms, five feedings/day. They were maintained up to cocoon stage.

Preparation of Silver Nanoparticles and *Spirulina* solution

Silver Nitrate AgNO₃ (Sigma Aldrich, UK) and Trisodium Citrate C₆H₅O₇Na₃ (Sigma Aldrich, UK) of analytical grade purity, were used as starting materials without further purification. The AgNps were prepared by using chemical reduction method according to the description of Lee and Meisel [20]. All solutions of reacting materials were prepared in distilled water. In typical experiment, 50 ml of 1·10⁻³ M AgNO₃ was heated to boiling. To this solution, 5 ml of 1% Trisodium Citrate was added drop by drop. During the process, solution was mixed vigorously. Solution was heated until colour change is evident (pale yellow). Then it was removed from the heating element and stirred until cooled to room temperature.

Mechanism of reaction could be expressed as follows:



Spirulina powder were purchased from Pharmaceutical company at Chidambaram and to prepare the experimental dose for 0.03% concentration.

Mulberry (*Morus alba* L.) MR₂ leaves treated with Silver Nanoparticles (AgNps) and *Spirulina* (SCP)



AgNps and *Spirulina* were diluted to 25% and 0.03% concentrations respectively; fresh MR₂ mulberry leaves were soaked in this two concentrations for 15 minutes and then were dried in air for 10 minutes. The treated leaves were used for feeding the 3rd, 4th and 5th instars larvae of silkworm *B. mori*. They were maintained up to cocoon stage.

Experimental groups

There are three experimental groups. 3rd, 4th and 5th instars of *B. mori* larvae fed with the following treated MR₂ mulberry leaves. Group T₁ larvae fed with distilled water treated mulberry leaves, it serve as a control, group T₂ larvae fed with 25% AgNps treated mulberry leaves, group T₃, larvae fed with 0.03% *Spirulina* solution treated mulberry leaves. They were maintained up to cocoon stage.

Statistical analysis

Data were analyzed by One-Way Analysis Of Variance (ANOVA) followed by Duncan's multiple range test (DMRT) using a commercially available statistics software package (SPSS® for Windows, V. 16.0,

Chicago, USA). Results were presented as means ± SD. P values < 0.05 were regarded as statistically significant.

RESULTS

Larval Parameters

Morphometric analysis of 3rd, 4th and 5th instar larvae

Table 1 displays that the morphometric data of control MR₂ leaves, AgNps and *Spirulina* treated MR₂ leaves fed 3rd instar *B. mori* larvae length, width and weight. The mean value of group T₁ were (1.6833±0.1472cm, 0.3500±0.0548cm and 0.1117±0.0075gm), respectively. The mean value of group T₂ were (1.9333±0.1633cm, 0.3833±0.0408cm and 0.1233±0.0103gm), respectively. The mean value of group T₃ were (1.7500±0.1049cm, 0.3500±0.0548cm and 0.1167±0.0121gm), respectively. In these three observations, 25% AgNps treated 3rd instar larvae length were significantly increased than the other two groups (T₁ and T₃).

Table 1
Morphometric data of AgNps and Spirulina treated 3rd instar of *B. mori* larvae.

Groups	3 rd instar of <i>B. mori</i> larvae		
	Length (cm) (Mean ± S.D)	Width (cm) (Mean ± S.D)	Weight (gm) (Mean ± S.D)
Control (T ₁)	1.6833±0.1472 ^a	0.3500±0.0548 ^a	0.1117±0.0075 ^a
MR ₂ Leaves + 25% AgNps (T ₂)	1.9333±0.1633 ^b	0.3833±0.0408 ^a	0.1233±0.0103 ^a
MR ₂ Leaves + 0.03% <i>Spirulina</i> (T ₃)	1.7500±0.1049 ^a	0.3500±0.0548 ^a	0.1167±0.0121 ^a

Values are Mean ± Standard Deviation of six observations. Values in the same column with different superscript letters (a & b) differs significantly at P<0.05 (DMRT).

