

**DOCOSAHEXAENOIC ACID REVERSES ALUMINIUM INDUCED ALTERATION
IN VITAMINS AND LIPID PROFILES IN RAT SERUM****DEVESH KUMAR JOSHI¹, MANISHA CHOUDHARY¹, SANDEEP TRIPATHI^{*1},
AMIT PAL² AND ABBAS ALI MAHDI³**¹*Department of Biotechnology, Nims Institute of Engineering & Technology, NIMS University, Jaipur-303121 India*²*Department of Biotechnology, Babasaheb Bhimrao Ambedkar University, Lucknow, India*³*Department of Biochemistry, King George's Medical University, Lucknow- India***ABSTRACT**

Aluminium (Al) is the third most widespread metal in the environment. It is known to be toxic for human and animals. We have studied the effect of simultaneous oral treatment of docosahexaenoic acid on aluminum induced changes in serum lipid profile and vitamins. A total of twenty four male albino rats were taken and divided in to four groups (N=6). First control group was treated with normal saline, second was treated with 100mg DHA, third was treated with 100mg AlCl₃ and fourth group was treated with both AlCl₃ and DHA. Aluminum treated rats showed a significant increase in serum low density lipoprotein (LDL), very low density lipoprotein (VLDL), triglyceride (TG), Cholesterol (TC) and lipid peroxide level (LPx) while decreased concentration of high density lipoprotein (HDL) and vitamins (A,C and E). DHA administration with Al showed reversal changes near to control in serum lipoprotein ie., LDL, VLDL, HDL, vitamins (A, C and E) and TG level. Moreover, DHA reduces the rate of lipid peroxidation. The results indicate that DHA has some beneficial effect in preventing Al induced toxicity.

KEYWORDS: Aluminum, Docosahexaenoic acid, lipid peroxidation, lipoproteins, vitamins**SANDEEP TRIPATHI**Department of Biotechnology, Nims Institute of Engineering & Technology, NIMS
University, Jaipur-303121 India**Corresponding author*

INTRODUCTION

Aluminum (Al) is a third most abundant element in the earth's crust followed by oxygen and silicon. It is used extensively in modern daily life. It is found in our food products, medicines and also added to drinking water for purification purposes¹. The high Al diet may lead to increase the deposition of Al in different tissues of the body especially in bones and brain^{2,3}. It is known to be a potential contributing factor in the aetiology of severe neurodegenerative disorders such as Alzheimer's disease (AD)^{4,5}. Al promotes the formation of amyloid- β protein and neurofibrillary tangles plaques by aggregating tau proteins in Alzheimer's disease⁶. Several studies reported that Al increases production of reacting oxygen species². The possible mechanism of Al induced toxicity has been related to cell damage via free radical production and increased lipid peroxidation (LPx) is one of the major consequences of oxidative stress². The primary targets of reactive oxygen species are cell-membrane polyunsaturated fatty acids, which, in turn, lead to damage in the cell structure and function. Some authors suggest that AL interacts with cell membrane directly and facilitate lipid degeneration⁷. There is an abundant evidence that Docosahexanoic acid (DHA) may have a protective role against oxidative burden by decreased lipid peroxidation rate⁸. Naturally, DHA is a major component of brain gray matter and of the retina in most mammalian species and is considered essential for normal neurological and cellular developments. DHA can also enter the blood by consuming food naturally containing DHA (eg: fish, fish oils, eggs, liver) or foods fortified with DHA-rich fish oils. Taking into consideration the above facts, the present study was conducted with an objective to explore the protective potential of DHA on AL induced biochemical deterioration in male albino rats and its correlate them with the time dependent severity of the disease process.

MATERIALS AND METHODS

Twenty four male *Rattus norvegicus*, Wistar strain rats (weight 220 ± 10 grams) were taken

from NIMS University animal house. The animals were separately housed in polypropylene cages in a room, which was maintained at a temperature of 22 ± 2 °C, relative humidity of 50 ± 10 % and 12h light dark cycles. They were fed a commercial pellet diet and allowed access to water ad libitum. The Institutional Animal Ethics Committee approved the study prior to the initiation of the experiment and also approved all experimental protocols.

