

**BIOPRODUCTION OF HUMAN DEFENSINS ANTIMICROBIAL PEPTIDES IN
TRANSGENIC PLANTS**

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ABSTRACT

Human defensins are small antimicrobial and cytotoxic peptides produced by neutrophils and epithelial cells. They exhibit broad range of antimicrobial properties and are thought to be ideal therapeutic agents because of their potential ability to circumvent the problems of acquired resistance often observed with other antimicrobial therapies. Therefore studies have been started to clone and express these peptides in bacterial and plant systems. There are certain major hurdles of producing the defensins peptides in bacteria like toxic action of the peptide on the host cells, costs of production by these technologies and scale up is also very difficult. Transgenic plants as bioreactors pave a new way of producing recombinant therapeutic defensins peptides. The present review paper describes an overview of human defensins including its classification, molecular structure, biosynthesis, localization, mechanism of action, potential medical application and production of human defensins by r-DNA technology in bacterial and plant systems.

KEYWORDS

Antimicrobial peptides, Human Defensins and Transgenic plants

INTRODUCTION

AntiMicrobial Peptides (AMPs) are the cytolytic peptides that are produced by the living organisms of all types (vertebrates and invertebrates) including humans in response to infection and exert antibiotic like action against pathogenic microorganisms, e.g. bacteria, fungi, viruses and parasites¹⁻². They are produced on epithelial surface and in phagocytic cells and plays an important role in innate and adaptive defence systems³. Thus they are having both offensive and defensive function. **Human Defensins** are small, cationically charged, cysteine-rich endogenous antibiotic peptides with a molecular weight of 4-5 kDa⁴. In recent years considerable research has been started on human defensins because of their potential medical and pharmaceutical application⁵⁻⁸.

CLASSIFICATION:

The eukaryotic antimicrobial peptides are divided into four groups according to their structural features: Cysteine- free α helices, extended Cysteine- free α helices with a predominance of one or two amino acids, loop structures with one intramolecular disulfide bond, and β sheet structures which are stabilized by two or three intermolecular disulfide bonds⁹⁻¹⁰. **Human** defensins are part of the fourth group. The human defensins are classified into three main groups according to their structural differences: the α defensins, β defensins and θ defensins¹¹. To date six α defensins have been identified in humans. Four of these designated human neutrophil peptides (HNP) 1, 2, 3 & 4, form part of the armory of neutrophils, where they participate in systemic innate immunity. The remaining two, Human Defensins (HD) 5 and 6, are expressed in intestinal Paneth cells and probably contribute

to innate defense of the GI mucosal surface¹². The β defensins have been isolated from both leukocytes and epithelial cells and classified into four main types as Human β defensins (HBD) (HBD1-to HBD-4)¹³.

MOLECULAR STRUCTURE

Human defensins are 29-35 amino acids cationic peptide, including six invariant cysteins whose intramolecular disulfide bonds cyclise and stabilise them in a complexly folded, triple stranded β sheet configuration. The peptides HNP-1, HNP-2, and HNP-3 are rich in cystine, arginine, and aromatic residues, but are devoid of sulfhydryl groups and carbohydrate moieties¹⁴⁻¹⁶.

BIOSYNTHESIS:

The neutrophil granulocytes is the most numerous leukocytes in peripheral blood. The development from a multipotent progenitor cell to a mature neutrophil takes place in the bone marrow over a period of 10-14 days. Human α defensins (HNPs) are produced from 93-95 amino acids precursors by stepwise, tissue specific, proteolytic removal of 64 amino terminal residues to produce the mature defensins¹⁷⁻²⁰. The early proteolytic processing events include two sequential cleavages, each removing 19 amino terminal aa residues, that yield 75 and 56 aa prodefensins respectively. The first cleavage is the removal of a signal peptide and second by preaspartate proteolytic cleavage in the bone marrow²¹⁻²⁴. The subsequent processing steps occurs in the peripheral blood neutrophils and convert these 56 aa prodefensins to mature (30 aa) HNP-1 and HNP-3.