Table 2 shows that the morphometric data of control MR₂ leaves, AgNps and *Spirulina* treated MR₂ leaves fed 4th instar *B. mori* larvae length, width and weight. The mean value of group T₁ were (5.7333±0.2422cm,

0.5333±0.0816cm and 0.4233±0.0344gm), respectively. The mean value of group T₂ were (6.0167±0.1472cm, 0.6167±0.0753cm and 0.5450±0.0404gm), respectively. The mean value of group T₃ were (5.8833±0.1941cm,

0.5667±0.0816cm and 0.4883±0.0183gm), respectively. In these three observations, 25% AgNps treated 4th instar larvae length,

width and weight were significantly increased than the other two groups (T₁ and T₃).

Table 2
Morphometric data of AgNps and Spirulina treated 4th instar of *B. mori* larvae.

4 th instar of <i>B. mori</i> larvae			
Groups	Length (cm) (Mean ± S.D)	Width (cm) (Mean ± S.D)	Weight (gm) (Mean ± S.D)
Control (T ₁)	5.7333±0.24 22 ^a	0.5333±0.0816 ^a b	0.4233±0.0344 ^a
MR ₂ mulberry + 25% AgNps (T ₂)	6.0167±0.14 72 ^b	0.6167±0.0752 ^b	0.5450±0.0404 ^c
MR ₂ Leaves + 0.03% <i>Spirulina</i> (T ₃)	5.8833±0.19 41 ^{ab}	0.5667±0.0816 ^a	0.4883±0.0183 ^b

Values are Mean ± S.D of six observations. Values in the same column with different superscript letters (a, b & c) differs significantly at P<0.05 (DMRT).

Table 3 shows that the morphometric data of control MR₂ leaves, AgNps and *Spirulina* treated MR₂ leaves fed 5th instar of *B. mori* larvae length, width and weight. The mean value of group T₁ were (6.7167±0.2483cm, 1.0333±0.1211cm and 2.8350±0.0855gm), respectively. The mean value of group T₂ were (7.2500±0.1871cm, 1.1333±0.0816cm and

3.5583±0.2369gm), respectively. The mean value of group T₃ were (7.0500±0.1871cm, 1.0667±0.0816cm and 3.4467±0.2965gm), respectively. In these three observations, 25% AgNps and 0.03% *Spirulina* treated 5th instar larvae length and weight were significantly increased than control group (T₁).

Table 3
Morphometric data of AgNps and Spirulina treated 5th instar of *B. mori* larvae.

5 th instar of <i>B. mori</i> larvae			
Groups	Length (cm) (Mean ± S.D)	Width (cm) (Mean ± S.D)	Weight (gm) (Mean ± S.D)
Control (T ₁)	6.7167±0.2483 ^a	1.0333±0.1211 ^a	2.8350±0.0855 ^a
MR ₂ mulberry + 25% AgNps (T ₂)	7.2500±0.1871 ^b	1.1333±0.0816 ^a	3.5583±0.2369 ^b
MR ₂ Leaves + 0.03% <i>Spirulina</i> (T ₃)	7.0500±0.1871 ^b	1.0667±0.0816 ^a	3.4467±0.2965 ^b

Values are Mean ± S.D of six observations. Values in the same column with different superscript letters (a & b) differs significantly at P<0.05 (DMRT).

Cocoon and pupal parameters

Table 4 shows that the morphometric data of the control MR₂ leaves, AgNps and *Spirulina* treated MR₂ leaves fed *B. mori* larvae was found to be produced cocoon length, width and weight. The mean value of group T₁ larvae produced cocoon were (3.4000±0.3847cm, 2.1333±0.1211cm and 1.5117±0.1007gm), respectively. The mean and of group T₂ larvae produced cocoon were (3.6667±0.1633cm,

2.4000±0.0894cm and 2.2117±0.3632gm), respectively. The mean value of group T₃ larvae produced cocoon were (3.5333±0.1862cm, 2.3000±0.0894cm and 1.9933±0.0975gm), respectively. In these three observations, 25% AgNps and 0.03% *Spirulina* treated larvae produced cocoon width and weight was significantly increased than control group (T₁).