Treatment

Animals were randomly divided into four groups (n = 6) viz. Group 1 served as control treated with normal saline, Group 2 treated with 100mg / kg body weight of DHA⁹, Group three treated with 100mg /kg body weight of $AlCl_3$ and Group four treated with 100mg $AlCl_3$ + 100mg DHA for 90 days.

Dose

Each solution was prepared with 1% gum acacia formed an aqueous suspension which was directly introduced into the rat pharynx via a feeding cannula (The sharp edged of the tip of a hypodermic needle no. 16 was blunted by grinding on a stone and thereafter bent to 120° so that the curved needle could easily be introduced into the rat pharynx via oral cavity without the pointed tip lacerating the passage) to experimental groups and an equivalent volume of physiological saline was given to control groups for 90 days.

Blood collection and serum separation

After 45 and 90 days of treatment the blood was collected from the retro-orbital plexus of the rats in plain vial for serum separation. The vial was kept for some time. Serum from blood after clotting separated out and collected in clean centrifuge tube and again centrifuged for 5-minutes at 3000 rpm. The serum thus obtained was used for further use.

Serum Lipid profiles

Serum total cholesterol, high-density lipoprotein, triacylglycerol were determined using Randox Laboratory kit reagents and

VLDL was calculated using the formula TG/2.2 mmol/l. Low density lipoprotein (LDL) cholesterol was determined by differential subtraction of the sum of the cholesterol fractions from the total cholesterol.

Vitamin estimations in serum

Vitamin A was estimated by the previously described procedure using high pressure liquid chromatography (HPLC) system (Waters Limited Mulford USA)¹⁰. The retinyl acetate was used as an internal standard and retinol levels were expressed as µg/dl.

Serum vit-C was measured according to method of Lowry et al¹¹ using 2,4 dinitrophenyl hydrazine in presence of thiourea as a mild reducing reagent to form a red coloured compound bis-hydrazone, which was measured at 520nm in spectrophotometer.

Serum vitamin-E was estimated after xylene extraction & reduction of ferric to ferrous ions, which then forms a red coloured complex with α-α' Dipyriddy¹². Absorbance was read at 460nm by spectrophotometer and a correction for the carotenes was made after adding ferric chloride and measured at 520nm.

RESULT AND DISCUSSION

In the present study, we evaluated AI induced time dependent changes in serum vitamins and lipid profiles in male albino rats. Protective potential of DHA was also tested in time course changes. The concentration of vitamins A was found to be significantly ($p < 0.05$) reduced in AI treated rats on the day 45 and 90 when compared with their respective controls (Fig-1). While, DHA treatment found to reverse these changes near to control on day 90 as compared with AI treated rats. Retinoic acid (RA), the active metabolite of vitamin A (retinoid), has also been shown to control the expression of genes related to Amyloid precursor protein (APP) processing in AD. Vitamin A is a hydrophobic antioxidant^{13,14}, which is decreased in diabetes causes retinopathy. It has also been reported that omega 3 fatty acid increases the concentration of vitamin A¹⁵. Our results are similar to that of Berson et al¹⁶. He also observed that omega 3 fatty acid restore the vitamin A and increase

