



RESEARCH ARTICLE

PHARMACOLOGY

**ANTIOXIDANT ACTIVITY OF PEDALIUM MUREX FRUITS IN CARBON TETRA
CHLORIDE- INDUCED HEPATOPATHY IN RATS***Corresponding Author***MADHU BABU A****Department of Pharmacology, Vikas college of Pharmacy, Jangaon,
Andhra Pradesh. India.***Co Authors***SRINIVAS P², VENKATESHWARULU L¹ AND ANIL KUMAR CH²**¹Department of Pharmacology, Vikas college of Pharmacy, Jangaon, Andhra Pradesh. India.^{2,1} Department of Biochemistry, Vikas College of Pharmacy, Jangaon, Andhra Pradesh, India.**ABSTRACT**

In this study antioxidant activity of methanol extract of fruits of *pedalium murex* (MEC) was investigated using carbon tetrachloride (CCl₄)- intoxicated rat liver as the experimental model. The hepatotoxic rats were administered MEC for 90 days (daily, orally at the dose of 70 mg per kg body weight). Lipid peroxidation (LPO) in CCl₄ - intoxicated rats was evidenced by a marked increment in the levels of thiobarbituric acid reactive substances (TBARS) and diene conjugates (CD), and also a distinct diminution in glutathione (GSH) content in the liver. In CCl₄ + MEC – treated rats these biochemical parameters attained an almost normal level. The decreased activity of antioxidant enzymes, such as superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPX) and glutathione reductase (GRD) in CCl₄ –intoxicatedrats, and its retrieval towards near normalcy in CCl₄ + MEC- administered rats revealed the efficacy of MEC in combating oxidative stress due to hepatic damage. Elevated level of glutathione transferase(GTS) observed in hepatotoxic rats too showed signs of returning towards normalcy in MEC co-administered animals, thus corroborating the antioxidant efficacy of MEC. The findings provide a rationale for further studies on isolation of active principles and its pharmacological evaluation.



KEY WORDS

Antioxidant enzymes, Carbon tetrachloride, *Pedalium murex* Linn, Lipid peroxidation.

INTRODUCTION

Pedalium murex L. (Pedaliaceae) is a diffuse, more or less succulent herb found near the sea coast of south India,¹. The fruits as well as the leaves and stems produced milk mucilage when agitated, and it is recommended as a treatment for gonorrhoea². An infusion or extract prepared from leaves is diuretic and demulcent, useful in treating disorders of the urinary system such as ardor urine, dysuria, spermatorrhoea, and incontinence of urine. As an emmenagogue, the juice is used in puerperal diseases and also to promote lochial discharge³. The mucilage from leaves and young shoots is used as an aphrodisiac in seminal debility⁴. The petroleum ether extract of *P. murex* is effective against Japanese encephalitis vector culex⁵. The aqueous extract of the whole plant has been found to possess analgesic and anti-inflammatory properties⁶. Extensive phytochemical investigations on the plant have revealed the presence of Pedalitin and Pedalin (major flavanoids) along with Diosmetin, Dinatin, Dinatin-7-glucuronide, Quercetin, Quercimeritin, and Quercetin-7-glucorhamnoside⁷. Triterpenoids such as α -amyrin acetate, Rubusic acid, ursolic acid, and lupeol acetate are reported⁸. Steroids such as β -sitosterol⁹, Sapogenins¹⁰ and Diosgenin¹¹ have also been reported. Lipids¹², phenolic acids such as caffeic acid, ferulic acid, protocatechic acid, and vanillic acid⁹, and amino acids such as aspartic acid, glutamic acid, and histidine are other phytoconstituents present in *P. murex*¹³. Although the plant contains several phytoconstituents, they have not been evaluated for their pharmacological activities in detail. Since no scientific data are available on the plant. Therefore in the present work attempt has been made to study the antioxidant effect of

fruits of *Pedalium murex* in CCl₄-intoxicated experimental rats.