LOCALISATION:

Human defensins peptides have been found to be in the granules of phagocytes, intestinal paneth cells, on epithelial surfaces of the intestine and trachea²⁵. Microbes in the phagocytic vacuoles of granulocytes and certain macrophages encounter high concentrations of defensins²⁶. Increased transcription of defensin genes and stimulus dependent release of pre-synthesized defensin-containing cytoplasmic granules contribute to the local antimicrobial response²⁷. The α human defensins constitute more than 5% of total cellular protein in human Polymorpho Neutrophils (PMN). They comprise 30-50% of the total protein in the azurophilic granules of human neutrophils, the most abundant is HNP1²⁸⁻³⁰. Human neutrophil contain large amounts of three α defensins (HNP-1 to HNP-3), and a smaller amounts of a fourth, HNP-4. HNP 1-3 are detected in bone marrow cells by Northern blot analysis and in peripheral leucocytes by the reverse transcription-polymerase chain reaction³¹⁻³⁴. HNPs 1-3 are absent from mononuclear phagocytes systems such as peripheral monocytes, alveolar macrophages and Kuffler cells in liver³⁵. They are specific to the cells of neutrophil lineage and functions as microbicidal agents in the neutrophil mediated defense system³⁶⁻⁴¹. They are released upon neutrophil stimulation. Endogenous molecular forms of HNPs are isolated from normal bone marrow, plasma and peripheral blood neutrophils. Immunoperoxidase stains revealed HNP 1-3 to have a granular localization in the neutrophil's cytoplasm by light microscopy. Frozen thin section immunogold transmission electron microscopy showed HNP 1-3 to be localized in azurophilic granules⁴²⁻⁴⁹. Monocytes and macrophages generally lack defensins, but they release messengers that induce the synthesis of β defensins in epithelial cells⁵⁰⁻⁵⁹.

MECHANISM OF ACTION:

Human defensins AMPs are membrane active, They act by disrupting negatively charged target

cell surface molecule to which they are electrostatically attracted⁶⁰. Hydrophobic face of the amphipathic structure of defensins AMPs disrupts the lipid bilayer of bacterial cell wall; causing membrane depolarisation which leads to cell death⁶¹.

DIFFERENT THERAPEUTIC APPLICATIONS OF DEFENSINS:

All defensins are active against majority of gram positive and gram negative bacteria and thought to be an ideal candidate to be developed as an antimicrobial agent. Studies have suggested that defensins can be used as an adjunct to antitubercular drugs. The studies have shown that the combination of HNP-1, isoniazid, and rifampicin is active against *Mycobacterium tuberculosis H37Rv* in vitro, ex vivo, and in vivo, and synergism was observed on the basis of reductions in minimum inhibitory concentrations (MICs) of these agents⁶². Human beta defensin have been shown to contribute to the defence of the intestine against infection by luminal microsporidia spores⁶³. Some of the artificially synthesised α defensin peptide have also shown useful results against various microorganisms⁶⁴. Some reports also suggest that defensin antimicrobial peptides play an important role in host defence, particularly in the oral cavity where there is constant challenge by microorganisms⁶⁵. Some of the recent findings are that in defensin in addition to their antimicrobial and immunomodulatory effects, particularly, HNP-1–HNP-3 possess antiviral and toxin neutralizing properties⁶⁶. Some studies also suggest that HNP-1 along with some other agents like SgIII and DMT-1 are implicated in cell-mediated LDL oxidation for pathogenesis of atherosclerosis⁶⁷. Different studies have also suggested that antimicrobial defensin (HD 5 and HD 6) peptides are important in protection of the host against microbial invasion in states of intestinal inflammation⁶⁸.

PRODUCTION OF HUMAN DEFENSIN IN TRANSGENIC PLANTS:

Transgenic Plants are the plants that are produced in the lab using r-DNA technology in order to create plants with special characteristics by artificial insertion of genes from other species, and sometime entirely from different kingdom⁶⁹⁻⁷². The plants that are successfully used for making transgenic are rice, corn, tobacco, and soybean.