the visual activity. Vitamin C concentration was diminished on the 45th and 90th days. Administration of AI caused a significant decrease in vitamin C levels in the serum (Fig-2). The results of the present study, the reduced concentration of vitamin particularly, Vit E and C may be responsible for the excessive generation of free radical in the brain. The reduced form of the vitamin, ascorbic acid, is an especially effective antioxidant owing to its high electron-donating power and ready conversion back to the active reduced form. There is great interest in the clinical roles of Vitamin C because of evidence that oxidative damage is a root cause of several neurodegenerative disorders. Population studies show that individuals with high intakes of Vitamin C have lower risk of a number of chronic diseases, including heart disease, cancer, eye diseases, and neurodegenerative conditions¹⁷. Vitamin E is the potent antioxidant. It is found to be reduced in both 45 and 90 days of treatment (Fig-3). Vit E (α-Tocopherol and its derivatives) is a predominant chain breaking lipid - soluble antioxidant and is believed to be the primary free radical scavenger and prevent lipid peroxidation¹⁸. Antioxidants are known to reduce oxidative radical induced reaction. α-tocopherol is an important antioxidant in biological systems. It inhibits peroxidation of membrane lipid by scavenging lipid peroxy radical with formation of tocopheroxyl radical as a consequence¹⁹. In addition, α-tocopherol maintained the levels of antioxidant membrane-bound enzyme and the activities of antioxidant enzymes near the normal value thus emphasizing their effect as antioxidant^{20,21}. Chinoy and Memon²² demonstrated that Vit E has a very significant bearing on the amelioration of AI toxicity in human.

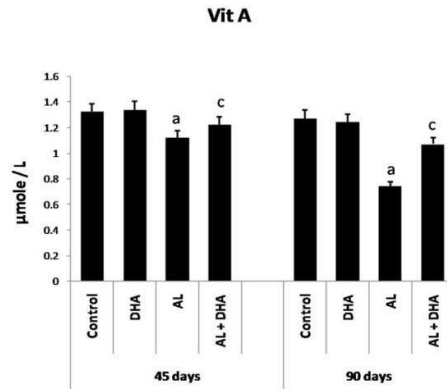


Figure.1

The concentration of vitamin A in serum of control and experimental group on 45 and 90 days. The results are expressed as Mean \pm SD six rat of each group (N=6). Superscripts relate significant ($p < 0.05$) comparison with Control Vs AL treated group (a) and AL treated Vs AL + DHA treated rats (c).

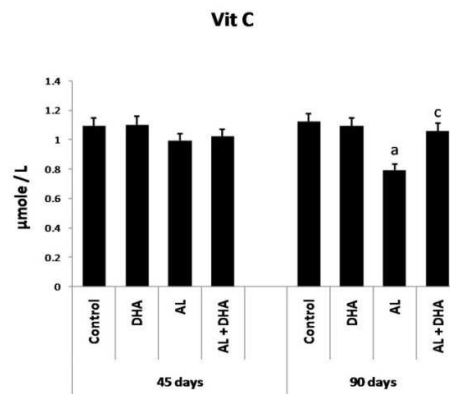


Figure.2

The concentration of vitamin C in serum of control and experimental group on 45 and 90 days. The results are expressed as Mean \pm SD six rat of each group (N=6). Superscripts relate significant ($p < 0.05$) comparison with Control Vs AL treated group (a) and AL treated Vs AL + DHA treated rats (c).

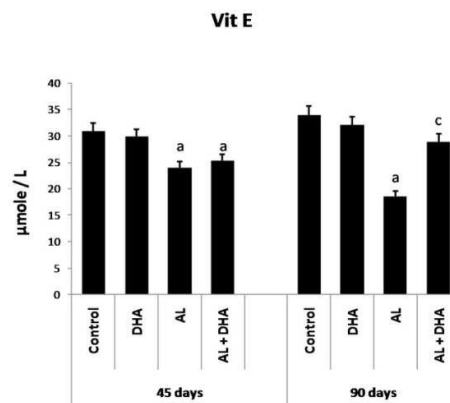


Figure.1

The concentration of vitamin E in serum of control and experimental group on 45 and 90 days. The results are expressed as Mean \pm SD six rat of each group (N=6). Superscripts relate significant ($p < 0.05$) comparison with Control Vs AL treated group (a) and AL treated Vs AL + DHA treated rats (c).