It has been hypothesized that one of the principal causes of CCl₄ –induced liver injury is LPO by free radical derivatives of CCl₄. Thus the antioxidant activity or the inhibition of the generation of free radicals is important in the protection against CCl₄ - induced Hepatopathy¹⁴

Antioxidant action has been reported to play a crucial role in the hepatoprotective capacity of many plants, In Ayurveda, an indigenous system of medicine in India, has a long tradition of treating liver disorders, with plant drugs. Thus search for crude drugs of plant origin with antioxidant activity has become a central focus of study of hepatoprotection. This may prove effective in alleviating tissue damages prevalent in organisms as a consequence of exposure to toxins of extrinsic or intrinsic origin.

MATERIALS AND METHODS

Plant Material

Fruits of *Pedalium murex* were collected from Kolkonda Village, Warangal District of Andhra Pradesh. The materials were identified and authenticated by experts in Department of Botany, Kakatiya University. The collected materials were air dried at 35 – 40°C for a week and pulverized in electric grinder. The powder obtained was successively extracted in petroleum ether (60-80°C), benzene, chloroform and methanol by using Soxhlet extractor. The methanol extract was then made to powder with the help of rotary evaporator under reduced pressure. Fruits of *Pedalium murex* yielded 2.6% w/w of powdered methanol extract (MEC) which was stored in refrigerator



for further use. LD 50 of MEC was found to be 180 mg/kg body weight of animals.

Experimental Animals

Albino rats either sex, weighing 230-250 were obtained from the animal house of Lalitha College of pharmacy. The animals were divided into 3 groups eight animals each. The Study was conducted after clearance from the Animal ethical committee.

CCI4 – induced liver damage

Hepatopathy was induced in animals by subcutaneous (sc) administration of CCl₄ at lower Abdomen twice a week at the dose of 1 ml per kg. Body weight in double the volume of liquid paraffin (lp) which served as a vehicle. CCl₄ was administered on the first and fourth day of every week.

Experimental Procedure

Body weight of animals was recorded and then they were divided into 3 groups of 8 rats each. Group – I animals served as control, which received sc administration of lp only twice a week at the dose of 3ml per kg body weight of each animal. Group – II constituted the hepatotoxic group which received sc administration of CCl₄ + lp twice a week as mentioned elsewhere. Group III were the herb-treated ones which received sc administration of CCl₄ + lp twice a week as mentioned above. They also received MEC daily at the dose of 70 mg/kg body weight (effective dose) of each rat in a suspension of 1 ml water, orally by intubation. A pilot study revealed that MEC evoked hepatoprotection at doses ranging 40-120 mg/kg body weight of animals. Animals were maintained at laboratory conditions for a period of 90 days. Animals were fasted overnight on the

89th day. On the next day, after recording body weight, the animals were sacrificed by decapitation and blood was collected by the incision of jugular vein. The liver was dissected out, blotted off blood, rinsed in phosphate buffered saline (pH 7.4) and immediately proceeded for biochemical estimations. Serum was prepared from the collected blood.

Biochemical Estimations

The measurement of thiobarbituric acid reactive substances (TBARS) was done as an index of LPO.¹⁵ CD content was found out by the method of Klein¹⁶. Activities of SOD and CAT were determined by the methods of Marklund and Marklund,¹⁷ and Aebi.H,¹⁸ respectively. GSH content was determined after deproteinisation by the method of Beutler and Kelly,¹⁹. GPX was assayed by the method of²⁰. Glutathione transferase (GTS) and GRD were assayed by the methods of²¹, and²² respectively.

Statistical Analysis

The results were presented as the mean \pm SEM. Student's 't' test was used to analyse statistical Significance.

RESULTS

The concentration of TBARS and CD was significantly higher in liver of CCl₄- treated rats, as compared to normal control animals. (Table 1). These constituents were found to attain a near Conversely, GSH content in liver of Group – II animals showed a significant decline when compared with controls. But in Group III animals GSH content was found to attain near normalcy.