Agrobacterium tumefaciens is a plant pathogenic bacteria that causes crown gall disease in dicotyledonous plants. The disease is characterized by a tumorous growth on the infected plant. Molecular studies of the disease have revealed that crown gall tumour develops as a result of genetic transformation of the plant by the bacterium. The bacteria make a heritable change in the DNA of the plant and change results in the formation of tumour by the plant. Thus it is an example of naturally occurring genetic engineering and if we can clone our protein/peptides of interest in the Ti Plasmid i.e. Tumour Inducing plasmid of *A. tumefaciens* and genetically transform the selected plant for example tobacco, the tobacco plants will start expressing the protein/peptide of our interest.

Significance of Using *Nicotiana tabaccum* as Transgenic Plant: low cost fully functional recombinant proteins have been reported. e.g. relaxin does not serve as a host for human pathogen non food, non feed crop, it does not pose any danger of contamination. Basic peptides and protein can easily be extracted from acidic part. Various antimicrobial peptides and proteins that have been started to be cloned and expressed in the transgenic plants are vaccines (Hepatitis B & Rabies), erythropoietin, human growth hormone, interferons, and various interleukins⁷³⁻⁷⁶. First Genetically engineered tobacco plants with tolerance by expressing genes encoding for insecticidal proteins from *Bacillus thuringiensis*

was reported for the first time in 1987. Thereafter many transgenic plants have been developed by laboratory methods, for example in 1994 Flavr Savr tomato which was the first modern transgenic crop approved for sale in the USA with the advantage of longer shelf life⁷⁷. In 1995 Margherita Zanetti et al have produced cathelicidins **antimicrobial** peptides in transgenic tobacco. A defensin like peptide was produced by Cary et al in the year 2000 in *Nicotiana* plant. In 2002 Carla M et al cloned and expressed *SMAP-29*- a mammalian antimicrobial peptide of innate immunity⁷⁸. Chenming Zhang in 2005 had successfully produced large quantity of recombinant protein, prorelaxin in transgenic *Nicotiana* plant⁷⁹. In 2006 Gloria Arenas and Sergio H. Marshall have shown that a defensin like Antimicrobial Peptide from *Mytilus edulis-chilensis* was cloned into a binary vector and successfully used to transfer in *Nicotiana tabaccum* L. The resulting plant is showing protective effect from a bacterial pathogen *Pseudomonas syringae* pv. *Syringae*⁸⁰.

CONCLUSION

Thus the transgenic plants for production of therapeutic peptides are promising tool for large scale production of this peptide. Further production of defensin like peptide in transgenic plants will also increase immunity towards various biotic and abiotic stresses of plants. The plants will become more resistible for microbial infections. Since human defensin has role in increasing the immunity of the humans therefore plants with defensin expression in it will surely increase the immunity of the individuals who will take it. The purified defensins can also be used as a potential target to be developed as a drug.