Table-1
Effect of Aluminum on lipid profiles in rats on 45 and 90 days of treatment

	45 days				90 days			
	Control	DHA	AL	AL + DHA	Control	DHA	AL	AL + DHA
TL (mg/ml)	360.5± 38.4	371.5 ± 43.4	276.7 ± 33.8 ^a	319.7 ± 31.3 ^c	405±27.5	419.5 ± 27.5	347.4 ± 33.2 ^a	394.6 ± 36.4 ^c
TC (mg/ml)	87.55 ± 9.7	91.5 ± 12.7	117.3 ± 23.9 ^a	101.2 ± 13.9 ^c	96.6 ± 12.8	101.3 ± 14.2	137.2 ± 32.7 ^a	127.6 ± 23.4 ^c
TG (mg/ml)	75.6 ± 2.5	69.6 ± 2.5	83.57 ± 2.2 ^a	73.1 ± 1.8 ^c	65.17 ± 7.2	68.34 ± 5.9	79.4 ± 12.7 ^a	74.7 ± 19.7 ^c
LPx (nmole/ml)	1.27±0.1	1.22 ± 0.1	1.98 ± 0.2 ^a	1.41 ± 0.2 ^c	1.34 ± 0.1	1.33 ± 0.2	2.57 ± 0.2 ^a	1.83 ± 0.2 ^c

Values are expressed as mean ± SD for six animals (N=6) in each group. The values of total lipid (TL), total cholesterol (TC), triglycerides (TG) and lipid peroxide levels (LPx) in control and experimental groups on 45 and 90 days. Sstatistical significance was determined by one way ANOVA followed by Neumann Keules post hoc test between groups. Superscripts relate significant ($p < 0.05$) comparison with AL treated vs control (a) and AL Vs AL + DHA treated rats (c).

Table-2
Effect of Aluminum on lipoproteins in rats on 45 and 90 days of treatment

	45 days				90 days			
	Control	DHA	AL	AL + DHA	Control	DHA	AL	AL + DHA
LDL (mg/dl)	46.6 ± 6.9	49.3 ± 6.8	62.9 ± 4.3 ^a	57.8 ± 3.3	43.6 ± 5.9	46.6 ± 6.8	102.9 ± 5.6 ^a	77.5 ± 6.6 ^c
VLDL (mg/dl)	27.4 ± 1.2	29.2± 1.3	36.1 ± 1.1 ^a	30.2 ± 1.2	22.4 ± 1.7	23.6 ± 1.7	42.4 ± 1.4 ^a	29.4 ± 1.7 ^c
HDL (mg/dl)	59.8 ± 4.3	57.3 ± 5.3	43.4 ± 2.1 ^a	53.2 ± 2.14 ^a	57.12 ± 1.5	55.3 ± 3.1	29.9 ± 2.7 ^a	41.5 ± 4.4 ^c

Values are expressed as mean ± SD for six animals (N=6) in each group. The values of low density lipoprotein (LDL), Very low density lipoprotein (VLDL) and high density lipoprotein (LDL) in control and experimental groups on 45 and 90 days. Sstatistical significance was determined by one way ANOVA followed by Neuman Keules post hoc test between groups. Superscripts relate significant ($p < 0.05$) comparison with AL treated vs control (a) and AL Vs AL + DHA treated rats (c).

We observed alterations in the lipid profiles following AL treatment (Table-1). The concentration of lipid was found to be significantly reduced in AI treated rats on both 45 and 90 days of treatment. The reduction of lipid may be correlate with the AI induced increased lipid peroxidation. Although biological mechanism of AI is not fully understood²³ it has been reported that AI promote the production of free radicals resulting in AI-Fe interdependence reaction²⁴. Total cholesterol and triglycerides were found to be elevated in AL treated rats when compared with the controls on the day 45 and 90. The increase in serum TG may be due to hypoactivity of lipoprotein lipase in blood vessels which breaks up TG. High serum cholesterol level may results from the hepatic dysfunction²⁵. Previously it has been reported that AI exposure accelerates neurodegeneration through lipid peroxidation via free radical production²⁶. In the present experiment, there was a significant increase in LPx after AI administration as we were found that significant ($p < 0.05$) elevated levels of MDA in the rat serum on the day 45 and 90. It is suggested that AI is responsible for oxidative burden. AL produces a subtle rearrangement in the membrane structure that facilitated the oxidative action of iron²⁷. Effects of AI on lipid peroxidation in various tissues such as liver, kidney, testis and brain of different animals were investigated and AI increased the rate of lipid peroxidation in some studies^{28,29,30,31}, while these changes were not observed in some studies^{23,32}. The concentrations of lipoproteins were found to be reduced significantly following AL exposure (Table-2). Increased levels of LPO or VLDL and LDL (β -lipoproteins) in treated rats are known to be cytotoxic to cells and tissues. Previously it is reported that during hyperlipemia, lipoproteins may initiate and maintain atheromatous lesions by endothelial cell injury and lipid accumulation³³. Recently it has been emphasized that hyperlipemic β -lipoproteins are cytotoxic to cells and tissues presumably due to enhanced levels of associated LPO³⁴. Lipids play significant role in various disorders like cardiovascular, hyperlipidemia fatty liver. It modifies the