Table: 1
Effect of *Pedalium murex* fruit on the antioxidant status of liver in rats

Parameters	Group I	Group II	Group III
Thiobarbituric acid reactive substances			
TBARS (μ mol / mg protein)	0.81 \pm 0.05	1.08 \pm 0.04*	0.86 \pm 0.04**
Diene conjugates – CD (μ mol / 100 g tissue)	0.24 \pm 0.03	0.72 \pm 0.06*	0.28 \pm 0.05**
Reduced glutathione – GSH			
(μ mol / 100 g tissue)	378.6 \pm 17.6	206.4 \pm 10.8*	349.2 \pm 11.9**

Values are mean \pm SEM of 8 animals in each group.

* $P < 0.01$ as compared to Group I.

** $P < 0.01$ as compared to Group II.

Activities of antioxidant enzymes are presented in Table 2. The levels of SOD, CAT, GPX, and GRD recorded a significant decline in CCl₄- administered rats, when compared with normal controls. In CCl₄ +MEC- treated rats, the activities of these enzymes attained a near-normalcy. However, the activity of GTS was significantly higher in CCl₄ – treated animals, which was brought down towards normalcy in herb-treated rats.

Table: 2
Effect of *Pedalium murex* Linn on activity of antioxidant enzymes

Parameters	Group I	Group II	Group III
Superoxide dismutase - SOD (U / mg protein)	12.63 \pm 0.31	7.69 \pm 0.38*	11.93 \pm 0.59**
Catalase - CAT (U / mg protein)	8.38 \pm 0.39	4.86 \pm 0.32*	7.93 \pm 0.41**
Glutathione peroxidase – GPX (U / mg protein)	0.82 \pm 0.09	0.59 \pm 0.07 *	0.80 \pm 0.09**
- GRD (U / mg protein)	6.94 \pm 0.52	2.99 \pm 0.32*	5.99 \pm 0.44**
Glutathione transferase –GTS (μ mol / mg protein)	8.28 \pm 0.82	15.61 \pm 0.94*	8.19 \pm 0.61**

Values are mean \pm SEM of 8 animals in each group

* $p < 0.01$ as compared to Group I.

** $p < 0.01$ as compared to Group II.



DISCUSSION

Ample experimental and epidemiological studies support the involvement of oxidative stress in the pathogenesis and progression of several chronic diseases.²³ It is now known that oxygen, indispensable for maintaining life, sometimes becomes toxic and results in the generation of most aggressive agents such as reactive oxygen species (ROS). The high reactivity of ROS may trigger a host of disorders in body resulting in tissue damage and necrosis in many instances.²⁴ CCl₄ – mediated hepatotoxicity was taken here as the experimental model for liver injury. It has been established that CCl₄ is accumulated in hepatic parenchymal cells and metabolically activated by cytochrome P-450 dependent monooxygenases to form a trichloromethyl free radical (CCl₃•) which alkylates cellular proteins and other macromolecules with a simultaneous attack on polyunsaturated fatty acids in the presence of oxygen to produce lipid peroxides leading to liver damage.²⁵ A study using methanol extract of fruits of *pedalium murex* having doses ranging 40 - 120 mg/ kg body weight revealed the extract with dose 70 mg/ kg body weight offering the maximum hepatoprotection with respect to different liver marker enzymes, such as aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP) and gamma glutamyl transpeptidase (GGT). The body has an effective mechanism to prevent and neutralize the free radical – induced damage. This is accomplished by a set of endogenous antioxidant enzymes, such as SOD, CAT, GPX and GRD etc. When the balance between ROS production and antioxidant defenses is lost, 'oxidative stress' results, which through a series of events deregulates the cellular functions leading to various pathological conditions²⁶. Any compound, natural or synthetic, with antioxidant properties might contribute towards the partial or total alleviation of this type of damage.

In the present study, elevated level of TBARS and CD observed in CCl₄- treated rats indicates excessive formation of free radicals and activation of LPO system resulting in hepatic damage. TBARS produced as byproducts of LPO that occurs in hydrophobic core of bio-membranes.²⁷ The significant decline in the concentration of these constituents in the liver tissue of CCl₄ +MEC administered rats indicates anti-lipid peroxidative effect of *Pedalium murex*.