REFERENCES

1. Anita G and Gowthaman R, Strategies for development of fungus-resistant transgenic plants, *Current Science*, 84 (3): 330-340, (2003).
2. Hong PJ, Brian CS, Andreas S, Rose L, Guthmillerd JM, Georgia KJ, Brian FT, Joseph PM, Andre R, Tomas GP and McCray BJ, Discovery of new human β defensins using a genomics-based approach, *Gene*, 263: 211-218, (2001).
3. Lalitha G, Kirti PB, Characterization of defensin (Tfgd2) from *Trigonella foenum-graecum* Sudar Olli, *Current Science*, 93(3): 365-369, (2007).
4. Kalita A, Verma I and Khuller GK, Role of Human Neutrophil Peptide-1 as a Possible Adjunct to Antituberculosis Chemotherapy, *The Journal of Infectious Diseases*, 190:1476–80, (2004).
5. Date Y, Nakazato M, Shiomi K, Toshimori H, Kangawa K, Matsuo H and Matsukura S, Localization of human neutrophil peptide (HNP) and its messenger RNA in neutrophil series, *Ann Hematol*, 69: 73-77, (1994).
6. Fionnuala TL, John N, Derek L, Brett G, Pat H and John JM, Antimicrobial activity of truncated α defensin (human neutrophil peptide (HNP)-1) analogues without disulphide bridges, *Molecular Immunology*, 45: 190–193, (2008).
7. Robert IL, Multispecific myeloid defensins, *Current Opinion in Hematology*, 14: 16–21, (2007).
8. Falco A, Masa V, Tafalla, Perez L, Coll JM and Estepa A, Dual antiviral activity of human alpha-defensin-1 against viral haemorrhagic septicaemia rhabdovirus (VHSV): Inactivation of virus particles and induction of a type I interferon-related response, *Antiviral Research*, 76: 111–123, (2007).
9. Kevin LP, Melissa HB and Robert EWH, Recombinant DNA procedures for producing small antimicrobial cationic peptides in bacteria, *Gene*, 134: 7-13, (1993).
10. Yi-Quan T, Jun Y, Christopher JM and Michael ES, Isolation, Characterization, cDNA Cloning, and Antimicrobial Properties of Two Distinct Subfamilies of α -Defensins from Rhesus Macaque Leukocytes, *Infection and Immunity*, 67, 11: 6139–6144, (1999).
11. Markus AH, Hofera J, Peter S, Katharina P and Gerhard JZ, Host antimicrobial proteins as endogenous immunomodulators, *Immunology Letters*, 119: 4–11, (2008).
12. Rose L, David M, Lide L and Gan T, The structure of neutrophil defensin genes, *Federation of European Biochemical Societies*, 321, 3: 267-273, (1993).
13. Jens MS, Epithelial Peptide Antibiotics, *Biochemical Pharmacology*, 57, 121–134, (1999).
14. Samantha L and Henry BL, Therapeutic peptides, *Trends in Biotechnology*, 21, (12): 556-56, (2003).
15. Tom WL, Groeneveld, Tamara HR, Leendert AT, Dafne LH, Vanessa B, Jan WD, Pieter SH, Mohamed RD and Anja R, Human neutrophil peptide-1 inhibits both the classical and the lectin pathway of complement activation, *Molecular Immunology*, 44: 3608–3614, (2007).
16. Fang XM, Shu Q, Chen QX, Book MH, Sahl G, Hoefft A and Stuber F, Differential expression of α and β defensins in human peripheral blood, *European Journal of Clinical Investigation*, 33: 82–87, (2003).
17. Alexander G, Martin K, Peter F, Bernd V, Hartmut S, Ulrich K, Eduard FS, Thomas M, Klaus F and Dominik MA, The Expression Patterns of Peritoneal