Table 4
Morphometric data of AgNps and Spirulina treated *B. mori* larvae produced cocoon.

Groups	Cocoon of <i>B. mori</i>		
	Length (cm) (Mean ± S.D)	Width (cm) (Mean ± S.D)	Weight (gm) (Mean ± S.D)
Control (T ₁)	3.4000±0.384 7 ^a	2.1333±0.121 1 ^a	1.5117±0.1007 ^a
MR ₂ mulberry + 25% AgNps (T ₂)	3.6667±0.163 3 ^a	2.4000±0.089 4 ^b	2.2117±0.3632 ^b
MR ₂ Leaves + 0.03% <i>Spirulina</i> (T ₃)	3.5333±0.186 2 ^a	2.3000±0.089 4 ^b	1.9933±0.0975 ^b

Values are Mean ± S.D of six observations. Values in the same column with different superscript letters (a & b) differs significantly at P<0.05 (DMRT).

Table 5 shows that the morphometric data of the control MR₂ leaves, AgNps and *Spirulina* treated MR₂ leaves fed *B. mori* larvae produced cocoon shell and pupal weight. The mean value of group T₁ larvae produced cocoon shell and pupal weight were (0.3883±0.0376gm and 1.1233±0.1256gm), respectively. The mean value of group T₂ larvae produced cocoon shell and pupal weight were (0.4767±0.0308gm and

1.5683±0.0673gm), respectively. The mean value of group T₃ larvae produced cocoon shell and pupal weight were (0.4250±0.0187gm and 1.5542±0.0964gm), respectively. In these three observations, 25% AgNps treated larvae produced cocoon shell weight was significantly increased than other two groups (T₁ and T₂). But 25% AgNps and 0.03% *Spirulina* treated larvae produced pupa weight was significantly increased than control group (T₁).

Table 5
Morphometric data of AgNps and Spirulina treated *B. mori* larvae produced cocoon shell and pupal weight.

Groups	Cocoon shell Weight (gm) (Mean \pm S.D)	Pupal Weight (gm) (Mean \pm S.D)
Control (T ₁)	0.3883 \pm 0.0376 ^a	1.1233 \pm 0.1256 ^a
MR ₂ mulberry + 25% AgNps (T ₂)	0.4767 \pm 0.0308 ^b	1.5683 \pm 0.0673 ^b
MR ₂ Leaves + 0.03% <i>Spirulina</i> (T ₃)	0.4250 \pm 0.0187 ^a	1.5542 \pm 0.0964 ^b

Values are Mean \pm S.D of six observations. Values in the same column with different superscript letters (a & b) differs significantly at $P < 0.05$ (DMRT).

DISCUSSION

In the present study, the larval and cocoon length, width and weight were significantly increased in some groups. Many researchers showed that the larval characters improve by different concentrations of complementary compounds such as ascorbic acid, folic acid, thiamin, vitamin B complex etc., [2, 21, 22]. Muniandy *et al.* [23] have been reported that multi-vitamins and mineral compounds could increase the food intake, growth and conversion efficiency of silkworm. In the present study, it has been observed that silkworms fed by the particular dose of AgNps (25%) and *Spirulina* (0.03%) have enhanced the larval, cocoon and pupal characters were concomitantly increased from 3rd to 5th instars, suggested that AgNps and *Spirulina* which were stimulate silkworm to feed more amount of nutrients intake than the control. This work is corroborated with Nirwani and Kaliwal [22], suggested that this enhancement in larval, cocoon and pupal parameters related to phagostimulation of folic acid. Several authors also reported these effects about ascorbic acid [24-27].