composition, structure, and stability of cellular membranes. Excess lipid in the blood is considered to accelerate the development of arteriosclerosis. An altered lipid metabolism can alter the cardiac function by changing the properties of cardiac cell membrane and these changes may contribute to the cell death. High levels of circulating cholesterol and its accumulation in heart tissue are well associated with cardiovascular damage. Recent reports indicate that cytotoxicity induced by LDL and very low density lipoprotein can result from lipoprotein oxidation. Oxidation in these studies is indicated by the relative amount of MDA equivalents or thiobarbituric acid-reacting substances (TBARS) residing on the lipoprotein. The present data showed that the presence of DHA with AI reduces its harmful effect on all above measured biochemical parameters and their levels become near to the normal values of control. These results are in good accordance with other investigators^{35,36} who found that DHA maintained the levels of antioxidant membrane-bound enzyme and the activities of antioxidant enzymes near the normal value thus emphasizing their effect as antioxidant. Restoration of lipoproteins was found after the treatment of DHA. These results were agreed with the finding of Pan et al³⁷. He showed that the hypolipidemic actions of dietary PUFAs. A separate study shows that rats initially maintained on a diet deficient in fish oil have poor learning and memory skills. When they are transferred to a diet supplemented with DHA, however, they demonstrate dramatic improvement in learning and memory^{38,39}.

CONCLUSION

On the basis of result, it can be suggested that AL produces more free radicals and in turn this decreases the level of vitamins in the serum and deleterious effect in lipid profiles. Moreover, DHA ameliorate these changes. From these results, we could suggest that, DHA should be taken to compensate the deleterious effect of AL and potential risk for Alzhiemr's Disease.