- Defensins, *Perit Dial Int*, 27: 654–662, (2007).
18. Cunliffe RN, Defensins in the gastrointestinal tract, *Molecular Immunology*, 40: 463–467, (2003).
 19. Lisa KR, Janice R, Meenakshi B and Gill D, Expression of β Defensin Genes in Bovine Alveolar Macrophages, *Infection and Immunity*, 66 (2): 878–881, (1998).
 20. Frye M, Bargon J, Lembcke B, Wagner TOF and Gropp R, Differential expression of human α and β defensins mRNA in gastrointestinal epithelia, *European Journal of Clinical Investigation*, 30: 695–701, (2000).
 21. Harder J, Bartels J, Christophers E and Schröder JM, A Peptide antibiotic from human skin, *Nature*, 387: 861, (1997).
 22. Thorgerdur S, Pia A, Mina D, Martin M, Artur S and Mikael B, In Silico Identification and Biological Evaluation of Antimicrobial Peptides Based on Human Cathelicidin LL-37, *Antimicrobial Agents And Chemotherapy*, 50(9): 2983–2989, (2006).
 23. Robert B, Xiaorong W, Michael Z and James MW, The peptide antibiotic LL-37yhCAP-18 is expressed in epithelia of the human lung where it has broad antimicrobial activity at the airway surface, *Proc. Natl. Acad. Sci. USA*, 95, 9541–9546, (1998).
 24. Margherita Z, Renato G, Domenico R, Cathelicidins: a novel protein family with a common proregion and a variable C-terminal antimicrobial domain, *FEBS Letters*, 374: 1-5, (1995).
 25. Reddy KVR, Yedery RD and Aranha C, Antimicrobial peptides: premises and promises, *International Journal of Antimicrobial Agents*, 24: 536–547, (2004).
 26. Brogden KA, Ackermann M, Paul BM and Brian FT, Antimicrobial peptides in animals and their role in host defences, *International Journal of Antimicrobial Agents*, 22: 465-478, (2003).
 27. Ganz T, Defensins: antimicrobial peptides of vertebrates, *Comptes Rendus. Biologies*, 327: 539–549, (2004).
 28. Tae-Joon P, Ji-Sun K, Sung-Sub C and Yongae K, Cloning, expression, isotope labeling, purification, and characterization of bovine antimicrobial peptide, lactophorin in *Escherichia coli*, *Protein Expression and Purification*, 65: 23–29, (2009).
 29. Arash I and Richard LG, Antimicrobial peptides, *The Journal of American Academy of Dermatology*, 10: 381-390, (2005).
 30. Ganz T and Robert I L, Antimicrobial peptides of vertebrates, *Current Opinion in Immunology*, 10:41-44, (1998).
 31. Ganz T and Robert I L, Antibiotic peptides from higher eukaryotes: biology and applications, *Molecular Medicine Today*, 5: 292-297, (1999).
 32. Gudmundur HG and Birgitta A, Neutrophil antibacterial peptides, multifunctional effector molecules in the mammalian immune system, *Journal of Immunological Methods*, 232: 45-54, (1999).
 33. Luis R, Ganz T, Eukaryotic antibiotic peptides: not only a membrane business, *DDT*, 4 (6): (1999)
 34. Robert EWH & Hans-Georg S, Antimicrobial and host-defense peptides as new anti-infective therapeutic strategies, *Nature Biotechnology*, 24 (12): 1551-1556, (2006).
 35. Michael ES & Andre JO, Mammalian defensins in the antimicrobial immune response, *Nature Immunology*, 6 (6): 551-557, (2005).
 36. John K. Spitznagel, Antibiotic Proteins of Human Neutrophils, *J. Clin. Invest*, 86: 1381-1386, (1990).
 37. Boman HG, Antibacterial peptides: basic facts and emerging concepts, *Journal of Internal Medicine*, 254: 197–215, (2003).