AgNps attach the surface of pathogenic bacterial cell membrane and penetrate to release the silver ions and disturb the functions of bacteria [28-29]. The AgNps target the bacterial cell membrane, leading to a dissipation of the proton motive force, Consequently AgNps need to reach the cell membrane to achieve an antibacterial effect [28, 30]. Considering that the bacterial cell membrane is the site of active transport, respiratory chain components, energy transducing systems, membrane stages in the biosynthesis of phospholipids, peptidoglycan, LPS and capsular polysaccharides, and the anchoring for DNA [31], an alteration of the membrane's integrity would have a great impact in bacterial growth. The AgNps were effective in inhibiting bacterial growth in a dose and time dependent manner [32]. The Pathogenic bacterial cells exposed to millimolar Ag⁺ doses suffer morphological changes such as cytoplasm shrinkage and detachment of cell wall membrane, DNA condensation and localization in an electron-light region in the center of the cell, and cell membrane degradation allowing leakage of intracellular contents [33, 34]. Physiological changes occur together with the morphological changes. Bacterial cells enter an active, but non-culturable state in which physiological levels



can be measured but bacteria are not able to grow and replicate. The AgNps enter the gut of silkworm *B. mori* from AgNps (25%) treated MR₂ mulberry leaves it inhibits the pathogenic bacterial growth and multiplication to prevent the bacterial diseases and enhance the feed efficacy, larval growth rate, silk protein synthesis and silk production.

Since most of these multi-vitamin compounds are composed of ascorbic acid, it could be thought that the increase in larval weight is due to an enhancement of feeding activity. Although, in treated larvae the vitamins as cofactors can facilitate the metabolic pathway. Similar findings have also been observed in the present study that AgNps and *Spirulina* act as vitamins to stimulate the feeding activity in the silkworms. Therefore, AgNps and *Spirulina* can improve the food digestibility and increase the larval, cocoon and pupal parameters. *Spirulina* is a rich source of protein and the most powerful and well-balanced source of nutrition. One kg of *Spirulina* is said to be equivalent to 1000 kg of assorted vegetables [35]. The growth-promoting effect of water-soluble proteins and vitamins viz., B₂, B₆ and C are found in *Spirulina* and it is treated on silkworm with vitamins and amino acids enhance the larval and cocoon length, width and weight, cocoon shell and pupal weight. Therefore, in the end, the study recommends that 0.03% concentration of aqueous solution of *Spirulina* as feed to silkworm found to effectively increase single larval and cocoon weight, shell and pupal weight. The specific dose of *Spirulina* 0.03% concentration contains the maximum amount of essential amino acids and vitamins which determine the specificity for various metabolic activities in silkworm *B. mori*.

In this study, cocoon parameters were changed in different treatments. Previously, it was reported that enrichment of mulberry leaves by some vitamins could increase the cocoon yield. Nirwani and Kaliwal [22] have determined that folic acid causes a significant increase in economical parameters such as female and male cocoon weight. Evanglista *et al.* [36] and Balasundaram *et al.* [37] have also reported that the larval and cocoon length, width and weight were increased under multi-vitamin and vitamin C treatments. The enrichment of mulberry leaves with AgNps and *Spirulina* increases the larval, cocoon and pupal parameters. Increase in these insects was related to metabolisms other than proteins. It is assumed that fortification of diet supports the metabolism of carbohydrates and lipids.

CONCLUSION

In conclusion, the AgNps and *Spirulina* could increase some biological characters of silkworm *B. mori*, but this enhancement could economically improve the Sericultural goals. In the present study, the treatment of AgNps and *Spirulina* at the concentration of 25% and 0.03% respectively may have beneficial effects on the growth of the silkworm larval, cocoon and pupal parameters and also increase the quantity of silk by enhancing the feed efficacy. So, this supplementation could be prescribed to the farmers to get more quantity of silk.