REFERENCES

1. Nayak, P, Aluminum: impacts and disease. *Environ. Res*, 89, 111–115, (2002).
2. Tripathi S, Mahdi AA, Nawab et al. Influence of age on aluminium induced lipid peroxidation and neurolipofuscin in frontal cortex of rat brain: A behavioral, biochemical and ultrastructural study. *Brain Res*, 1253: 107-116, (2009).
3. Tripathi S, Mahdi AA, Hasan M et al. Protective potential of Bacopa monniera (Brahmi) extract on aluminum induced cerebellar toxicity and associated neuromuscular status in aged rats. *Cell Mol Biol*, 57: 3-15, (2011)
4. Drago, D., Bettella, M., Bolognin, S., Cendron, L., Scancar, J., Milacic, R., Ricchelli, F., Casini, A., Messori, L., Tognon, G., Zatta, P, Potential pathogenic role of beta-amyloid (1–42)-aluminum complex in Alzheimer's disease. *Int. J. Biochem. Cell Biol*, 40, 731–746, (2008).
5. Walton, J.R., Aluminum hippocampal neurons from humans with Alzheimer's disease. *Neurotoxicology*, 27, 385–394, (2006).
6. Exley, C., Esiri, M.M., Severe cerebral congophilic angiopathy coincident with increased brain aluminum in a resident of Camelford, Cornwall, UK. *J. Neurol. Neurosurg. Psychiatry*, 77, 877–879, (2006).
7. Fraga CG, Oteiza PI, Golub MS, Gershwin ME, Keen CL: Effects of aluminum on brain lipid peroxidation. *Toxicol Lett*, 51, 213-219, (1990).
8. Hiratsuka S, Ishihara K, Kitagawa T, Wada S, Yokogoshi H. Effect of dietary docosahexaenoic acid connecting phospholipids on the lipid peroxidation of the brain in mice. *J Nutr Sci Vitaminol (Tokyo)*, 54, (6):501-6, (2008).
9. Choudhary M, Joshi DK, Tripathi S, Kumar S, Mahdi AA. protective effect of docosahexaenoic acid on aluminium induced autonomic and gross behavioural changes in male albino rats *Int. J. Res. Pharm. Sci.* 2012, 2(3),131-145
10. Craft NE, Bulux J, Valdez C, Li Y, Solomons NW. Retinol concentrations in capillary dried blood spots from healthy volunteers: method validation. *Am J Clin Nutr*, 72(2): 450-454, (2000).
11. Lowry OH, Lopez JA, Bessey OA. The determination of ascorbic acid in small amounts of blood serum. *J Biol Chem*, 160: 609-615, (1945).
12. Baker H and Frank O. Vitamin status in metabolic upsets. *World Rev Nutr Diet*, 9: 124-160, (1968).
13. Dobrucki R, Radomska A. Retinol palmitate as a model substance to test antioxidant properties in vitro on the example of captopril. *Pharmazie*, 57(9):635–7, (2002).
14. Tesoriere L, D'Arpa D, Re R, Livrea MA. Antioxidant reactions of all-trans retinol in phospholipid bilayers: effect of oxygen partial pressure, radical fluxes, and retinol concentration. *Arch Biochem Biophys*, 343(1):13–8, (1997).
15. Kouchak A, Djalali M, Eshraghian M, Saedisomeolia A, Djazayeri A, Hajianfar H. The effect of Omega-3 fatty acids on serum paraoxonase activity, vitamins A, E, and C in type 2 diabetic patients. *J Res Med Sci.*, 16(7):878-84, (2011).
16. Berson EL, Rosner B, Sandberg MA, Weigel-DiFranco C, Willett WC. ω -3 intake and visual acuity in patients with retinitis pigmentosa receiving vitamin A. *Arch Ophthalmol*, 130(6):707-11, (2012).
17. Martin A, Youdim K, Szprengiel A, Shukitt-Hale B, Joseph J. Roles of vitamins E and C on neurodegenerative diseases and cognitive performance. *Nutr Rev.* 2002 Oct;60(10 Pt 1):308-26.
18. Cerolini, S.; Maldjian, A.; Surai, P. and Nobil, R. Viability, Susceptibility to peroxidation and fatty acid composition of boar semen during liquid storage. *Anim. Reprod. Sci*, 58 (1-2): 99-111, (2000).