38. Xian CR, Shu L, Jin CH, Xiao LJ, Xiao MH, Jian JH, Chen ZJ, Jun MZ and Fu QH, A novel carrier molecule for high-level expression of peptide antibiotics in *Escherichia coli*, *Protein Expression and Purification*, 36: 11–18, (2004).
39. Sylvia SLH, Ganz T and Robert IL, Neutrophil Defensins: Purification, Characterization, and Antimicrobial Testing, *Methods In Enzymology*, 236: 161- 172, (1998).
40. Li-gang S, Xi-cheng L, You-yong L, Genyu W and Wen-mei L, Soluble expression of active human β -defensin-3 in *Escherichia coli* and its effects on the growth of host cells, *Chin Med J*, 120, 8: 708-713, (2007).
41. Jack BC, Niels B, Isolation of neutrophil precursors from bone marrow for biochemical and transcriptional analysis, *Journal of Immunological Methods*, 232: 191–200, (1999).
42. Kathleen AD, Robert IL, Tomas G and Mitchell K, Isolation and characterization of human defensin cDNA clones, *Proc Natl Acad Sci*, 85: 7327-7331, (1998).
43. Hiromu T, Mitsuo K, Shigeru K, Hironori T, Yoshida R, Kazuo I, Ryusuke M, Hironobu K, Kohei H, Toshiaki U and Takayuki E, Cloning and expression of human defensin HNP-1 genomic DNA in *Escherichia coli*, *Journal of Microbiological Methods*, 25: 287-293, (1996).
44. Sylvia SLH, Alane SKP and Robert IL, Characterization of Defensin Precursors in Mature Human Neutrophils, *Blood*, 79: 1532-1537, (1992).
45. Josef JS, Angela U, Martin S, Monika SK, Hans Christian K, Human defensins, *J Mol Med*, 83: 587–595, (2005).
46. Michael ES, Sylvia SLH, Ganz T, James WS and Robert IL, Primary Structures of Three Human Neutrophil Defensins, *J. Clin Invest*, 76: 1436-1439, (1985).
47. Ganz T, Michael ES, Dorothy S, Sylvia SLH, Daher K, Dorothy FB and Robert IL, Defensins: Natural Peptide Antibiotics of Human Neutrophils, *J Clin Invest*, 76: 1427-1435, (1985).
48. Patricia MRA, Edward JH and John ALA, Copy number polymorphism and expression level variation of the human α -defensin genes DEFA1 and DEFA3, *Human Molecular Genetics* 14 (14): 2045–2052, (2005).
49. Xiongying C, Guozhen L, Gongyin Y, Haijiao W, Xiaojing S, Kechun W, Jiahua X, Illimar A, Screening and cloning of antimicrobial DNA sequences using a vital staining method, *Gene*, 430: 132–139, (2009).
50. Michael PP, Defensins and acne, *Molecular Immunology*, 40: 457–462, (2003).
51. Winnie D, Mona BE and Paul K, Intestinal defensin gene expression in human populations, *Molecular Immunology*, 40: 469–475, (2003).
52. Guozhang Z, Erik de L, Jacek L and Wuyuan L, Molecular Determinants for the Interaction of Human Neutrophil α Defensin 1 with its Propeptide, *J Mol Biol*, 381:1281–1291, (2008).
53. Marzena P, Jacek L, Expression and purification of recombinant human α defensins in *Escherichia coli*, *Protein Expression and Purification*, 49: 1–8, (2006).
54. Eric BM, Ann H, Nita S, John PR, Ralph JDB, Eduardo R and Charles LB, Human Enteric Defensins Gene Structure And Developmental Expression, *The Journal Of Biological Chemistry*, 271, 8: 4038–4045, (1996).
55. Michael Z, Antimicrobial peptides of multicellular Organisms, *Nature*, 415: 389-394, (2002).
56. Schaubert J and Richard LG, Antimicrobial peptides and the skin immune defense