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REFERENCES

- [1] Etebari K. Effect of enrichment of mulberry leaves (*Morus alba*) with some vitamins and nitrogenous compounds on some economic traits and physiological characters of silkworm *Bombyx mori* (Lepidoptera: Bombycidae). Isfahan University of Technology, Iran; 2002.
- [2] Etebari K, Kaliwal B, Matindoost L. Supplementation of mulberry leaves in sericulture, theoretical and applied aspects. *Int J Indust Entomol* 2004; 9: 14-28.
- [3] Islam MR, Ohayed Ali MA, Paul DK, Sultana S, Banu NA, Islam MR. Effect of salt, nickel chloride supplementation on the growth of silkworm, *Bombyx mori* L. (Lepidoptera: Bombycidae). *J Biol Sci* 2004; 4: 170-172.
- [4] Etebari K, Fazilati M. Effect of feeding on mulberry's supplementary leaves with multi-mineral in some biological and biochemical characteristics of silkworm (*Bombyx mori*). *J Sci Technol Agric Natur Resour* 2003; 7: 233-244.
- [5] Jia ZS, Tang MC, Wu JM. The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals. *Food Chem* 1999; 64: 555-559.
- [6] Chen JJ, Li XG. Hypolipidemic effect of flavonoids from mulberry leaves in triton WR-1339 induced hyperlipidemic mice. *Asia Pacific J Clinical Nutrition* 2007; 16: 290-294.
- [7] Wang J, Wu FA, Zhao H, Liu L, Wu QS. Isolation of flavonoids from mulberry (*Morus alba* L.) leaves with macroporous resins. *African J Biotechnol* 2008; 7: 2147-2155.
- [8] Lee CY, Sim SM, Cheng HM. Systemic absorption of antioxidants from mulberry (*Morus alba* L.) leaf extracts using an in situ rat intestinal preparation. *Nutr Res* 2007; 27: 492-497.
- [9] Middleton E, Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease and cancer. *Pharmacol Ther* 2000; 52: 673-751.
- [10] Snijman PW, Swanevelder S, Joubert E. The antimutagenic activity of the major flavonoids of rooibos (*Aspalathus linearis*): Some dose-response effects on mutagen activation-flavonoid interactions. *Mutation Res* 2007; 631: 111-123.
- [11] Kerem Z, German-Shashoua H, Yarden O. Microwave-assisted extraction of bioactive saponins from chickpea (*Cicer arietinum* L.). *J Sci Food Agric* 2005; 85: 406-412.
- [12] Rai M, Yadav A, Gade A. Silver nanoparticles as a new generation of antimicrobials. *Biotechnol Adv* 2009; 27: 76-83.
- [13] Panyala NR, Pen˜a-Me˜ndez EM, Havel J. Silver or silver nanoparticles: A hazardous threat to the environment and human health. *J Appl Biomed* 2008; 6: 117-129.
- [14] Li Y, Leung P, Yao L, Song QW, Newton E. Antimicrobial effect of surgical masks coated with nanoparticles. *J Hosp Infect* 2006; 62: 58-63.
- [15] Elechiguerra JL, Burt JL, Morones JR, Camacho-Bragado A, Gao X, Lara HH. Interaction of silver nanoparticles with HIV-1. *J Nanobiotechnology* 2005; 3: 6.
- [16] Venkataramana P, Srinivasa Rao TVS, Srinivasula Reddy P, Suryanarayana N. Effect of *Spirulina* on the larval and cocoon characters of silkworm, *Bombyx mori* L. *Proc Nat Acad Sci India* 2003; 73B(1): 89-94.
- [17] Horie. Recent advances in sericulture. *Ann Rev Entomol* 1980; 23: 49-71.
- [18] Ito T, Inokuchi. Nutritive effects of D-amino acids on the silkworm, *Bombyx mori* L. *Journal of Insect Physiology* 1981; 27(7): 447-453.
- [19] Laskar N, Datta M. Effect of Alfalfa tonic and its organic ingredients on growth and development of silkworm *Bombyx mori* L.