19. Arita, M.; Sato, Y. Arai, H. and Inoue K. Binding of alphotocopherylquinone, an oxidized form of alpha-tocopherol, to glutathione S-transferase in the liver cytosol. *FEBS Lett*, 436:424–6, (1998).
20. Ithayarasi, A.P and Devi, C.S. Effect of alpha-tocopherol on lipid peroxidation in isoproterenol induced myocardial infarction in rats. *Indian J Physiol Pharmacol*; 41:369–76, (1997).
21. El-Demerdash F.M, Effects of selenium and mercury on the enzymatic activities and lipid peroxidation in brain, liver, and blood of rats. *J Environ. Sci. Health B*; 36: 489–99,(2001).
22. Chinoy, N.J. and Memon, M.R. Beneficial effects of some vitamins and calcium on fluoride and aluminium toxicity on gastrocnemius muscle and liver of male mice. *Fluoride*, 34:21–33, (2001).
23. Farina M, Lara FS, Brandao R, Jacques R, Rocha JBT: Effects of aluminum sulfate on erythropoiesis in rats. *Toxicol Lett*, 132: 131-139, (2002).
24. Tripathi S, Somashekar BS, Mahdi AA, Gupta A, Mahdi F, Hasan M, Roy R, Khetrpal CL. Aluminum mediated metabolic changes in rat serum and urine: A proton magnetic resonance study. *J Bio Mol Toxicol*, 22(2): 119-127, (2008).
25. Kojima K, Maki K, Tofani I, Kamitani Y, Kimura M. Effects of grape seed proanthocyanidins extract on rat mandibular condyle. *J Musculoskeletal Neuronal Interact*, 4(3):301-7, (2004).
26. Gupta, V.B., Suram, A., Hegde, M.I., Zecca, L., Garruto, R.M., Ravid, R., Shanker, S.K., Stein, R., Shanmngavelu, P., Rao, K.S.J., Aluminum in Alzheimer's disease: are we still at a crossroad? *Cell.Mol. Life Sci.* 62, 1–16, (2005).
27. Zatta P, Kiss T, Suwalsky M, Berthon G: Aluminium (III) as a promoter of cellular oxidation. *Coordin Chem Rev*, 228: 271-284, (2002).
28. Deloncle, R., Huguet, F., Fernandez, B., Quellard, N., Babin, P.H., Guillard, O., Ultrastructural study of rat hippocampus after chronic administration of aluminum l-glutamate: an acceleration of the aging process. *Experimental Gerontology*, 36, 231–244, (2001).
29. Chugh SN, Arora V, Sharma A, Chugh K: Free radical scavengers and lipid peroxidation in acute aluminium phosphide poisoning. *Indian J Med Res*,104: 190-193, (1996).
30. Guo CH, Huang CJ, Chiou YL, Hsu GSW:Alteration of trace element distribution and testis ACE activity in mice with high peritoneal aluminum. *Biol Trace Elem Res*,86: 145-157, (2002).
31. Julka D, Gill KD: Effects of aluminium on regional brain antioxidant defense status in wistar rats. *Res Exp Med*,196: 187-194, (1996).
32. Swain C, Chainy GBN: Effects of aluminium sulfate and citric acid ingestion on lipid peroxidation and on activities of superoxide dismutase and catalase in cerebral hemisphere and liver of developing young chicks. *Mol Cell Biochem*,187: 163-172. (1998).
33. Harker LA, Ross R, Slichter SJ, Scott CR Homocystine-induced arteriosclerosis. The role of endothelial cell injury and platelet response in its genesis. *J Clin Invest*, 58(3):731-41, (1976).
34. Zenker G, Költringer P, Boné G, Niederkorn K, Pfeiffer K, Jürgens G. Lipoprotein(a) as a strong indicator for cerebrovascular disease.*Stroke*,17(5):942-5,(1986).
35. Kusic B, Miric D, Zoric L, Ilic A, Dragojevic I. Antioxidant capacity of lenses with age-related cataract. *Oxid Med Cell Longev*, 467-130,(2012).
36. Andreo U, Elkind J, Blachford C, Cederbaum AI, Fisher EA. Role of superoxide radical anion in the mechanism of apoB100 degradation induced by DHA in hepatic cells. *FASEB J*, 25(10):3554-60,(2011).
37. Pan M, Cederbaum AI, Zhang YL, Ginsberg HN, Williams KJ, Fisher EA. Lipid peroxidation and oxidant stress regulate hepatic apolipoprotein B

- degradation and VLDL production. *J Clin Invest*, 113(9):1277-87, (2004).
38. Yurko-Mauro K, McCarthy D, Rom D, Nelson EB, Ryan AS, Blackwell A, Salem N Jr, Stedman M; MIDAS Investigators. Beneficial effects of docosahexaenoic acid on cognition in age-related cognitive decline. *Alzheimers Dement*,6:456-64, (2010).
 39. Johnson EJ, McDonald K, Caldarella SM, Chung HY, Troen AM, Snodderly DM. Cognitive findings of an exploratory trial of docosahexaenoic acid and lutein supplementation in older women. *Nutr Neurosci*, 1:75-83, (2008).