- system, *J Allergy Clin Immuno* 3: 27, (2008).
57. Ganz T and Robert IL, Defensins, *Current Opinion in Immunology*, 6:584-589, (1994).
58. Peng L, Zhinan X, Xiangming F, Fang W, Peilin C, High-level expression of soluble human β defensin-2 in *Escherichia coli*, *Process Biochemistry*, 39: 2199–2205, (2004).
59. Zhinan X, Fang W, Peng L, Xiangming F and Peilin C, Expression of Human β -Defensin-2 With Multiple Joined Genes in *Escherichia coli*, *Applied Biochemistry and Biotechnology* 120: 1-13, (2005).
60. Oliver B, Petra R, Matthias M, Paul S, Christoph G, Sascha J, Georg T, Rainer P, Matthias L, Joachim GO and Ernst K, A novel horse α -defensin: gene transcription, recombinant expression and characterization of the structure and function, *Biochem. J*, 407: 267–276, (2007).
61. Yi L, Yufang M, Xu W, Jianguo L, Zhang C, Zhang K, Guoqing L, Ren L, The first antimicrobial peptide from sea amphibian, *Molecular Immunology*, 45: 678–681, (2008).
62. Patricia MS, Role of antimicrobial peptides in host defense against mycobacterial infections, *Peptides*, 29: 1836-1841, (2008).
63. Leitch GJ And Ceballos C, A Role of Antimicrobial Peptides in intestinal microsporidiosis, *Parasitology*, 136:175–181, (2009).
64. Robert L, Alan KL and Tomas G, Defensins: antimicrobial and cytotoxic peptides of mammalian cells, *Annual Review of Immunology*, 11:105-28, (1993).
65. Chen H, Zhinan X, Li P, Fang X, Yin X, Naizheng X and Peilin C, Recent advances in the research and development of human defensins, *Peptides*, 27: 931–940, (2006).
66. Chunyan H, Rui H, Fen D, Fang Z, Lei W, Junzhu W, LDL oxidation by THP-1 monocytes: Implication of HNP-1, SgIII and DMT-1, *Clinica Chimica Acta*, 402: 102–106, (2009).
67. Carl L, Maurice C, Jacopo V, Sofie K, Adam R, Bruce MC and Philippe B, Antimicrobial Activity Spectrum, cDNA Cloning, and mRNA Expression of a Newly Isolated Member of the Cecropin Family from the Mosquito Vector *Aedes aegypti*, *The Journal of Biological Chemistry*, 274 (29): 20092–20097, (1999).
68. Timothy JF, Nedra DK and Robert EWH, Mode of Action of the Antimicrobial Peptide Indolicidin, *The Journal Of Biological Chemistry*, 271 (32): 19298–19303, (1996).
69. Stephane C, Mohamed A, Jan M, Veronique V, Jean-Pierre LC, Pierre N and Antoine D, Structure, Synthesis, and Molecular Cloning of Dermaseptins B, a Family of Skin Peptide Antibiotics, *The Journal of Biological Chemistry*, 273 (24): 14690–14697, (1998).
70. Maria LM, Jose MS, Maria D, Donatella B, Maurizio S and Luis R, Temporins: Small Antimicrobial Peptides with Leishmanicidal Activity, *The Journal of Biological Chemistry*, 280 (2): 984–990, (2005).
71. Jeffrey WC, Kanniah R, Jesse M. J and Thomas EC, Transgenic expression of a gene encoding a synthetic antimicrobial peptide results in inhibition of fungal growth in vitro and in planta, *Plant Science*, 154: 171–181, (2000).
72. Mateja Z, Bostjan J, Iva HB and Roman J, Expression, purification and structural studies of a short antimicrobial peptide, *Biochimica et Biophysica Acta*, 1788: 314–323, (2009).
73. Bhargava A, Osusky M, Hancock RE, Benjamin SF, William WK and Misra S: Antiviral indolicidin variant peptides, Evaluation for broad-spectrum disease resistance in transgenic *Nicotiana*



- tabacum, *Plant Science*, 172: 515–523, (2007).
74. Amicis D, Barbara K, Margherita Z and Stefano M, Production of a recombinant antimicrobial peptide in transgenic plants using a modified VMA intein expression system, *FEBS Letters*, 519: 141-146, (2002).
75. Goddijn OJM and Pen J, Plants as bioreactors, *TIBTECH*, 13: 379-410, (1995).
76. Daniell H, Streatfield SJ and Wycoff K, Medicinal Molecular farming: production of antibodies, biopharmaceuticals and edible vaccines in plants, *Trends in Plant Science*, 6 (5): 219-226, (2001).
77. Larrick JW and Thomas DW, Producing proteins in transgenic plants and animals, *Current opinion in biotechnology*, 12: 411-418, (2001).
78. Carla M, Francesca KAB, Mark A, Paul BM and Brian FT, Antimicrobial peptides in animals and their role in host defences, *International Journal of Antimicrobial Agents*, 22: 465-478, (2003).
79. Scott B, Fabricio MB, Qiang C, Kevin VC and Chenming Z, Expression of Porcine Prorelaxin in Transgenic Tobacco, *Ann. N.Y. Acaemic Science*, 1041: 77–81, (2005).
80. Sergio HM and Gloria A, Antimicrobial peptides: A natural alternative to chemical antibiotics and a potential for applied biotechnology, *Electronic Journal of Biotechnology*, 6 (2): 271-284, (2003).