- Race Nistari. Environment and Ecology 2000; 18(3): 591-596.
- [20] Lee PC, Meisel DJ. Phys Chem 1982, 86: 3391-3395.
- [21] Sarker A, Haque M, Rab M, Absar N. Effects of feeding mulberry (*Morus sp.*) leaves supplemented with different nutrients to silkworm (*Bombyx mori*) L. Curr Sci 1995; 69: 185-188.
- [22] Nirwani RB, Kaliwal BB. Effect of folic acid on economic traits and the change of some metabolic substances of bivoltine silkworm, *Bombyx mori* L. Korean J Seric Sci 1996; 38: 118-123.
- [23] Muniandy S, Sheela M, Nirmala ST. Effect of vitamins and minerals (Filibon) on food intake, growth and conversion efficiency in *Bombyx mori*. Environ Ecol 1995; 13: 433-435.
- [24] Dobzhenok NV. Effects of ascorbic acid on the physiological condition of the codling moth and its resistance to fungus and bacterial infection. Zakhist Roslin 1974; 19: 3-7.
- [25] Ito T. Silkworm Nutrition in the silkworm an important laboratory tool. Tazima Y (ed) Ko Ltd, Tokyo 1978; 121-157.
- [26] Singh T, Reddy GP. Feeding behaviour of castor semilooper, *Achoea janata* Linn. To sterols, ascorbic acid and castor leaves. Indian J Ectomol 1981; 50: 530-532.
- [27] El-Karaksy IA, Idriss M. Ascorbic acid enhances the silk yield of the mulberry silkworm *Bombyx mori*. J Appl Entomol 1990; 109: 81-86.
- [28] Sondi I, Salopek-Sondi B. Silver nanoparticles as antimicrobial agent: a case study on *E. coli* as a model for gram-negative bacteria. J Colloid Interface Sci 2004; 275: 177-182.
- [29] Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, Tam PK, Chiu JF, Che CM. Proteomic analysis of the mode of antibacterial action of silver nanoparticles. J Proteome Res 2006; 5: 916-924.
- [30] Lok CN, Ho CM, Chen R, He QY, Yu WY, Sun H, Tam PK, Chiu JF, Che CM. Silver nanoparticles: partial oxidation and antibacterial activities. J Biol Inorg Chem 2007; 12: 527-534.
- [31] Salton MRJ, Kim KS. Structure. In: Baron S, editor. Medical Microbiology, 2009.
- [32] Yamanaka M, Hara K, Kudo J. Bactericidal actions of a silver ion solution on *Escherichia coli*, studied by energy-filtering transmission electron microscopy and proteomic analysis. Appl Environ Microbiol 2005; 71: 7589-7593.
- [33] Feng Q, Wu J, Chen G, Cui F, Kim T, Kim J. A mechanistic study of the antibacterial effect of silver ions on *Escherichia coli* and *Staphylococcus aureus*. J Biomed Mater Res 2000; 52: 662-668.
- [34] Jung W, Koo H, Kim K, Shin S, Kim S, Park Y. Antibacterial activity and mechanism of action of the silver ion in *Staphylococcus aureus* and *Escherichia coli*. Appl Environ Microbiol 2008; 74: 2171-2178.
- [35] Vijayalakshmi M, Sherley PG, Santhy KS, Sarojini Sukumar. Impact of supplementation of *Spirulina* on silkworm, *Bombyx mori* L. Proceedings on "Sericulture Technology Transfer", Kongunadu College, India 2004.
- [36] Evangelista A, Carvalho AD, Takahashi R, De Carvalho AD. Performance of silkworm (*Bombyx mori* L.) fed with vitamin and mineral supplement. Revista de Agriculture Piracicaba 1997; 72: 199-204.
- [37] Balasundaram D, Selvisabhanayakam, Mathivanan V. Studies on comparative feed efficacy of mulberry leaves MR₂ and MR₂ treated with vitamin C on *Bombyx mori* (L.) (Lepidoptera: Bombycidae) in relation to larval parameters. J Curr Sci 2008; 12(2): 677-682.