GSH is a major non-protein thiol in living organisms which plays a central role in coordinating the body's antioxidant defence processes. Perturbation of GSH status of a biological system has been reported to lead to serious consequences.²⁶ Decline in GSH content in the liver of CCl₄- intoxicated rats, and its subsequent return towards near-normalcy in CCl₄ +MEC- treated rats reveal antioxidant effect of *Pedalium murex* fruits. Explanations of the possible mechanism underlying the hepatoprotective properties of drugs include the prevention of GSH depletion and destruction of free radicals²⁸. These two factors are believed to attribute to the hepatoprotective properties of fruits of *pedalium murex*. SOD, CAT and GPX constitute a mutually supportive team of defence against ROS. SOD is a metalloprotein and is the first enzyme involved in the antioxidant defence by lowering the steady-state level of O₂⁻. CAT is a heme protein, localized in the peroxisomes or the microperoxisomes. This enzyme catalyses the decomposition of H₂O₂ to water and oxygen and thus protecting the cell from oxidative damage by H₂O₂ and OH⁻. GPX is a seleno-enzyme two third of which (in liver) is present in the cytosol and one third in the mitochondria. It catalyses the reaction of hydroperoxides with reduced glutathione to form glutathione disulphide (GSSG) and the reduction product of the hydroperoxide. In our study, decline in the activities of these enzymes in CCl₄-



administered rats revealed that LPO and oxidative stress elicited by CCl₄ – intoxication have been nullified due to the effect of fruits of *pedalium murex*. This observation perfectly agrees with those of ²⁹ who investigated hepatoprotective and antioxidant activity of *Bohemia nivea*.

GTS plays an essential role in liver by eliminating toxic compounds by conjugating them with glutathione. GRD is concerned with the maintenance of cellular level of GSH (especially in the reduced state) by effecting fast reduction of oxidized glutathione to reduced form. The activities of these enzymes were found to be in the reverse order. In liver tissues of CCl₄- administered rats, level of GTS registered a significant increment, whereas that of GRD recorded a decline. However, these enzymes restored an almost normal activity in CCl₄ + MEC – administered rats, thus unearthing the antioxidant effect of *pedalium murex*.

Natural antioxidants strengthen the endogenous antioxidant defences from ROS ravage and restore the optimal balance by neutralizing the reactive species. They are gaining immense importance by virtue of their critical role in disease prevention. In conclusion, it can be said that methanol extract of Fruits of *pedalium murex* exhibit a liver protective effect against CCl₄- induced hepatotoxicity and possessed anti-lipid peroxidative and antioxidant activities. Efforts are in progress here to isolate and purify the active principle involved in the hepatoprotective efficacy of this medicinal plant.

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REFERENCES

1. Nadkarni KM. Indian Materia Medica. Popular Prakashan, Bombay; 1982.
2. Mhaskar KS, Blatter E, Caiur JF. In: Kritikar and Basu's illustrated Indian Medicinal Plants, their usage in Ayurveda and Unani medicines, Vol-8, Indian Medicinal Science Series No 93,PID; New Delhi, 2000, pp. 2555-2559.
3. Chopra RN, Nayar SL, Chopra IC. Glossary of Indian Medicinal Plants, National Institute of cience Communication (CSIR), New Delhi; 1996.
4. Shukla VN, Khanuja SPS. Chemical, Pharmacological and Botanical studies on *Pedalium murex*. Journal of Medicinal and Aromatic Plant Sciences 26: 64-69, 2004.
5. Venkatarathina KT, Muthusamy VA, Ramanathan S et al. Evaluation of the petroleum ether extracts of *Pedalium murex* against Japanese encephalitis vector culex Tritaenlorhynchus. Antiseptic 102: 335-336, 2005.
6. Muralidharan P, Balamurugan G. Analgesic and anti-inflammatory activities of aqueous extract of *Pedalium murex* Linn. Biomedicine 28: 84-87, 2008.
7. Subramanian SS, Nair AGR. Flavonoids of the leaves of *Pedalium murex* Linn. Phytochemistry 11: 464-465, 1972.
8. Prasad TNV, Sastry KV. A note on the chemical examination of *Pedalium murex* Linn. leaves. Indian Drugs 25: 84-85, 1998.
9. ShuklaYN, Thakur RS. Heptatriacontan-4-one, Tetratriacontanyl octacosanoate and other constituents from *Pedalium murex* Linn. Phytochemistry 22: 973-974, 1983.
10. Harvey SK. A brief comparative pharmacognostic study of certain



- indigenous drugs. *Natural Medicinal Journal* 9: 519.
11. Mangle MS, Jolley CI. HPTLC studies on *Tribulus terrestris* (Chota ghokru) and *Pedaliium murex* (Bada ghokru). *Indian Drugs* 35: 189-194, 1998.
 12. Bhakuni RS, Shukla YN, Thakur RS. Flavonoids and other constituents from *Pedaliium murex* Linn. *Phytochemistry* 31:2917-2918, 1992.
 13. Rastogi JN, Sharma OD, Loiwai SD. Amino acids in certain medicinal plants. *Bull Pure Appl Sci* 1: 11-12, 1982.
 14. Castro, J.A., Ferrya, G.C, Castro, C.R., Sasame, H, Fenos, O.M. and Gillette, J.R. Prevention of Carbon tetrachloride-induced necrosis by inhibitors of drug metabolism. Further studies on the mechanism of their action. *Biochem. Pharmacol* 23, 295-302, (1974).
 16. Yagi, K. Lipid peroxides and Human disease. *Chem. Phys. Lipids*. 45, 337-351, (1987).
 17. Habig, W.H., Pabst, M.J. and Jakpoby, W.B. Glutathione transferase : A first enzymatic step in mercapturic acid formation. *J. Biol. Chem.* 249, 7130-7139, (1974).
 18. Aebi, H. Catalase In : *Methods in Enzymatic Analysis*, Ed Bergmeyer. H.U. Academic Press, New York, Vol 3, p. 276- 286, (1983).
 19. Beautler, E. and Kelley, B.M. The Effects of Sodium nitrate on red cell glutathione. *Experientia* 19, 96-97, (1963).
 20. Rotruck, J.T., Pope, A.L. and Gantter, H.E. Selenium: Biochemical roles as a component of glutathione peroxidase. *Sci.* 179, 588-590, (1973).
 21. Habig, W.H., Pabst, M.J. and Jakpoby, W.B. Glutathione transferase : A first enzymatic step in mercapturic acid formation. *J. Biol. Chem.* 249, 7130-7139, (1974).
 22. Racker, E Glutathione reductase (liver and yeast) In : *Methods in Enzymology*, Eds Colowick, S.P. and Kaplan, N.O. Academic Press, New York, Vol VII, p 722-725, .(1955)
 23. Sunita Tewari, Vani Gupta and Sandeep Bhattacharya. Comparative study of antioxidant potential of tea with and without additives. *Ind. J. Physiol. Pharmacol.* 44 (2), 215-219, (2000).
 24. Prasad Varier, S., Venkatachalam, S.R., Ramesh Chander. and Paul Thomas. Dietary antioxidantsnatural defence against disease. *Aryavaidyan* 12(3), 149-158, (1999).
 25. Recknagel, R.O. A new direction in the study of carbon tetrachloride hepatotoxicity. *Life Sci.* 33, 401-408, (1983).
 26. Uday Bandyopadhyay, Dipak Das. and Ranajit Banerjee, K. Reactive oxygen species: oxidative damage and pathogenesis. *Curr. Sci.* 77 (5), 658-665, (1999).
 27. Fraga, C., Leibovitz, B. and Tappel, A Halogenated compounds as inducers of lipid peroxidation in tissue slices. *Free Rad. Biol. Med.* 3, 119-123, (1987).
 28. Valenzuela, A., Lagos, C., Schmidt, K. and Videla, K, Silymarin protection against hepatic lipid peroxidation induced by acute ethanol intoxication in the rat. *Biochem. Pharmacol.* 3, 2209-2212, (1985).
 29. Chun-Chun Lin., Ming-Hong Yen, Tsae-Shiuan Lo. and Jer-Min Lin. Evaluation of the hepatoprotective and antioxidant activity of *Boehmeria nivea* var. *nivea* and *B. nivea* var, *tenacissima*. *J. Ethnopharmacol* 60, 9-17, (1998).